DIGITAL COMMONS @ UNIVERSITY OF SOUTH FLORIDA

University of South Florida Digital Commons @ University of South Florida

USF Tampa Graduate Theses and Dissertations

USF Graduate Theses and Dissertations

3-25-2014

Influence of Diet on Element Incorporation in the Shells of Two Bivalve Molluscs: Argopecten irradians concentricus and Mercenaria mercenaria

William Noland Elsaesser University of South Florida, william.elsaesser@cardno.com

Follow this and additional works at: https://digitalcommons.usf.edu/etd

Part of the Biogeochemistry Commons, and the Other Oceanography and Atmospheric Sciences and Meteorology Commons

Scholar Commons Citation

Elsaesser, William Noland, "Influence of Diet on Element Incorporation in the Shells of Two Bivalve Molluscs: Argopecten irradians concentricus and Mercenaria mercenaria" (2014). USF Tampa Graduate Theses and Dissertations.

https://digitalcommons.usf.edu/etd/5010

This Dissertation is brought to you for free and open access by the USF Graduate Theses and Dissertations at Digital Commons @ University of South Florida. It has been accepted for inclusion in USF Tampa Graduate Theses and Dissertations by an authorized administrator of Digital Commons @ University of South Florida. For more information, please contact digitalcommons@usf.edu.

Influence of Diet on Element Incorporation in the Shells of Two Bivalve Molluscs:

Argopecten irradians concentricus and Mercenaria mercenaria

by

William Noland Elsaesser

A dissertation submitted in partial fulfillment of the requirements for the degree of Doctor of Philosophy College of Marine Science University of South Florida

Co-Major Professor: Norman J Blake, Ph.D. Co-Major Professor: Pamela Hallock Muller, Ph.D. Sandra E Shumway, Ph.D. Peter Betzer, Ph.D. Karolyn Hansen, Ph.D.

> Date of Approval: March 25, 2014

Keywords: Bay scallop, northern quahog, shell chemistry, algal diet

Copyright © 2014, William Noland Elsaesser

Acknowledgments

Thank you to the Old Salt Fishermen's Society and the Sanibel-Captiva Shell Club for fellowships that partially funded this endeavor. Thank you to Bay Shellfish for allowing me to conduct the *Mercenaria mercenaria* spawns and initial rearing at the hatchery facility. Thank you to Ethan Goddard and Anthony Greco for their technical expertise and assistance with the ICP analyses and SEM x-ray analysis. Completion of this work would not have been possible without your contributions.

I would like to say thank you to my fellow lab members for assistance when schedules demanded my attention elsewhere and to Dr. Eric Monte for his assistance with the re-evaluation of statistical methodology. And finally, I would like to thank my committee for their patience and guidance throughout.

Table of Contents

	Tables	IV
List of	f Figures	xv
Abstra	act	xxii
1.	Introduction The bivalve mollusc shell Bivalve shell deposition Tissues involved Shell formation Bivalve shell mineralogy Minor and trace elements in bivalve shells Bivalve shells as environmental proxies Objectives	1 2 3 6 11 13 16 19
2.	Elemental composition in the shells of juvenile Argopectin irradians concentricus fed different diets: a first look Introduction Materials and Methods Algal cultures Specimen maintenance Feeding experiment SEM microanalysis ICP-OES Results Survivorship Growth Elemental analysis (ICP-OES) Left versus right valve Differences in elemental composition by diet received Discussion	21 23 23 25 26 27 27 27 27 28 30 30 35 36
1.	Introduction The bivalve mollusc shell Bivalve shell deposition Tissues involved Shell formation Bivalve shell mineralogy Minor and trace elements in bivalve shells Bivalve shells as environmental proxies Objectives Elemental composition in the shells of juvenile <i>Argopectin</i> <i>irradians concentricus</i> fed different diets: a first look Introduction Materials and Methods Algal cultures Specimen maintenance Feeding experiment SEM microanalysis ICP-OES Results Survivorship Growth Elemental analysis (ICP-OES) Left versus right valve Differences in elemental composition by diet received Discussion Conclusion	

3.	Elemental composition in the valves of juvenile Mercenaria			
	mercenaria fed unique algal diets: implications for ecological			
	patterns as well as for rearing bivalves for laboratory			
	experiments and aguaculture	45		
		45		
	Materials and Methods	47		
	Algae cultures	47		
	Spawning and rearing	48		
	Feeding experiment	49		
	ICP analysis	51		
	Statistical analysis	52		
	Results	53		
	Growth	53		
	Shell length and valve mass by diet	58		
	Differences between the left and right valves:			
	valve length and mass	66		
	Elemental shell chemistry	69		
	Statistical comparison of elemental shell chemistry			
	between left and right valves	83		
	Comparison of elemental shell chemistry by diet	86		
	Correlation analysis: time, growth, and element			
	association	135		
	Discussion	137		
	Shell length and mass	142		
	Left versus right valve elemental chemistry	145		
	Elemental shell composition differences as a			
	function of diet	147		
	Correlation analyses	157		
	Conclusion	159		
4.	Can phosphorous normalization indicate dietary			
	influence on elemental chemistry in bivalve shells?	161		
	Introduction	161		
	Methods	164		
	Results	164		
	Discussion	188		
	Conclusion	194		
5.	Temporal changes in the elemental shell chemistry of			
	Mercenaria mercenaria during starvation-induced stress			
	and death	195		
	Introduction	195		
	Methods	196		
	Results	197		
	Discussion	208		
	Conclusion	212		

ii

6.	Conclusions Recommendations for future work	214 221
Refer	rences	224
Appe	ndix A: Chapter 4 Post Hoc analysis results	248
Appe	ndix B: Chapter 5 Post Hoc analysis summaries	267

List of Tables

Table 1.1:	Microstructure groups and categories with associated varieties and constituent microstructures (after Carter, 1980).	14
Table 2.1:	Available lines, Limits of Quantitation (LOQ), and Minimum Detection Limits (MDL) for the USF Paleoclimatology, Paleoceanography Biogeochemistry Laboratory - Perkin- Elmer	
	4300 DV ICP-OES	28
Table 2.2:	Mann-Whitney Rank Sum Test– scallop left versus right valve pools by element	35
Table 2.3:	t-tests - scallop left versus right valve pools by element	36
Table 2.4:	Results for analysis of variance of scallop valve pool K/Ca by diet	38
Table 2.5:	Results of pairwise comparison (Holm-Sidak method) of scallop valve pool K/Ca by diet	38
Table 2.6:	Results for analysis of variance of scallop valve pool Mg/Ca by diet	39
Table 2.7:	Results of pairwise comparison (Holm-Sidak method) of scallop valve pool Mg/Ca by diet	40
Table 3.1A:	Median left valve length (mm) by diet and month collected	55
Table 3.1B:	Median right valve length by diet and month collected	55
Table 3.2:	Median mass (g) of the analyzed left valves by	
	diet and month collected	57
		iv

Table 3.3:	Median mass (g) of the right valves by diet and month collected	58
Table 3.4A:	Results for the Kruskal-Wallis analysis of shell length among diets	60
Table 3.4B:	Results for post hoc pair wise comparisons of shell length between diets	60
Table 3.5A:	Results of analyses of left valve mass by treatment	62
Table 3.5B:	Results of post hoc pairwise analyses of left valve mass and treatment	63
Table 3.6A:	Results of analysis of right valve mass comparison by treatment	65
Table 3.6B:	Results of post hoc pairwise comparisons of right valve shell mass by diet	65
Table 3.7A:	Results of analyses of all valves collected, left vs. right valve mass	67
Table 3.7B:	Summary results of analyses of baseline data versus experimental composite	68
Table 3.8A:	Summary of results for diet specific left and right valve comparisons	68
Table 3.8B:	Results of post hoc pairwise analyses of diet specific left and right valve comparisons	69
Table 3.9:	Results for left versus right valve comparisons with respect to Ni/Ca	84
Table 3.10:	Results for left versus right valve comparisons with respect to Zn/Ca	84
Table 3.11:	Significance of diet-specific left and right valve comparisons	85
Table 3.12:	Results of the comparison of the left and right valves from the control group with regard to B/Ca	87 v

Table 3.13:	Results of the comparison of the left and right valves from the control group with regard to Si/Ca	88
Table 3.14:	Results of the comparison of the left and right valves from the control group with regard to Zn/Ca	89
Table 3.15:	Results of the comparison of the left and right valves from the Cm group with regard to Ba/Ca	90
Table 3.16A:	Results for analysis of variance of left valve B/Ca between experimental diets	92
Table 3.16B:	Results for pairwise comparison of left valve B/Ca between experimental diets	92
Table 3.17A:	Results for analysis of variance of right valve B/Ca between experimental diets	93
Table 3.17B:	Results of pairwise comparison of right valve B/Ca between experimental diets	94
Table 3.18A:	Results of analysis of variance for left valve Ba/Ca among diets	96
Table 3.18B:	Results of pairwise comparison of left valve Ba/Ca between diets	96
Table 3.19A:	Results of analysis of variance of right valve Ba/Ca among diets	97
Table 3.19B:	Results of pairwise comparison of right valve Ba/Ca between diets	98
Table 3.20A:	Results of analysis of variance of left valve Co/Ca among diets	99
Table 3.20B:	Results of pairwise comparison of left valve Co/Ca between diets	100
Table 3.21A:	Results of analysis of variance of left valve Fe/Ca among diets	102
Table 3.21B:	Results of pairwise comparison of left valve Fe/Ca between diets	102 vi

Table 3.22A:	Results of analysis of variance of right valve Fe/Ca among diets	103
Table 3.22B:	Results of pairwise comparisons of right valve Fe/Ca between diets	104
Table 3.23A:	Results of analysis of variance of left valve K/Ca among diets	106
Table 3.23B:	Results of pairwise comparisons of left valve K/Ca between diets	106
Table 3.24A:	Results of analysis of variance of right valve K/Ca among diets	107
Table 3.24B:	Results of pairwise comparisons of right valve K/Ca between diets	108
Table 3.25A:	Results of analysis of variance of left valve Li/Ca among diets	110
Table 3.25B:	Results of pairwise comparisons of left valve Li/Ca between diets	110
Table 3.26A:	Results of analysis of variance of right valve Li/Ca among diets	111
Table 3.26B:	Results of pairwise comparisons of right valve Li/Ca between diets	112
Table 3.27A:	Results of analysis of variance of left valve Mg/Ca among diets	113
Table 3.27B:	Results of pairwise comparisons of left valve Mg/Ca between diets	114
Table 3.28A:	Results of analysis of variance of right valve Mg/Ca among diets	115
Table 3.28B:	Results of pairwise comparisons of right valve Mg/Ca between diets	115
Table 3.29A:	Results of analysis of variance of left valve Mn/Ca among diets	118 vii

Table 3.29B:	Results of pairwise comparisons of left valve Mn/Ca between diets	118
Table 3.30A:	Results of analysis of variance of right valve Mn/Ca among diets	119
Table 3.30B:	Results of pairwise comparisons of right valve Mn/Ca between diets	120
Table 3.31A:	Results of analysis of variance of left valve P/Ca among diets	122
Table 3.31B:	Results of pairwise comparisons of left valve P/Ca between diets	122
Table 3.32A:	Results of analysis of variance of right valve P/Ca among diets	123
Table 3.32B:	Results of pairwise comparisons of right valve P/Ca between diets	123
Table 3.33A:	Results of analysis of variance of left valve Si/Ca among diets	125
Table 3.33B:	Results of pairwise comparisons of left valve Si/Ca between diets	126
Table 3.34A:	Results of analysis of variance of right valve Si/Ca among diets	127
Table 3.34B:	Results of pairwise comparisons of right valve Si/Ca between diets	127
Table 3.35A:	Results of analysis of variance of left valve Sr/Ca among diets	129
Table 3.35B:	Results of pairwise comparisons of left valve Sr/Ca between diets	130
Table 3.36A:	Results of analysis of variance of right valve Sr/Ca among diets	131
Table 3.36B:	Results of pairwise comparisons of right valve Sr/Ca between diets	131 viii

Table 3.37A:	Results of analysis of variance of left valve Zn/Ca among diets	133
Table 3.37B:	Results of pairwise comparisons of left valve Zn/Ca between diets	133
Table 3.38A:	Results of analysis of variance of right valve Zn/Ca among diets	134
Table 3.38B:	Results of pairwise comparisons of right valve Zn/Ca between diets	135
Table 3.39A:	Pearson correlation analysis results of significance for element/calcium ratio associations with time, shell length, and valve mass	138
Table 3.39B:	Direction of association for initial Pearson correlations	138
Table 3.40:	Spearman's correlation analysis results of significance for element/calcium ratio associations with time, shell length, and valve mass	139
Table 3.41:	Best fit correlation function for data description	139
Table 3.42:	Significant element/calcium ratio correlation results – Pearson correlation	140
Table 3.43:	Significant element/calcium ratio correlation results- Spearman correlation	141
Table 4.1:	Results of Kruskal-Wallis analysis for (B/P)/Ca by diet	165
Table 4.2:	Results of Kruskal-Wallis analysis for (B/P) by diet	166
Table 4.3:	Results of Kruskal-Wallis analysis for (Ba/P)/Ca by diet	168
Table 4.4:	Results of Kruskal-Wallis analysis for (Ba/P) by diet	168 ix

Table 4.5:	Results of Kruskal-Wallis analysis for (Co/P)/Ca by diet	169
Table 4.6:	Results of Kruskal-Wallis analysis for (Co/P) by diet	170
Table 4.7:	Results of Kruskal-Wallis analysis for (Fe/P)/Ca by diet	172
Table 4.8:	Results of Kruskal-Wallis analysis for (Fe/P) by diet	173
Table 4.9:	Results of Kruskal-Wallis analysis for (K/P)/Ca by diet	174
Table 4.10:	Results of Kruskal-Wallis analysis for (K/P) by diet	175
Table 4.11:	Results of Kruskal-Wallis analysis for (Li/P)/Ca by diet	176
Table 4.12:	Results of Kruskal-Wallis analysis for (Li/P) by diet	177
Table 4.13:	Results of Kruskal-Wallis analysis for (Mg/P)/Ca by diet	178
Table 4.14:	Results of Kruskal-Wallis analysis for (Mg/P) by diet	179
Table 4.15:	Results of Kruskal-Wallis analysis for (Mn/P)/Ca by diet	180
Table 4.16:	Results of Kruskal-Wallis analysis for (Mn/P) by diet	181
Table 4.17:	Results of Kruskal-Wallis analysis for (Si/P)/Ca by diet	183
Table 4.18:	Results of Kruskal-Wallis analysis for (Si/P) by diet	184
Table 4.19:	Results of Kruskal-Wallis analysis for (Sr/P)/Ca by diet	185 x

Table 4.20:	Results of Kruskal-Wallis analysis for (Sr/P) by diet	186
Table 4.21:	Results of Kruskal-Wallis analysis for (Zn/P)/Ca by diet	187
Table 4.22:	Results of Kruskal-Wallis analysis for (Zn/P) by diet	188
Table 4.23:	Interpreted ranking of algae element quotas as depicted in Ho et al. (2003)	190
Table 5.1:	Summary of analysis of variance on ranks for B/Ca	198
Table 5.2:	Summary of analysis of variance on ranks for Ba/Ca	199
Table 5.3:	Summary of analysis of variance for Co/Ca	200
Table 5.4:	Summary of analysis of variance on ranks for Fe/Ca	201
Table 5.5:	Summary of analysis of variance on ranks for K/Ca	202
Table 5.6:	Summary of analysis of variance on ranks for Li/Ca	203
Table 5.7:	Summary of analysis of variance on ranks for Mg/Ca	204
Table 5.8:	Summary of analysis of variance on ranks for Mn/Ca	205
Table 5.9:	Summary of analysis of variance on ranks for Si/Ca	206
Table 5.10:	Summary of analysis of variance on ranks for Zn/Ca	207
Table A1:	Results summary of the post hoc analyses (Dunn's Method) for (B/P)/Ca differences between diets	2/18
		z40 xi

Table A2:	Results summary of the post hoc analyses (Dunn's Method) for (B/P) differences between diets	249
Table A3:	Results summary of the post hoc analyses (Dunn's Method) for (Ba/P)/Ca differences between diets	250
Table A4:	Results summary of the post hoc analyses (Dunn's Method) for (Ba/P) differences between diets	250
Table A5:	Results summary of the post hoc analyses (Dunn's Method) for (Co/P)/Ca differences between diets	251
Table A6:	Results summary of the post hoc analyses (Dunn's Method) for (Fe/P)/Ca differences between diets	252
Table A7:	Results summary of the post hoc analyses (Dunn's Method) for (Fe/P) differences between diets	253
Table A8:	Results summary of the post hoc analyses (Dunn's Method) for (K/P)/Ca differences between diets	254
Table A9:	Results summary of the post hoc analyses (Dunn's Method) for (K/P) differences between diets	255
Table A10:	Results summary of the post hoc analyses (Dunn's Method) for (Li/P)/Ca differences between diets	256
Table A11:	Results summary of the post hoc analyses (Dunn's Method) for (Li/P) differences between diets	256
Table A12:	Results summary of the post hoc analyses (Dunn's Method) for (Mg/P)/Ca differences between diets	257

Table A13:	Results summary of the post hoc analyses (Dunn's Method) for (Mg/P) differences between diets	258
Table A14:	Results summary of the post hoc analyses (Dunn's Method) for (Mn/P)/Ca differences between diets	259
Table A15:	Results summary of the post hoc analyses (Dunn's Method) for (Mn/P) differences between diets	260
Table A16:	Results summary of the post hoc analyses (Dunn's Method) for (Si/P)/Ca differences between diets	261
Table A17:	Results summary of the post hoc analyses (Dunn's Method) for (Si/P) differences between diets	262
Table A18:	Results summary of the post hoc analyses (Dunn's Method) for (Sr/P)/Ca differences between diets	262
Table A19:	Results summary of the post hoc analyses (Dunn's Method) for (Sr/P) differences between diets	263
Table A20:	Results summary of the post hoc analyses (Dunn's Method) for (Zn/P)/Ca differences between diets	264
Table A21:	Results summary of the post hoc analyses (Dunn's Method) for (Zn/P) differences between diets	265
Table B1:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for B/Ca	267
Table B2:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for Ba/Ca	267
Table B3:	Statistical analysis results summary – pairwise comparison procedure – for Co/Ca	268 xiii

Table B4:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for Fe/Ca	268
Table B5:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for K/Ca	268
Table B6:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for Li/Ca	269
Table B7:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for Mg/Ca	269
Table B8:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for Mn/Ca	270
Table B9:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for Si/Ca	270
Table B10:	Statistical analysis results summary – Dunn's pairwise comparison procedure – for Zn/Ca	270

List of Figures

Figure 1.1:	General molluscan bivalve shell features	4
Figure 1.2:	General molluscan bivalve shell structure relative to the extrapallial space and mantle epithelium (Jacobs et al., 2006)	5
Figure 1.3:	Nacre formations as described by Bevelander and Nakahara (1969), as interpreted in Jacob et al. (2008)	7
Figure 1.4:	Model of shell calcification from Levi-Kalisman et al. (2001)	8
Figure 1.5:	Model of shell calcification in both prismatic and nacreous layers (Nudelman et al., 2007)	12
Figure 2.1:	Percent survival of Argopecten irradians concentricus by diet received	29
Figure 2.2:	Growth rate (micrometers per day) of <i>Argopecten irradians concentricus</i> by diet received	29
Figure 2.3:	Si/Ca in pooled scallop shells by diet received	31
Figure 2.4:	Fe/Ca in pooled scallop shells by diet received	31
Figure 2.5:	K/Ca in pooled scallop shells by diet received	32
Figure 2.6:	Mg/Ca in pooled scallop shells by diet received	32
Figure 2.7:	Cu/Ca in pooled scallop shells by diet received	33
Figure 2.8:	Mn/Ca in pooled scallop shells by diet received	33
Figure 2.9:	P/Ca in pooled scallop shells by diet received	34

xv

Figure 2.10:	Cd/Ca in pooled scallop shells by diet received	34
Figure 2.11:	K/Ca versus diet received	37
Figure 2.12:	Mg/Ca versus diet received	39
Figure 3.1:	Shell length versus time by diet received	54
Figure 3.2:	Median mass of the analyzed left valves by diet and month collected	56
Figure 3.3:	Median mass of the right valves per diet and month collected	57
Figure 3.4A:	Median left valve length by treatment	59
Figure 3.4B:	Median right valve length by treatment	59
Figure 3.5:	Median left valve masses by treatment	62
Figure 3.6:	Median mass of the collected right valves by treatment	64
Figure 3.7:	Median left (L) and right (R) valve mass by diet received	67
Figure 3.8A:	Median B/Ca versus diet received for the collected left valves by collection month	70
Figure 3.8B:	Median B/Ca versus diet received for the collected right valves by collection month	71
Figure 3.9A:	Median Ba/Ca versus diet received for the collected left valves by collection month	71
Figure 3.9B:	Median Ba/Ca versus diet received for the collected right valves by collection month	72
Figure 3.10:	Median Co/Ca versus diet received for the collected left valves by collection month	72
Figure 3.11A:	Median Cu/Ca versus diet received for the collected left valves by collection month	73

Figure 3.11B:	Median Cu/Ca versus diet received for the collected right valves by collection month	73
Figure 3.12A:	Median Fe/Ca versus diet received for the collected left valves by collection month	74
Figure 3.12B:	Median Fe/Ca versus diet received for the collected right valves by collection month	74
Figure 3.13A:	Median K/Ca versus diet received for the collected left valves by collection month	75
Figure 3.13B:	Median K/Ca versus diet received for the collected right valves by collection month	75
Figure 3.14A:	Median Li/Ca versus diet received for the collected left valves by collection month	76
Figure 3.14B:	Median Li/Ca versus diet received for the collected right valves by collection month	76
Figure 3.15A:	Median Mg/Ca versus diet received for the collected left valves by collection month	77
Figure 3.15B:	Median Mg/Ca versus diet received for the collected right valves by collection month	77
Figure 3.16A:	Median Mn/Ca versus diet received for the collected left valves by collection month	78
Figure 3.16B:	Median Mn/Ca versus diet received for the collected right valves by collection month	78
Figure 3.17A:	Median P/Ca versus diet received for the collected left valves by collection month	79
Figure 3.17B:	Median P/Ca versus diet received for the collected right valves by collection month	79
Figure 3.18A:	Median Si/Ca versus diet received for the collected left valves by collection month	80
Figure 3.18B:	Median Si/Ca versus diet received for the collected right valves by collection month	80

Figure 3.19A:	Median Sr/Ca versus diet received for the collected left valves by collection month	81
Figure 3.19B:	Median Sr/Ca versus diet received for the collected right valves by collection month	81
Figure 3.20A:	Median Zn/Ca versus diet received for the collected left valves by collection month	82
Figure 3.20B:	Median Zn/Ca versus diet received for the collected right valves by collection month	82
Figure 3.21:	Left versus right valve for the Control feeding group with regard to B/Ca	87
Figure 3.22:	Left versus right valve for the Control feeding group with regard to Si/Ca	88
Figure 3.23:	Left versus right valve for the Control feeding group with regard to Zn/Ca	89
Figure 3.24:	Left versus right valve for the Cm feeding group with regard to Ba/Ca	90
Figure 3.25:	Left valve median B/Ca values compared to diet	91
Figure 3.26:	Left valve median Ba/Ca values compared to diet	95
Figure 3.27:	Left valve median Co/Ca values compared to diet	99
Figure 3.28:	Left valve median Fe/Ca values compared to diet	101
Figure 3.29:	Left valve median K/Ca values compared to diet	105
Figure 3.30:	Left valve median Li/Ca values compared to diet	109
Figure 3.31:	Left valve median Mg/Ca values compared to diet	113

Figure 3.32:	Left valve median Mn/Ca values compared to diet	117
Figure 3.33:	Left valve median P/Ca values compared to diet	121
Figure 3.34:	Left valve median Si/Ca values compared to diet	125
Figure 3.35:	Left valve median Sr/Ca values compared to diet	129
Figure 3.36:	Left valve median Zn/Ca values compared to diet	132
Figure 4.1:	Comparison of average elemental quotas among different algal taxa from Ho et al. (2003)	163
Figure 4.2:	Median (B/P)/Ca ratios versus diet received	165
Figure 4.3:	Median B/P ratios in <i>M. mercenaria</i> shells compared to diet	166
Figure 4.4:	Median (Ba/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	167
Figure 4.5:	Median Ba/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	168
Figure 4.6:	Median (Co/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	169
Figure 4.7:	Median Co/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	170
Figure 4.8:	Median (Fe/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	171
Figure 4.9:	Median Fe/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	172 xix

Figure 4.10:	Median (K/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	173
Figure 4.11:	Median K/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	174
Figure 4.12:	Median (Li/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	176
Figure 4.13:	Median Li/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	177
Figure 4.14:	Median (Mg/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	178
Figure 4.15:	Median Mg/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	179
Figure 4.16:	Median (Mn/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	180
Figure 4.17:	Median Mn/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	181
Figure 4.18:	Median (Si/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	182
Figure 4.19:	Median Si/P ratios with associated 95% confidence intervals In <i>Mercenaria</i> shells by diet	183
Figure 4.20:	Median (Sr/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	185

Figure 4.21:	Median Sr/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	186
Figure 4.22:	Median (Zn/P)/Ca ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	187
Figure 4.23:	Median Zn/P ratios with associated 95% confidence intervals in <i>Mercenaria</i> shells by diet	188
Figure 5.1:	Median B/Ca compared to month collected with associated 95% confidence intervals	198
Figure 5.2:	Median Ba/Ca compared to month collected with associated 95% confidence intervals	199
Figure 5.3:	Median Co/Ca compared to month collected with associated 95% confidence intervals	200
Figure 5.4:	Median Fe/Ca compared to month collected with associated 95% confidence intervals	201
Figure 5.5:	Median K/Ca compared to month collected with associated 95% confidence intervals	202
Figure 5.6:	Median Li/Ca compared to month collected with associated 95% confidence intervals	203
Figure 5.7:	Median Mg/Ca compared to month collected with associated 95% confidence intervals	204
Figure 5.8:	Median Mn/Ca compared to month collected with associated 95% confidence intervals	205
Figure 5.9:	Median Si/Ca compared to month collected with associated 95% confidence intervals	206
Figure 5.10:	Median Zn/Ca compared to to month collected with associated 95% confidence intervals	207

Abstract

Recently, biogenic carbonates have received much attention as potential proxies of environmental change; however, a major pathway of elemental incorporation is often overlooked when making interpretations or designing experiments. This research experimentally examines the influence of diet on elemental composition in juvenile shells of the bay scallop, *Argopecten irradians concentricus,* and the northern quahog, *Mercenaria mercenaria*.

Exploratory trials were conducted using *Argopecten irradians concentricus* juveniles fed different algal diets: *Isochrysis, Chaetoceros, Pavlova, Tetraselmis*, or a mix of all four in a 2:1:2:2 ratio. No differences between the left and right valves were revealed, thus, subsequent analysis of the dietary influence on shell chemistry utilized both valves. Only Mg/Ca and K/Ca were significantly different between the diet groups, though different influences were determined.

Experiments with juvenile *Mercenaria mercenaria* compared shell chemistries among clams fed unicellular diets of *Isochrysis* sp. (CCMP1324), *Pavlova pinguis* (CCMP609), *Chaetoceros mulleri* (CCMP1316), *Isochrysis sp.* (CCMP1611) culture, *Pavlova sp.* (CCMP1209), or *Chaetoceros galvestonensis* (CCMP186), a mixed diet of all species in equal ratios (Mixed), or no food (starvation control). The results indicate that diet can influence shell chemistry either directly or indirectly, with degree of influence varying by diet and mollusc species.

Additional information concerning the use of alternative element ratios and changes in the shell chemistry due to starvation-induced stress are also presented. Altogether, the present research provides valuable information concerning shell dynamics and potential diet-associated fluxes, thus demonstrating the need to consider the composition of dietary inputs when assessing environmental associations with elemental shell chemistries.

1. Introduction

The chemistry of biogenic carbonate shells is complex and influenced by a wide variety of processes. Consequently, many scientific applications are utilizing shell chemistries to interpret evolutionary, paleoenvironmental, environmental and chemical processes. To interpret trends in shell chemistries in response to environmental change, it is essential to understand how basic biological processes and environmental changes influence the elemental composition of a shell (Strasser et al., 2008).

Physical processes, especially the influence of temperature, are among the parameters most commonly considered as influencing shell chemistries, in part because rates of ion substitution can be strongly temperature dependent (Kennedy, 1969; Klein et al., 1996; Weiner and Dove, 2003; Schone, 2008; Strasser et al., 2008). Likewise, the chemistry of the water in which an organism was living and growing has been shown to influence shell chemistry (e.g., Panella and McClintock, 1968; Bryan, 1973; Carriker et al., 1980; Lorens and Bender, 1980; Rhoads and Lutz, 1980; Rainbow, 1993; Puten et al., 1996; Stecher et al., 1996; Leng and Pierce, 1999; Surge et al., 2003; Gilliken et al., 2006). Post-mortem processes (taphonomy) are also essential to consider, as are a variety of other processes, depending upon the organism considered, the environment in which it lives, and how it feeds.

Yet dietary contribution to elemental shell composition and variation of elemental constituents related to dietary algal composition are rarely if ever considered in studies and interpretations of shell chemistries. Thus, one of the primary mechanisms influencing the ratios of ions in the fluids from which a shell is precipitated is almost unknown.

This dissertation presents an examination of dietary influence on elemental concentrations in the shells of two commercially important species of bivalve. In Chapter 2, I examine the effects of the diet of young bay scallops, *Argopecten irradians concentricus,* on their shell chemistries. In Chapters 3-5, I report more extensive experiments that examined dietary and other associated influences on shell dynamics of *Mercenaria mercenaria.* To provide the background to understand how diet might influence shell chemistries, the basics of bivalve molluscan shell morphologies and shell deposition are reviewed in this introductory chapter.

The Bivalve Mollusc Shell

The outward and distinguishing feature of the bivalve mollusk is a shell made up of a pair of valves that provide protection and support for internal organs and processes (Carefoot and Donovan, 1995). Figure 1.1 depicts the general features of a marine bivalve molluscan shell.

The shell itself is an accretionary exoskeleton composed primarily of organics and calcium carbonate that are organized into layers. The outer-most layer is the organic periostracum, under which lies the mineralized portion of the shell, which usually is composed of aragonite, calcite or a combination of these minerals. The

extrapallial space separates the shell from the epithelial cells of the mantle. Figure 1.2 illustrates a partial section through a bivalve shell, showing its structure and relationship to the mantle epithelium.

The following sections provide information on the internal tissues associated with shell formation, the proposed mechanisms of shell formation, an overview of bivalve molluscan shell mineralogy, control of element incorporation, and a summary of previous findings related to elemental composition with regard to environmental and biologic influences.

Bivalve shell deposition

The process of shell deposition is often discussed at two different levels: 1) the major players and process associated with the transfer of necessary materials into the staging area for shell deposition, and 2) the formation of the shell and nucleation of the composite crystals. Each level will be discussed separately.

Tissues involved

Most sources refer to the tissues and spaces that are involved in shell deposition as "compartments", with the mantle (outer mantle epithelium), extrapallial space, and the inner shell surface identified as the primary compartments (Crenshaw, 1980; Wheeler, 1992). The secondary compartment most often identified is the blood sinus of the mantle tissue; this is the major vessel for either food-derived or medium- (seawater) - derived materials to be transported to the mantle epithelium (Crenshaw, 1980).

The outer mantle epithelium is associated with the transfer and deposition of materials due to its position adjacent to both the extrapallial fluid and the blood sinus, which is positioned between the two epithelial layers. The role of the mantle epithelium in shell deposition is believed to be limited mostly to the marginal edge, based on the



В.

Α.

Figure 1.1: General molluscan bivalve shell features. A. External shell features (Florida International University), B. Valve interior (Amqueddfa Cymru – National Museum Wales).

Pallial line



Figure 1.2: General molluscan bivalve shell structure relative to the extrapallial space and mantle epithelium (Jacob et al., 2008).

rate of shell deposition at the shell margin (Zischke et al., 1970; Wheeler and Wilbur, 1977) and upon high metabolic activity/respiratory rates described by studies such as Jodrey and Wilbur (1955). As such, the functional role of the outer mantle epithelium is further divided into two regions referred to as the central/proximal zone and the marginal zone; these zones are commonly illustrated in the histological classification of cell types and ultrastructure inherent to these cells (Crenshaw, 1980). The marginal zone, which is primarily associated with shell lengthening and thickening (Wheeler and Wilbur, 1977), is characterized by tall columnar cells with numerous mitochondria, well developed endoplasmic reticula and Golgi apparatus (Crenshaw, 1980). The central zone, which is more associated with acid-base metabolism and shell dissolution, is characterized by cuboidal epithelium with fewer mitochondria, less developed endoplasmic reticula, and minimal Golgi apparatus (Tsujii, 1968; Neff, 1972; Crenshaw, 1980). Transformation to columnar secretory cells can take place in the central zone to allow for shell repair (Tsujii, 1968).

The fluid-filled extrapallial space is the environment in which shell deposition takes place (Wilbur and Simkiss, 1972). The composition of the fluid is directly related to the metabolic activity of the outer mantle cavity, as hydrophobic barriers prevent direct association with the seawater (Crenshaw, 1980). As with the mantle, the pallial fluid can be categorized by the portion of the outer mantle with which it is associated. Though this is species specific, the outer mantle epithelium can attach at the pallial line to partition the pallial fluid into two unique components associated with the marginal and central zones (Crenshaw, 1980).

Shell formation

Shell formation is a complex process comprised of multiple stages and necessitating many biological products. Though the process has been studied for many years, there are still components that have not been accounted for and thus the exact mechanism is not fully understood.

The main components of the shell, and those consistently thought to be involved in the formation of the shell, are simply the organic matrix and calcium carbonate. Wilbur (1964) and Wilbur and Simkiss (1968) hypothesized that the organic matrix plays a significant role in shell formation as a depositional net that aids nucleation or that controls growth of the mineral. This idea led to different hypotheses related to shell formation. Bevelander and Nakahara (1969), for example, suggested that heterogeneous nucleation and growth occurs within established organic compartments (Figure 1.3). Studies such as Falini et al. (1996), however, suggest epitaxial growth of the mineral at active sites on the organic-matrix surface. Commonalities between the hypotheses are specific to the molecules suggested to be actively involved in biomineralization and include chitin, acidic glycoproteins and silk-like proteins (Bevelander and Nakahara, 1980; Levi-Kalisman et al., 2001), with chitin determining crystal orientation of aragonite (Blank et al., 2003) and the acidic proteins controlling nucleation, polymorph, texture and morphology of the crystals (Mann et al., 1989; Weiner and Addadi, 1997; Levi-Kalisman, 2001). Figure 1.4 depicts the model of shell calcification from Levi-Kalisman et al. (2001). These descriptions, however, are based on development of primarily nacreous layers, as this has been the focus of shell mineralization studies. The following discussion will provide a general description of shell formation based on recent studies and accounting for differences between shell layers and in mineralogy.







Figure 1.4: Model of shell calcification from Levi-Kalisman et al. (2001).

The depositional environment for mineral precipitation is isolated from the exterior environment (seawater) by the periostracum, which is the outermost layer of the bivalve shell comprised of highly cross-linked proteins, and the mantle epithelium (Addadi et al., 2006). This isolation allows for biological control of mineral formation (Simkiss and Wilbur, 1989; Weiner and Dove, 2003). This represents the first stage in shell formation.

The second stage is the secretion of necessary components into the extrapallial space and the construction of the organic matrix. The main components in the shell matrix are β -chitin, a hydrophobic silk protein, and other hydrophobic proteins, many of which are rich in aspartic acid (Addadi et al., 2006; Furuhashi et al., 2009). Cells of the outer mantle epithelium secrete a mixture of carbohydrates, proteins and lipids into the pallial space, all of which participate in the formation of the organic matrix (Lowenstam and Weiner, 1989; Gotliv et al., 2005). Addadi et al. (2006) suggest that the epithelial

cells secrete chitin first, which forms the general mold for the mineral, then assorted proteins, followed by silk protein in the form of a gel, which fills the chitin form and helps maintain spatial separation of lamellar sheets. In addition to the identified components, Meyer et al. (2007) identified specific phosphoproteins associated with secretory cells at the mantle edge, which they proposed to be significant in mineralization as phosphoprotein extracts have been shown to bind calcium-carbonate crystals while fixed in a hydrogel (Mount, 1999). The phosphoproteins, however, can also hinder crystal growth when in a solution (Wheeler et al., 1987).

Once the matrix (the form and foundation for mineralization) has been established, the mineral components are introduced. Vesicles containing amorphous calcium carbonate secreted from the mantle epithelial cells (Addadi et al., 2008), and/or hemocytes containing calcium carbonate crystals, deliver materials to the mineralization front (Mount et al., 2004). The presumed nacre-nucleation sites contain carboxylates and sulfates (Crenshaw and Ristedt, 1976; Nudelman et al., 2006), as seen in *Nautilus* shells. Similar sites have been seen in the shells of *Atrina rigida*, though less constricted to specific zones of the matrix (Nudelman et al., 2006). The acidic matrix components likely direct crystal formation through anionic domains (Sudo et al., 1997; Gotliv et al., 2005; Myers et al., 2007). Occluded material (acidic proteins) during mineralization is presumed to be responsible for different morphologies, as well as for alteration of solubility and mechanical characteristics (Addadi et al., 2006; Nudelman et al., 2007).

Prismatic layer formation is similar to that of nacre formation in origin and structure (Nakahara and Bevelander, 1970). The first stage is again the formation of an organic matrix juxtaposed to the periostracum and serving as a boundary for crystal

formation (Bevelander and Nakahara, 1969). These lamellae are then said to fragment and form envelopes within the compartments being formed. These envelopes contain dense extrapallial fluid (ground substance) enclosed during their formation. Crystal formation initiates within the envelopes at the inner margin with interprismatic material derived from the inner lamellae or ground substance (Nakahara and Bevelander, 1970). Nudelman et al. (2007) further describes the formation of prismatic layers atop an already formed mineral layer. In this process the first stage is suggested to be the deposition of a meshwork of chitin fibers. The next stage is the deposition of amorphous calcium carbonate onto the chitin fibers. The final stage is the crystallization of the calcium carbonate by epitaxial nucleation such that crystal orientation in each layer is maintained. Nudelman et al. (2007) further states that chitin fibers are occluded during mineralization. Figure 1.5 provides a schematic of the suggested mineralization mechanism for both prismatic layers and nacreous layers as described in Nudelman et al. (2007).

Biomineralization mechanisms remain speculative (Furuhashi et al., 2009). Lack of comparative research has prohibited a comprehensive view of molluscan shell formation (Furuhashi et al., 2009). While the structure of the bivalve shell has been generalized by many researchers, no attempt has been made to determine differences between even similarly constructed shells (Addadi et al., 2006). As illustrated in Kobayashi and Samata (2006), the components and molecular weight of the organic matrix are dependent upon the crystal structure and taxon of mollusc. Because the organic matrix has been shown to control mineralization, biological controls (genetic
controls) governing the organic matrix and thus crystal structure are an extremely important factor. It follows that not every molluscan shell would be the same.

Weiner and Traub (1980) suggested that chitin was not present in all molluscan shells, though since then, there has been a lack of research aimed at examining different taxa for chitin (Furuhashi et al., 2009). The presence of silk fibroin has not been verified in all molluscan shells (Ghiselin et al., 1967) and the protein structure lacks the helical and b-spirals characteristic of silk fibroin (Weiner and Traub, 1980). Absence of silk fibroin in prismatic layers was also suggested by Furuhashi et al. (2009).

The mechanisms for shell formation most likely differ in taxa adapted to different natural stresses (Crenshaw, 1980). Comprehensive research comparing a wide range of species and techniques will be necessary to determine all biological products associated with shell formation and the potentially numerous mechanisms of shell-layer mineralization based on evolution of the molluscan species (Crenshaw, 1980).

Bivalve shell mineralogy

The larval shell of a bivalve mollusc is constructed initially of amorphous calcium carbonate (Weiss et al., 2002), which is then transformed to aragonite. The larval shell of all bivalves is composed of aragonite with similar ultrastructure (Carriker and Palmer, 1979), implying that the larval shell is conserved through evolution (Taylor, 1973).





The shells at later life stages are also under the influence of evolutionary adaptations, and as such, the mechanisms for deposition are genetically and biological controlled. For example, taxonomic differences can be found in the calcium carbonate mineral precipitated, the direction of allometric growth, shape of the skeleton, thickness, hardness, coloration, and other shell factors (Rhoades and Lutz, 1980). All bivalve shells, however, are composites of calcium carbonate polymorphs (calcite, aragonite, or possibly vaterite), and an organic matrix as previously identified, but the mineral structure of the calcium carbonate and combinations thereof are species specific (Kobayashi and Samata, 2006).

Carter (1980) identifies seven microstructure groups and five microstructure categories in the Bivalvia (Table 1.1). The primitive molluscan shell is assumed to have been composed of simple aragonitic prisms and nacre, as found in the presumed ancestral Monoplacophora (Taylor, 1973); variations have occurred through time to produce the listed groups and categories. Possible advantages to evolutionary changes in mineralogy, architecture and microstructure include breakage resistance, abrasion resistance, resistance to shell dissolution, fracture localization and deflection, flexibility, lower density, economy of secretion, and variable rates of vertical shell growth (Carter, 1980).

Minor and trace elements in bivalve shells

Many minor and trace elements can be incorporated into the calcium carbonate shell of bivalve mollusks (Brookes and Rumsby, 1965; Kobayashi, 1975). These elements can be associated with pigments (Foxx, 1966), part of the structural components (Dodd, 1967), substituted for calcium or carbon, adsorbed, or associated

with physiological and environmental factors (Rosenburg, 1980).

Table 1.1: Microstructure groups and categories with associated varieties and constituent microstructures (after Carter, 1980).

Microstructure Groups and	Microstructure categories and
principal varieties	constituent microstructures
Prismatic	Aragonitic Prismatic
Simple prismatic	
Fibrous prismatic	
Spherulitic prismatic	
Composite prismatic	
Spherulitic	Calcitic prismatic
Laminar	Nacreous (aragonitic)
Nacreous	
Regularly foliated	
Crossed	Porcelaneous (aragonitic)
Crossed lamellar	Aragonitic crossed lamellar
Crossed acicular	Aragonitic crossed acicular
Complex crossed lamellar	Aragonitic complex crossed lamellar
Crossed-matted/lineated	Aragonitic crossed-matted/lineated
	Aragonitic homogenous
Microstructure Groups and	Microstructure categories and
principal varieties	constituent microstructures
Homogenous	Foliated (calcitic)
Homogenous s.s.	Regularly foliated
Granular	Calcitic crossed lamellar
	= crossed foliated
	Calcitic complex crossed lamellar
	= complex crossed foliated
Isolated Spicules or spikes	
Isolated crystal morphotypes	

Incorporation of the elements that make up the shell can be achieved in two ways. The inner mantle tissue can uptake the elements from the surrounding environment or the elements can be derived from the ingestion of particles (feeding) (Elinger, 1972). Both meet the same end; all elements are transferred to the blood sinus, are transported to the outer mantle epithelium and eventually into the pallial fluid for incorporation in the shell.

One major control of element incorporation is the chemical restriction associated with physical limitations of size for incorporation into the crystal lattice of the polymorphs of calcium carbonate. Aside from chemical restrictions, factors that determine the inclusion, distribution, or concentration of minor and trace elements range from ontogenetic regulation, physiological regulation, association with environmental dynamics, or can be seemingly random in nature (Rosenburg, 1980).

Magnesium can readily substitute for calcium in the crystal structure of calcite because of its relatively similar atomic radius, and has potential for environmental, ontogenetic, evolutionary and stoichiometric associations (Rosenberg, 1980). Strontium, in contrast, most readily substitutes for calcium in aragonite; Sr incorporation is widely studied as an indicator of temperature (Rosenburg 1980). The incorporation of both Mg and Sr can be influenced by many environmental factors. References to incorporation of other elements including manganese, boron, barium, lead, cadmium, iron, nickel, copper, zinc, as well as oxygen and carbon isotopes, are common in the scientific literature, with postulated influences including algal blooms altering media concentrations, temperature, salinity, ontogenetic variation, growth rate, tidal fluctuations, seasonal patterns, pollution, and sediment loading (e.g., Rucker and Valentine, 1961; Rosenburg, 1980; Carriker et al., 1996; Leng and Pearce, 1999; Putten et al., 2000; Lazareth et al., 2003). The studies, however, as noted by Rosenburg (1980), often assume cause and effect relationships without investigating all possible

factors in the elemental concentration and thus inconsistent results are common (Strasser et al., 2008).

Bivalve shells as environmental proxies

Ecological stresses can be recorded as changes in the chemistry of the molluscan bivalve shell. Their accretionary skeleton, sedentary nature, species-specific longevity, general hardiness, and preservation potential, make bivalves ideal candidates for studies of environmental influences on shell chemistry (Dodd, 1965; Phillips, 1977; Rhoads and Lutz, 1980). Presented here are some examples of experimental interpretations based on studies of shell-associated elements.

As noted earlier, Mg readily substitutes for Ca, especially in calcite, though studies have reported mixed results. Early researchers, including Chave (1954) and Wolfe et al. (1967), recognized the need for experiments to determine cause and effect relationships. Rosenburg (1980) demonstrated that Mg concentrations varied more in bivalves than other groups examined. Loren and Bender (1980) found possible confounding factors based on experimental conditions. Recent studies by Kleine et al. (1996) and Putten et al. (2000) have demonstrated that Mg concentrations covary with temperature, but Putten et al. (2000) also noted a sudden deviation indicating that the relationship was not constant. Other studies have correlated Mg with salinity (e.g., Milliman, 1974). Though observations relating Mg incorporation with environmental factors are inconclusive, evolutionary trends suggest biologically associated factors.

Strontium incorporation has been reported to be associated with multiple environmental factors though controversy surrounds these observations. Dodd (1965)

found that Sr increased with increasing temperature in the calcite layers in *Mytilus edulis*, and decreased with increasing temperature in the aragonitic shell layers. Thompson and Chow (1955), however, found no correlation with temperature in the multiple species they inspected. Hallam and Price (1968) surmised no correlation between growth rates and Sr, while Gilken et al. (2005) concluded that Sr was positively correlated with growth rate. Meanwhile, Hamer and Jenkins (2007) concluded that Sr was negatively correlated with growth rate. Stanton and Dodd (1970) concluded that Sr incorporation decreases with age, while Crisp (1975) found the opposite, and Strasser et al. (2008) reported mixed results.

Results with trace elements such as manganese have been equally problematic. Blanchard and Chasteen (1976) surmised the amount of substitution of Ca²⁺ by Mn²⁺ was correlated with tidal level, though the authors did not consider the oxygenation of the environment (Rosenburg, 1980). Crisp (1975) attempted to correlate the Mn concentration to salinity, while Strasser et al. (2008) correlated Mn/Ca levels with that of seawater concentration, potentially confounded by biological activity. Carre et al. (2006) suggested positive Mn/Ca association with growth rates while Strasser et al. (2006) showed negative correlation.

Multiple associations have been suggested for barium concentrations in shell, which is one of a very few elements potentially linked to algae dynamics thus far. Dorval (2007) suggested that Ba/Ca can be associated with salinity though more commonly, as in Stecher et al. (1996), Putten et al. (2000), Lazareth et al. (2007), and Thebault et al. (2009), is correlated with phytoplankton blooms (diatoms mostly). In the latter study, fresh water influences related to both phytoplankton blooms and increased Ba are

discussed as potential cofactors. Alternative environmental correlations were interpreted by Risk et al. (2010), who surmised a direct correlation of Ba in the shells of Chilean bivalves with local soil erosion. Coffey et al. (1997) also found possible correlation with fluvial inputs. Gillikin et al. (2006) linked dissolved concentrations of Ba with shell concentrations in *Mytilus edulis*.

Lead concentrations in shells are often correlated with the concentration in the natural environment, despite the complication of incorporation versus adsorption. Studies by both Ferrel et al. (1973) and Clarke et al. (1976) found a direct correlation between the lead concentration in the shell samples with that of the environment, though Clarke et al. examined the dead shells of Corbicula manillensis, while Ferrel et al. (1973) focused on the live shells of Crassostrea virginica. This presented the question of whether the measured lead was actually a component of the shell or simply adsorbed to the shell surface (Lutz and Rhoades, 1980). Other studies have discriminated between the dissolved and particulate phases of lead, including Pitts and Wallace (1994) who correlated lead in Mya arenaria shells with dissolved lead concentrations, while Borgoin (1990) associated the lead in Mytilus edulis shells with environmental concentrations of particulate Pb. More recently, Putten et al. (2000) found that their results were inconclusive and the lead concentrations in the shells of Mytilus edulis were not necessarily representative of the seasonal variability of environmental concentrations.

Objectives

The chemistry of the biogenic carbonate shell is clearly very complex (Wilbur, 1972). Carefully designed experimental studies are essential to understand the effects that environmental parameters and basic biological processes have on elemental components of the shell (Strasser et al., 2008). Moreover, all factors - such as diet – need to be included in the analyses to avoid misinterpretation and exclusion of primary contributors to shell chemistry.

This dissertation presents a first examination of the influence of algal diet on elemental concentrations in the shell of young bay scallops, *Argopecten irradians concentricus.* However, the major focus of my research is the detailed examination of dietary and other associated influences on shell chemistries of *Mercenaria mercenaria*. Similarly, few studies have determined if differences can occur between the two valves of a bivalve shell.

The elements analyzed in the shells of experimental *Argopecten* and *Mercenaria* include silver (Ag), aluminum (Al), arsenic (As), boron (B), beryllium (Be), calcium (Ca), cadmium (Cd), cobalt (Co), chromium (Cr), copper (Cu), iron (Fe), potassium (K), lithium (Li), magnesium (Mg), manganese (Mn), molybdenum (Mo), sodium (Na), nickel (Ni), phosphorous (P), lead (Pb), scandium (Sc), selenium (Se), silica (Si), tin (Sn), strontium (Sr), titanium (Ti), thallium (TI), vanadium (V) and zinc (Z). This suite of elements represents those available using the Perkin-Elmer 4300 DV ICP-OES in the USF Paleoclimatology, Paleoceanography Biogeochemistry Laboratory, but also includes the majority of elements frequently examined in calcium carbonate shells.

The following questions are addressed:

- Question 1. Do different algal diets contribute to unique elemental patterns in the bivalve shell?
- Question 2. Do diets based on different species of the same algal genus result in different elemental shell chemistries in bivalve molluscs? Furthermore, does origin of the algae affect associated elemental signatures?
- Question 3. Are elemental signatures distinct enough to differentiate the species of algae ingested? Is there a method which might increase the resolution between algal species ingested?
- Question 4. Do elemental signatures differ significantly between valves of a bivalve mollusc? If so, do differences suggest a genetic/evolutionary mechanism or possibly correlate with functionality?
- Question 5. Do elemental patterns contributed by diet to the bivalve molluscan shell depend on the mineralogy of the shell and species of bivalve?
- Question 6. Do the valves collected from dead bivalve molluscs (empty shell with no tissue) retain the elemental characteristics of the living population?

2. Elemental composition differences in the shell of juvenile *Argopecten irradians concentricus* fed different diets: a first look

Introduction

In general, the two pathways used for the incorporation of material into the pallial space [region of shell deposition (Wilbur and Simkiss, 1972)] are 1) direct internalization from the surrounding sea water or 2) feeding (Crenshaw, 1980). Though many studies concentrate on associations between the concentration of elements in seawater and the concentrations of those elements in the calcium carbonate shells of marine bivalves, little is known about the contribution of diet to elemental shell chemistry.

Ho et al. (2003) presented analysis of the cellular content of 15 phytoplankton species with regard to C, N, P, S, K, Mg, Ca, Sr, Fe, Mn, Zn, Cu, Co, Cd and Mo. In general, their study found that K concentrations are higher than seawater and Mg lower, except in the case of diatoms, where Mg and K are both relatively high. Ho et al. (2003) also repeated that the major nutrients C, N, P, S are variable with the average quotas of the organic biomass being similar to that of Redfield et al. (1963); and that the trace metals following the general pattern of Fe>Mn>Zn>Cu>Co=Cd>Mo. The coccolithophores examined by Ho et al. (2003) had higher Mn, Co, and Cd quotas compared to the diatoms; this was explained by the possible difference between metal requirements of oceanic (the coccolithophores) and neritic species (the diatoms). The

oceanic diatom examined, however, had low Fe, Mn, and Cu quotas and higher Co and Cd quotas compared to the coastal diatoms analyzed, thus lending support to the researcher's interpretation.

Besides the elemental content differences among algal taxonomic classes, researchers (e.g., Brown et al., 1997) have demonstrated varied nutritional value and biochemical composition of specific algae with regard to vitamin, amino acid, sugar, protein, and carbohydrate content, which may have implications for selective feeding upon certain algal classes. Bivalves such as the eastern oyster, *Crassostrea virginica*, select food particles based upon size, shape, concentration (Ward and Shumway, 2004) and biochemical composition, as demonstrated in the black lipped oyster (Brown et al., 1996).

Assimilation of material from ingested food particles is directly related to the food itself, i.e., size, biochemical composition, quantity and cell wall (Reinfelder and Fisher, 1991; Bayne, 1993). Furthermore, the partitioning of an element within an algal cell has been directly related to assimilation efficiencies of specific elements studied in *Mytilus edulis* shells and soft tissues (Wang and Fisher, 1996). Because some elements are more readily available for assimilation from certain algal cells than from others, selective feeding could potentially limit the elements internalized, providing for a mechanism of diet-associated elemental shell signatures related to the ecology of the organism and the specific cells being ingested.

The present research is a first look at the potential for variable elemental concentrations in the shell of *Argopecten irradians concentricus* due to differential diets provided in an aquacultural setting. The bay scallop, *Argopecten irradians concentricus*,

is a relatively fast growing hermaphroditic, filter-feeding bivalve with an average life span of 18 months. The shell consists of three layers: an inner and outer calcitic foliated layer, and a middle aragonitic crossed lamellar layer (Kennedy et al., 1969). Due to the short life span, the bay scallop is not considered an ideal experimental animal for longterm environmental association; however, the rapid growth rate and shell composed of both aragonite and calcite make the bay scallop a good specimen for exploratory research related to feeding dynamics.

Materials and Methods

Algal cultures

Six months prior to the start of the feeding trials, selected phytoplankton cultures were procured from Provasalli - Guillard Culture Center for Marine Phytoplankton [*Pavlova lutheri* (CCMP 1325)(Pl) and *Tetraselmis* sp. (CCMP 963)(Ttm)] or from Bay Shellfish Aquaculture [*Chaetoceros mulleri* (Cm) and *Isochrysis galbana* (TI)]. All cultures were acclimated to laboratory conditions for 48 hours prior to transfer to 150 ml sterile sea water for use as experimental stock cultures.

Stock and feeder cultures were maintained in a separate enclosure within the hatchery. Temperature was maintained at 22⁰ C. Both standard and full spectrum fluorescent lights were used for illumination.

Specimen rearing and maintenance

Six adult bay scallops, *Argopecten irradians concentricus,* were spawned using thermal induction. Three million fertilized eggs were collected and placed in a 500L tank

with a heat source and filtered air supply. The resultant larvae were held in the same tank for 48 hours before the first water change occurred.

Water was obtained from two 800 L settling tanks filled with water pumped from Tampa Bay and filtered using a sand filter, a diatomaceous earth filter, and two charcoal filters. The water pumped to the laboratory was further treated with an ultraviolet sterilizing unit and filtered a final time through a 1 μ m sock filter. The day prior to a water change, an alternate 500L tank was filled and fitted with a heat source and filtered air supply. The larval tanks were maintained at 26^oC and salinity of 30±2.

The water was changed every other day. The larvae were retained on a 35 µm sieve for the first two water changes and a 75 µm sieve for the remainder of the larval stage. The larvae were transferred to the alternate rearing tank immediately at which time they received the first of four daily feedings (a mixture of algal species composed of *Isochrysis galbana, Pavlova lutheri, and Chaetoceros mulleri* in a 2:2:1 ratio). After the animals were transferred, the drained tanks were cleaned using a dilute glacial acetic acid solution, rinsed with tap water, cleaned again with a mild Alconox solution, and rinsed with Reverse Osmosis water. The tank was allowed to dry for 24 hours before being filled with bay water.

Once 1% of the larvae were at pediveliger stage, five clumps of faux grass made by bundling strips of black plastic were added for settling substrate. The water change methodology altered such that only 50% of the water was replaced every other day. This procedure was followed until the post-set scallops were 450 µm in size (Lu and Blake, 1999).

Feeding experiment

Scallops attaining 450 µm (shell height) were randomly collected, divided into five groups of 150, then further divided into groups of fifty to provide three replicate experimental groups for each of the five treatments. Each group was placed into a 22L tank with a heat source and filtered air supply. The same temperature and salinity parameters used during the rearing process were used during the experiment.

Each experimental group of scallops was treated identically except for diet received. Each of four groups received only one of the following algal species for nutritional supplement: *Isochrysis galbana, Pavlova lutheri, Chaetoceros mulleri,* or *Tetraselmis sp.* The fifth group was fed the same mixed diet received during the larval phase (*Isochrysis galbana, Pavlova lutheri, and Chaetoceros mulleri* in a 2:2:1 ratio).

All algal cultures used for feeding experiments were allowed to grow until a concentration of $4x10^6$ cells ml⁻¹ was attained. Each treatment received a feeding of $1x10^5$ cells ml⁻¹ per day for the first two days. Subsequently, the daily ration was increased by 10,000 cells per milliliter per day every two days. Using these guidelines, a volumetric measure of algae to be fed daily was determined and divided into three or four doses spread equally throughout the day.

The feeding experiment was terminated after thirty days. All living animals were collected, the valves separated and the tissue excised by use of a stainless steel scalpel. Each shell was then rinsed in RO water and lightly brushed to remove foreign organic matter. Up to thirty left valves per feeding group were then placed in sealed glass vials for elemental analysis by Inductively Coupled Plasma Optical Emission

Spectroscopy (ICP-OES) and for inspection by Energy Dispersive X-ray Spectroscopy (EDS x-ray analysis).

SEM microanalysis

A Hitachi 2000[®] scanning electron microscope with a Sun System electron dispersion system attachment was utilized for the x-ray analysis. Five valves from each of the five feeding groups were used for the EDS portion of the research. The clean and dried valves were adhered to carbon stubs such that the umbo of each valve faced in the same direction and the inner shell surface came into contact with the carbon tape. This orientation helped to minimize shadowing affects as well as time needed for the analysis.

The SEM was set for variable pressure imaging and the vacuum set for 25 Pa. A magnification of 60X was used with a 12.3 mm working distance. No aperture was utilized and both 5 KV and 15 KV beams were used on each valve with an initial x-ray time of 200 seconds.

Each valve was inspected point by point in five different regions along the leading marginal edge and at a sixth point at the umbo. All x-ray times were adjusted depending upon count rate acquired to make sure all data collected were comparable. The results of this procedure were utilized to calibrate the ICP unit based on the general detection limits of the EDS system and peaks present in the analysis.

ICP-OES

Centrifuge tubes, 15 ml, were cleaned by soaking in hot 2% trace pure nitric acid. After approximately 15 minutes, the tubes were removed from the cleaning solution and dried. Caps were placed on the tubes and the outside of the tube rinsed with MilliQ water to remove any acid residue from the handling surface.

The remaining valves from each feeding group were pooled into one sample per treatment per replicate. Each grouping was rinsed one more time in MilliQ water and then ground before being placed in the clean 15 ml centrifuge tubes and digested in 4 mls of 4% trace pure nitric acid after the methodology of He and Mai (2001). The inductively coupled plasma optical emission spectroscopy unit was calibrated for the inspection of a full sweep of trace elements, set for a linear fit, and run with a blank of 4% trace-pure nitric acid between every sample. Table 2.1 provides the detection and quantitation limits of the ICP-OES equipment used.

The resulting concentration measures from the inductively couple plasma optical emission spectroscope analyses were corrected using the determined dilution factors (mass of solute/ {mass of solute + mass of solvent}). The data were further transformed into element: calcium ratios.

Results

Survivorship

The highest survivorship was found in juvenile scallops fed the mixed diet and the *Pavlova pinguis* diet (96% and 98% respectively), as compared to the *Pavlova lutheri* and *Isochrysis galbana* with 84% and 88% respectively (Figure 2.1). The lowest

survivorships observed were the Chaetoceros mulleri (52%) and the Tetraselmis sp. (10%) feeding groups.

element	λ(nm)	LOQ* (ppb)	MDL** (ppb)	element	λ(nm)	LOQ* (ppb)	MDL** (ppb)
·		06/25/04 Axial View	06/25/04 Axial View			06/25/04 Axial View	06/25/04 Axial View
Ag	328.068	1.57	0.47	Mg	280.271	0.68	0.2
AI	396.153	5.35	1.6	Mg	285.213	1.21	0.36
As	188.979	95.76	28.73	Mn	257.61	0.16	0.05
В	249.677	16.6	4.98	Мо	202.031	8.81	2.64
Ва	233.527	0.63	0.19	Na	589.592	3.53	1.06
Be	313.107	0.25	0.08	Ni	231.604	2.79	0.84
Са	315.887	19.56	5.87	Р	213.617	56.21	16.86
Са	317.933	4.44	1.33	Pb	220.353	13.08	3.92
Са	422.673	2.21	0.66	Sb	206.836	46.45	13.93
Cd	228.802	1.94	0.58	Sc	361.383	0.16	0.05
Co	228.616	1.57	0.47	Se	196.026	150.12	45.04
Cr	267.716	0.54	0.16	Si	251.611	8.51	2.55
Cu	327.393	1.75	0.52	Sn	189.927	-	-
Fe	238.204	1.17	0.35	Sr	407.771	0.03	0.01
Hg	253.652	31.98	9.59	Sr	421.552	0.04	0.01
К	766.49	1.29	0.39	Ti	334.94	0.26	0.08
Li	670.784	0.07	0.02	TI	190.801	54.62	16.39
Mg	279.077	4.2	1.26	V	290.88	0.81	0.24
Mg	279.553	0.38	0.11	Zn	206.2	6.59	1.98

Table 2.1: Available Lines, Limits of Quantitation (LOQ), and Minimum Detection Limits (MDL) for the USF Paleoclimatology, Paleoceanography Biogeochemistry Laboratory - Perkin-Elmer 4300 DV ICP-OES

* - Limit of Quantitation, 10s of reagent blank, n=20
** - Minimum Detection Limit, 3s of reagent blank, n=20

Growth

The highest growth rates were observed in scallops fed the mixed diet (control) and the Pavlova pinguis diet (Figure 2.2). On average, the specimens fed the mixed algal diet grew 110 µm per day while the scallops fed Pavlova pinguis grew on average 103 µm per day. The slowest growth rate observed was for scallops fed the Tetraselmis sp. at

52 μm per day. The growth rates on remaining diets were intermediate with the *Chaetoceros*-fed group growing on average 86 μm per day, *Pavlova lutheri*-fed scallops at 79 μm per day, and the *Isochrysis*-fed scallops 82 μm per day.



Figure 2.1: Percent survival of Argopecten irradians concentricus by diet received.



Figure 2.2: Growth rate (micrometers per day) of Argopecten irradians concentricus by diet received.

Elemental analysis (ICP-OES)

Complications during the analysis of the shell samples by Inductively Coupled Plasma Optical Emission Spectroscopy, greater than expected mortality in some of the feeding groups and variable valve mass restricted the available sample size to just two replicates. Figures 2.3 – 2.10 provide the element/calcium ratios for the scallop shell pools analyzed for the left and right valves, respectively, from feeding trials 1 and 3. Those elements originally included in the analysis but not reported were below the detection limits of the ICP-OES for all diets in both trials as were the values missing in the presented figures. The variability between the trials and valves is large for the majority of the element ratios examined. Differences in Fe/Ca (Figure 2.4), K/Ca (Figure 2.5), and Mg/Ca (Figure 2.6) are apparently influenced by diet. Due, impart, to the high variability, no trends are evident in the other elemental ratios amongst the diets.

Left versus right valve

The different valves were compared regardless of diet to provide a generalized comparison of the composition of the left and right valves of the bay scallop. All valve groups were analyzed by element/Ca ratio and the results are provided in Tables 2.2 and 2.3. The test used was based on the properties of the individual data sets. None of the elemental ratios were significantly different between the two valves.



Figure 2.3: Si/Ca in pooled scallop shells by diet received. 1) Mixed algal diet, 2) *Chaetoceros mulleri* diet (Cm), 3) *Isochrysis galbana* diet (Ti), 4) *Pavlova pinguis* (Pp), 5) *Pavlova lutheri* diet (Pl), 6) *Tetraselmis* diet (Ttm).



Figure 2.4: Fe/Ca in pooled scallop shells by diet received. 1) Mixed algal diet, 2) *Chaetoceros mulleri* diet (Cm), 3) *Isochrysis galbana* diet (Ti), 4) *Pavlova pinguis* (Pp), 5) *Pavlova lutheri* diet (Pl), 6) *Tetraselmis* diet (Ttm).



Figure 2.5: K/Ca in pooled scallop shells by diet received. 1) Mixed algal diet, 2) *Chaetoceros mulleri* diet (Cm), 3) *Isochrysis galbana* diet (Ti), 4) *Pavlova pinguis* (Pp), 5) *Pavlova lutheri* diet (Pl), 6) *Tetraselmis* diet (Ttm).



Figure 2.6: Mg/Ca in pooled scallop shells by diet received. 1) Mixed algal diet, 2) *Chaetoceros mulleri* diet (Cm), 3) *Isochrysis galbana* diet (Ti), 4) *Pavlova pinguis* (Pp), 5) *Pavlova lutheri* diet (Pl), 6) *Tetraselmis* diet (Ttm).



Figure 2.7: Cu/Ca in pooled scallop shells by diet received. 1) Mixed algal diet, 2) *Chaetoceros mulleri* diet (Cm), 3) *Isochrysis galbana* diet (Ti), 4) *Pavlova pinguis* (Pp), 5) *Pavlova lutheri* diet (Pl), 6) *Tetraselmis* diet (Ttm).



Figure 2.8: Mn/Ca in pooled scallop shells by diet received. 1) Mixed algal diet, 2) *Chaetoceros mulleri* diet (Cm), 3) *Isochrysis galbana* diet (Ti), 4) *Pavlova pinguis* (Pp), 5) *Pavlova lutheri* diet (Pl), 6) *Tetraselmis* diet (Ttm).



Figure 2.9: P/Ca in pooled scallop shells by diet received. 1) Mixed algal diet, 2) *Chaetoceros mulleri* diet (Cm), 3) *Isochrysis galbana* diet (Ti), 4) *Pavlova pinguis* (Pp), 5) *Pavlova lutheri* diet (Pl), 6) *Tetraselmis* diet (Ttm).



Figure 2.10: Cd/Ca in pooled scallop shells by diet received. 1) Mixed algal diet, 2) *Chaetoceros mulleri* diet (Cm), 3) *Isochrysis galbana* diet (Ti), 4) *Pavlova pinguis* (Pp), 5) *Pavlova lutheri* diet (PI), 6) *Tetraselmis* diet (Ttm).

Differences in elemental composition by diet received

Because the left and right valves were determined not to be significantly different in regard to elemental composition, both valve pools per trial were analyzed together to increase the overall sample size sufficiently for statistical comparison. Two elemental ratios proved to be significantly different between various experimental diets, K/Ca and Mg/Ca. The results of the ANOVAs and post hoc pairwise analyses are provided in Tables 2.4 - 2.7, and shown in Figures 2.11 and 2.12.

Group	N	Missing	Median	25%	75%			
Si/Ca-L	12	3	1.40E-05	9.92E-06	3.08E-05			
Si/Ca-R	12	3	1.34E-05	1.03E-05	2.65E-05			
		Mann-Whiti	ney U Statistic= 42.000					
		T = 84.000 n(sm	all)= 9 n(big)= 9 (P = 0.9	930)				
Fe/Ca-L	12	3	6.01E-05	4.72E-05	1.40E-04			
Fe/Ca-R	12	3	5.82E-05	5.03E-05	1.13E-04			
		Mann-Whiti	ney U Statistic= 39.000					
		T = 87.000 n(sm	all)= 9 n(big)= 9 (P = 0.9	930)				
Mg/Ca-L	12	3	8.61E-03	8.09E-03	8.99E-03			
Mg/Ca-R	12	3	8.81E-03	8.18E-03	9.27E-03			
Mann-Whitney U Statistic= 46.000								
T = 80.000 n(small) = 9 n(big) = 9 (P = 0.659)								
Cu/Ca-L	6	2	1.08E-03	7.14E-04	1.28E-03			
Cu/Ca-R	6	2	8.87E-04	7.01E-04	1.00E-01			
Mann-Whitney U Statistic= 7.000								
	T = 19.0	000 n(small)= 4 n(b	oig)= 4 P(est.)= 0.885 P(exact)= 0.886				
Mn/Ca-L	6	2	1.08E-03	8.04E-04	1.25E-03			
Mn/Ca-R	6	2	9.21E-04	7.98E-04	9.04E-02			
	Mann-Whitney U Statistic= 7.000							
T = 19.000 n(small)= 4 n(big)= 4 P(est.)= 0.885 P(exact)= 0.886								
Cd/Ca-L	6	2	9.56E-04	6.35E-04	1.14E-03			
Cd/Ca-R	6	2	7.82E-04	6.21E-04	8.84E-02			
		Mann-Whit	ney U Statistic= 7.000					
T = 19,000 n(small) = 4 n(big) = 4 P(est.) = 0.885 P(exact) = 0.886								

Table 2.2: Mann-Whitney Rank Sum Test- scallop left versus right valve pools by element

Group Name	N	Missing	Mean	Std Dev	SEM		
K/Ca-R	12	3	8.08E-04	2.93E-04	9.77E-05		
K/Ca-L	12	3	6.88E-04	2.33E-04	7.75E-05		
Difference			0.0001	2			
t = 0.962 with 16 degrees of freedom. (P = 0.350)							
95 percent confidence interval for difference of means: -0.000144 to 0.000384							
Group Name	N	N Missing Mean Std Dev SEM					
P/Ca-L	12	3	1.80E-03	3.04E-04	1.01E-04		
P/Ca-R	12 3 1.83E-03 2.28E-04 7.59						
Difference -0.0000226							
t = -0.178 with 16 degrees of freedom. (P = 0.861)							
95 percent confidence interval for difference of means: -0.000291 to 0.000246							

Table 2.3: t-test – scallop left versus right valve pools by element

The potassium ratio differed significantly between the *Pavlova pinguis* and Mixed diets, the *Pavlova pinguis* and *Pavlova lutheri* diets and the *Isochrysis galbana* and Mixed diets, establishing Pp > Mixed, Pp > PI, and Ti > Mixed (Table 2.5). The magnesium ratio differed significantly between the Mixed diet and all the analyzed single algal species diets establishing Mixed > Cm, Ti, Pp, and PI (Table 2.7).

Discussion

The Mixed algal diet produced the fastest growth rates and the second highest survivorship of juvenile bay scallops. The animals receiving the *Pavlova lutheri* and *Isochrysis galbana* both had similar survival rates as well as similar growth rates. The *Pavlova pinguis* and *Chaetoceros mulleri* diets did, however, deviate from the effect of the other single algal diets. *Pavlova pinguis* was associated with the highest survivorship and the second highest growth rate, thus was more similar to the mixed

diet. The *Chaetoceros mulleri* fed group had similar growth rates to *Pavlova lutheri* and *Isochrysis galbana*, but a low survivorship.

In general, diets composed of multiple algal species are superior to those composed of single algal species (Romberger and Epifanio, 1981; Albentosa et al., 1993; Brown et al., 1997) and the present study supports that trend. Diatoms, however, have been shown to be rich in fatty acids and support healthy growth rates and survival of cultured *Argopecten irradians* (Milke et al., 2006). In the present study, the initial size of the scallops used may have affected ingestion of the *C. mulleri* and thus their growth rate and survival.



Figure 2.11: K/Ca versus diet received. The mean values for each of the diets are provided with the standard deviations.

Group Name	N	Missing	Mean	Std Dev	SEM
Mixed	5	1	4.73E-04	1.13E-04	5.67E-05
Cm	5	2	7.25E-04	1.62E-04	9.38E-05
Ті	5	2	8.75E-04	1.16E-04	6.71E-05
Рр	5	1	1.07E-03	2.59E-04	1.30E-04
PI	5	1	6.18E-04	8.37E-05	4.19E-05
Source of Variation	DF	SS	MS	F	Р
Between Groups	4	8.45E-07	0.00000211	8.059	0.002
Residual	13	3.41E-07	2.62E-08		
Total	17	1.19E-06			

Table 2.4: Results for analysis of variance of scallop valve pool K/Ca by diet.

Table 2.5: Results of pairwise comparison (Holm-Sidak method) of scallop valve pool K/Ca by diet.

Overall significance level = 0.05							
Comparison	Diff of Means	t	Unadjusted P	Critical Level	Significant?		
Pp vs. Mixed	0.000601	5.248	0.000157	0.005	Yes		
Pp vs. Pl	0.000456	3.985	0.00155	0.006	Yes		
Ti vs. Mixed	0.000402	3.249	0.00634	0.006	Yes		
Pp vs. Cm	0.000349	2.82	0.0145	0.007	No		
Ti vs. Pl	0.000257	2.079	0.0579	0.009	No		
Cm vs. Mixed	0.000252	2.039	0.0623	0.01	No		
Pp vs. Ti	0.000199	1.61	0.131	0.013	No		
PI vs. Mixed	0.000145	1.263	0.229	0.017	No		
Ti vs. Cm	0.00015	1.132	0.278	0.025	No		
Cm vs. Pl	0.000108	0.87	0.4	0.05	No		

The results of elemental analysis of the scallop valves were highly variable. Each trial and valve pool produced inconsistent trends for most all of the elements examined. Though the random design of the experiment potentially increased the differences reported between the left and right valves, the observed variability within each treatment was larger than anticipated. This, however, was similar to previous results with cultured bivalves (Carriker, 1996; Strasser et al., 2008). This consistent problem with cultured specimens complicates analysis and evaluation of elemental trends in the shell, especially when attempting calibration-type experiments in the laboratory.



Figure 2.12: Mg/Ca versus diet received. The mean values for each of the diets are provided with the standard deviations.

Group Name	Ν	Missing	Mean	Std Dev	SEM
Mix	5	1	1.03E-02	3.37E-04	1.69E-04
Cm	5	2	8.65E-03	1.79E-04	1.03E-04
Ті	5	2	8.47E-03	5.95E-04	3.44E-04
Рр	5	1	8.02E-03	2.52E-04	1.26E-04
PI	5	1	8.50E-03	4.59E-04	2.30E-04
Source of Variation	DF	SS	MS	F	Р
Between Groups	4	0.0000119	0.00000297	19.892	<0.001
Residual	13	1.94E-06	0.000000149		
Total	17	0.0000138			

Table 2.6: Results for analysis of variance of scallop valve pool Mg/Ca by diet.

Overall significance level = 0.05							
Comparison	Diff of Means	t	Unadjusted P	Critical Level	Significant?		
Mix vs. Pp	0.00225	8.244	0.00000161	0.005	Yes		
Mix vs. Pl	0.00177	6.493	0.0000203	0.006	Yes		
Mix vs. Ti	0.00181	6.128	0.0000361	0.006	Yes		
Mix vs. Cm	0.00163	5.511	0.0001	0.007	Yes		
Cm vs. Pp	0.000626	2.122	0.0536	0.009	No		
Pl vs. Pp	0.000478	1.751	0.103	0.01	No		
Ti vs. Pp	0.000444	1.505	0.156	0.013	No		
Cm vs. Ti	0.000182	0.577	0.574	0.017	No		
Cm vs. Pl	0.000148	0.5	0.625	0.025	No		
Pl vs. Ti	0.0000344	0.117	0.909	0.05	No		

Table 2.7: Results of pairwise comparison (Holm-Sidak method) of scallop valve pool Mg/Ca by diet

The initial hypothesis was that elemental incorporation should be significantly different based on diet, following the findings of Ho et al. (2003). The Mn/Ca, Cd/Ca, and Co/Ca ratios were expected to differ between valves of scallops fed the the Coscinodiscophyceae (diatom) diet and the Prymnesiophycea (Ti, Pp, and Pl) diets, while Fe/Ca, Zn/Ca, and Cu/Ca were hypothesized to be most influenced by the *Tetraselmis* diet. The lack of data related to the *Tetraselmis* diet (high mortality restricting sample size and elemental concentrations below detection limits) prevented any examination of the elemental contribution to the shell. Evidence for influence of Mn/Ca, Cd/Ca/ and Co/Ca on the shells of the scallops fed *Chaetocerous*, *Pavlova* or *Isochrysis* was also inhibited by the inconsistency in the reults across trials and diets. The data sets related to the other elements were restricted in size by the lack of analytical results for trial 2.

To examine the overall differences in the elemental ratios between the left and right valves, the element/Ca ratio was examined regardless the diet received. Either a Mann-Whitney test or a standard T-Test was used to determine any difference between

the right and left valve relevant to the elemental ratios. All analyses indicated no significant differences in any of the elemental ratios compared to valve orientation. After probe analysis of the valves of the eastern oyster, Carriker et al. (1996) noted an increased concentration of most elements in the right valves examined and surmised the differences to be the result of structural differences, pigmentation, or mode of elemental incorporation (incorporation from bathing solution or transport to extrapallial fluid through tissues). The present research used element/Ca normalization, thus results are not fully comparable with the mentioned oyster study. Also, the left and right valves were selected randomly in the present study, meaning the left and right valves analyzed were not necessarily from the same scallops. Due to the combination of the different diet groups, the random selection of the valves, and pooled analysis groups; the examination of left and right valve differences is more a determinant of average experimental population dynamics related to elemental composition versus a direct comparison of left and right valves. As such, further experimentation is needed to formulate conclusive statements as to 1) right and left valve similarity, 2) whether differences noted in previous research compared to the present research are due to the comparison of an inequivalved bivalve and a more equivalved bivalve, or differences in analytical procedure, and 3) increased dissimilarity as a result of specific diets and metabolic effects.

Because the analyses of the left and right valves did not reveal any significant differences in the present study, both left and right valve pools were used in the analyses to determine differences among diets with respect to elemental shell composition. The increased sample size allowed for statistical analysis using a One

Way ANOVA and post hoc Holmes-Sidak method. Two elemental ratios were found to differ significantly between specific treatments: Mg/Ca and K/Ca.

Magnesium has been proposed to fluctuate with temperature in certain species of bivalve (Rucker and Valentine, 1961; Dodd, 1965; Rosenburg, 1980); though, more recent research suggests temperature and salinity have minor to no influence on Mg/Ca in other bivalve species (Carre et al., 2006; Strasser et al., 2008). In Crassostrea virginica (Carriker, 1996) and certain gastropods (Foster and Cravo, 2003), Mg has been shown to increase with size and through ontogeny. To eliminate two previously suggested influences on Mg/Ca, salinity and temperature were held constant during my experiments. The analyses revealed Mg/Ca for the Mixed diet was higher than all other diets tested. Ho et al. (2003) suggests that of the five taxonomic classes of algae examined in their study, diatoms had the largest Mg/P quota while the others were mostly similar. This trend was not evident in my study. The fact that the Mixed algal diet was the only diet to differ significantly from the other treatments further confounds the possibility that diet was directly responsible for the difference. As previously mentioned, Carriker (1996) suggested a relationship between size and Mg in the shell of Crassostrea virginica. Another study, Carre et al. (2006), has suggested a correlation between calcification rate and Mg/Ca. The present study does more closely support a correlation between Mg/Ca and growth rate or size. The Mg/Ca value for the Pp diet, however, is not consistent with this conclusion.

Carriker et al. (1996) concluded that differences in the potassium concentrations observed were possibly from ingestion of sedimentary particles containing potassium, though specific examination of the algal cells being ingested was not included. In the

present study, no sediments were added to any of the tanks as was done in Carriker et al. (1996) and water was treated in the same manner. The sole difference between experimental groups was the diet received, thus indicating an indirect biologic effect or a direct relationship with the algae ingested. Visually, it would appear that the Pp diet was correlated with the highest K/Ca; however, the only statistical differences were between the Pp and Mixed diets, the Pp and Pl diets, and the Ti and Mixed diets. The findings of Ho et al. (2003) suggested K is in higher concentration in algal cells than in seawater and that diatoms have a much higher K quota than the other algal classes examined. These finding are not fully translated to the shells of the bay scallops in the present study. There is no significant difference between any of the single algal taxonomic diets as related to K/Ca other than Pp vs. Pl. The finding that the mixed diet shells contain less K as compared to Ca than Pp or Ti diets cannot be explained by growth rates, but the lower K/Ca as compared to the other diets might be due to the decreased influence of any single algal species. The most notable difference in K/Ca between diets was the comparison of the Pp and PI diets, as this suggests a difference between algal species and not just algal classes though growth conditions and where the culture isolate was derived could play a role in these differences as identified in Ho et al. (2003).

Conclusion

Much research has been aimed at elucidating trends that apply to paleothermometry, environmental conditions, or ecological conditions. Most methodologies that have compared a laboratory setting with natural conditions, however, have failed to directly address diet as a factor in elemental enrichment. The results of the present research support the possibility that diet can contribute to the elemental composition of the shell of *Argopecten irradians concentricus*. Furthermore, algal species may contribute differently to shell composition. The actual mechanism, however, is uncertain and seemingly element specific. Clearly research is needed to distinguish biologic influences, including diet, to determine the dominant influences of elemental composition of shells and relationship with bivalve ecology and culture.

3. Elemental composition differences in the valves of juvenile *Mercenaria mercenaria* fed unique algal diets: implications for ecological patterns as well as for rearing bivalves for laboratory experiments and aquaculture

Introduction

The incorporation of elements into biominerals, including the shells of bivalves, is influenced by both geochemical and biological factors (Schone, 2008), but which elements and in what concentration are most often attributed to the physiochemistry of the surrounding water (Gillikin et al., 2006). Temperature or salinity are commonly assumed to influence shell chemistry, especially with respect to magnesium (Milliman, 1974; Putten et al., 2000), strontium (Dodd, 1965), manganese (Crisp, 1975), and barium (Dorval, 2007). Recently, however, researchers have begun to focus their attention towards mechanisms and pathways of incorporation of these elements. New research and experimental designs continue to elucidate those elements that are more likely influenced by environmental dynamics and those more biologically controlled, and thus, not dependable as environmental proxies (Stecher et al., 1996; Freitas et al., 2006).

In an assessment of element ratios between the digestive gland and gill tissues of *Laternula elliptica*, Poigner et al. (2012) suggested that assimilation of Al, Ca, Fe, K, Mn, and Mg is predominantly from the dissolved phase while elements such as Cd, Cu,

and Sr are assimilated through digestion of particulates. In another study, incorporation of Ba and Mo into the shells of *Pecten maximus* were examined with the conclusion that Ba enrichment was due to direct incorporation of dissolved Ba in proportion to the levels present in seawater while Mo enrichment was most likely associated with trophic uptake (Tabouret et al., 2012). Interestingly, the enrichment of Ba in scallop shells was delayed compared to maximum concentrations of the element in surrounding sea water which agreed with observations made by Stecher and Kogut (1999) and Ganeshram et al. (2003) that suggested dissolved Ba increased following specific algal blooms. Tabouret et al. (2012) postulated the increased concentration of dissolved Ba subsequent to algal blooms increased bioavailability which ultimately influenced incorporation into bivalve shells.

Based on the results of the previous chapter and conclusions of Carre et al. (2006), Strasser et al. (2006), Jenkins (2007), Strasser et al. (2008) and others, I postulated that the factors contributing to differences in elemental shell chemistry of *Argopecten irradians* fed unique diets was most likely associated with differences in growth rates caused by nutritional variation, time/ontogenetic factors, health/metabolic changes, or direct incorporation of elements associated with the diet received. I concluded that the difference in Mg/Ca was most evidently associated with growth rate, though confounded by other biological factors, while K/Ca was more directly influenced by the algal diet. The actual mechanisms, however, were uncertain and seemingly element specific.

The research presented in this chapter was designed to examine the influence of diet on the elemental shell chemistry of the hard shell clam, *Mercenaria mercenaria*.
This clam is a dioecious, filter-feeding, slower growing bivalve with an average life span of four to eight years (when considering fishing pressure/harvest), an estimated natural life span around 46 years (Peterson and Fegely, 1986), and suggested to have a potential maximum life span of at least 106 years (Ridgway et al., 2011). The shell of *M. mercenaria* is also composed of three distinct layers: an outer prismatic layer and a middle and inner homogenous layer (Panella and Mac Clintock, 1968). All three shell layers of the hard shell clam are aragonitic.

The results will be used to analyze differences in the elemental shell chemistry of clams fed diets of specific single algal species as well as clams fed a mixture of single algal species and starved individuals. The overall findings will be used to further determine the role diet plays in elemental shell chemistry of this marine bivalve. This work will aid in determining direct dietary influence on shell chemistry, species-specific effects and differences due to shell mineralogy, elemental composition change due to metabolic/health changes, growth-rate influence on shell chemistry, and the difference between left and right valves related to effects on the animal due to diet and associated biological factors.

Materials and Methods

Algae cultures

All species isolates were purchased from Provasalli - Guillard Culture Center for Marine Phytoplankton. *Isochrysis* sp. (CCMP1324), *Pavlova pinguis* (CCMP609), and *Chaetoceros mulleri* (CCMP1316), *Isochrysis sp.* (CCMP1611), *Pavlova sp.* (CCMP1209), and *Chaetoceros galvestonensis* (CCMP186) cultures were received several months prior to the initiation of the feeding trials to allow for proper acclimation periods, ensure that adequate starter and stock cultures were available, as well as to allow sufficient time to determine the health of all cultures.

All cultures were maintained within enclosures in the laboratory under the same conditions. Temperature was maintained at approximately 22⁰ C and both standard and full spectrum fluorescent lights were used for illumination. The stock cultures were maintained in three volumes including test tubes, 200 ml flasks (150 ml cultures), and 2 liter flasks (1500 ml culture) while feeding cultures were cultured in both 22 l K-Wall tubes and Carboys, with great care taken to ensure all feeding cultures were true monocultures.

Spawning and rearing

Thirty adult *Mercenaria mercenaria* were spawned using a flow through water table and thermal induction at the Bay Shellfish Aquaculture facilities in Palmetto, FL. The resultant fertilized eggs were transferred to a heated, aerated 800 I tank and left for 48 hours before the first water change and first feeding occurred. Similar to the scallop trials described in Chapter 2, an adjacent tank was prepared the day prior to ensure the temperature of the water and salinities were constant (28^oC and a salinity of 30+/- 2). After sieving the larvae, they were immediately placed into the other tank and received their first dose of food, which was a mix of *Chaetoceros* sp., *Isochrysis* sp., *and Pavlova sp.* The emptied tank was then cleaned using muriatic acid and Alconox, thoroughly rinsed, and filled with filtered, UV treated bay water. These changes occurred daily until metamorphosis of the clam larvae.

After metamorphosis, the settled clams were moved to a partial recirculating system (new water was always filtered and UV treated), divided equally between 12 downweller trays, and fed through continuous line systems with the same algal mixture received as larvae until they reached approximately 1-2 mm shell length. At that time, the clams were moved to a brown-water system supplied by natural water pulled from Tampa Bay until they reached an average of 4 mm shell length.

Feeding experiment

A subsample of the reared clams was sieved on a 4mm screen and all individuals retained were kept for the feeding trials and transferred to the Aquarium Laboratory at USF College of Marine Science. The clams were volumetrically counted and divided among 8 separate 500 L tanks and acclimated to laboratory conditions for 36 hrs. with no supplemental algae added.

Each of the tanks was fitted with a titanium heat source, two air lines, and a polyethylene mesh to suspend the clams from the bottom of the tank. Approximately 500 *M. mercenaria* were placed in each tank.

An approximate 25-50% water change occurred every other day and feedings (approximately 100,000-317,000 cells/ml per feeding) occurred three - four times daily with the overall volume fed dependent upon cell concentration of the individual cultures. If algae cultures were volume restricted for feeding on a specific day, the feeding volume was adjusted for all feeding groups such that equivalent cell concentrations were received. The feeding regimens were one of six single algal species diets - *Isochrysis* sp. (CCMP1324, TI), *Pavlova pinguis* (CCMP609, Pav609), *Chaetoceros*

49

mulleri (CCMP1316, Cm), *Isochrysis sp.* (CCMP1611, ISO) culture, *Pavlova sp.* (CCMP1209, Pav1209), or *Chaetoceros galvestonensis* (CCMP186, Cg), as well as a mixed diet of all species in equal ratios (Mixed), or no food (starvation control, CTRL).

The water used for exchanges was pulled from Bayboro Harbor and filtered through a sand filter, two charcoal filters 10 micron sock filter and a five micron sock filter into two holding tanks fitted with a heat source and air supply (the DE filter used during the scallop experiments was eliminated to limit Si sources). On a maintenance day, the water in the two tanks was cycled through a UV sterilization unit and a one micron sock filter prior to use in the laboratory.

Salinity and temperature measurements were taken from both the holding tanks and the experimental tanks so that adjustments in the volume of water exchanged could be made to maintain temperature and salinity in the experimental units (28⁰C+/- 1 and a salinity of 30+/-2). Each tank was siphoned to remove waste and the desired level of water. The tanks were then filled with the water passing again through the UV unit and another 1 micron sock filter. On non-water change days, small siphon tubes were used to eliminate as much accumulated waste as possible without significantly affecting water volume and introducing inter-tank differences beyond experimental variables. The first feeding occurred immediately following the water exchange or cleaning, with the remaining feedings spread as equally as possible through the day.

Once every two to three months, 25 individuals were randomly selected and removed from each of the tanks (January baseline collection, March, May, July, and October). All valves were immediately separated and the tissue excised. The valves

50

were then lightly brushed to remove debris, rinsed in distilled water, and dried before placing them individually in acid washed vials for storage until processed.

ICP analysis

The left and right valves were isolated from each of the individuals collected for processing during the first two collections. Each valve was individually measured and weighed on a micro scale, then quickly rinsed in a series of dishes – distilled water - 0.01% nitric acid – distilled water – milliQ water. After the shell was rinsed and dried, it was placed in an acid-cleaned vial and digested in approximately 4 ml of heated 5-6% trace-pure nitric acid. This procedure was repeated for all valves collected during the first two collections. The valves were then analyzed for silver (Ag), aluminum (Al), arsenic(As), barium (Ba), boron (B), beryllium (Be), calcium (Ca), cadmium (Cd), cobalt (Co), chromium (Cr), copper (Cu), iron (Fe), potassium (K), lithium (Li), magnesium (Mg), manganese (Mn), molybdenum (Mo), sodium (Na), nickel (Ni), phosphorous (P), lead (Pb), scandium (Sc), selenium (Se), silica (Si), tin (Sn), strontium (Sr), titanium (Ti), thallium (TI), vanadium (V) and zinc (Z) using the ICP-OES with blanks of 5% trace-pure nitric acid inserted between the valve samples. The detection limits of the ICP unit used are provided in Table 2.1.

The preparation procedure was slightly modified for the third and final collections. Rates of dissolution of the shells was noticeably influenced by size, as larger shells caused clogging of the ICP tubes, therefore, the left valves were placed in a muffle furnace at 400 degrees over night after they had been cleaned. The resultant ash was then reweighed and combined with the nitric acid in the vials for analysis. The right valves were immediately placed in air-tight containers following cleaning and drying for later experimental use.

Statistical analysis

The results from the ICP-OES analysis were corrected using the established dilution factor per sample. The resultant concentrations were then sorted and evaluated based on the Relative Standard Deviation values (RSD), detection limitations of the ICP equipment, and returns from the associated blanks. All concentrations associated with RSD values greater than 10 were removed from the final data set. A statistical analysis was performed to compare the final data set with the data set containing the removed values to evaluate bias. The two data sets were not significantly different (P = 0.87) as a whole; as well, the concentrations per element were evaluated individually and found not to be significantly different except for zinc (P > 0.05).

The concentrations remaining in the final data set were transformed to element/calcium ratios for comparison. The element: calcium ratios were initially analyzed using one-way analysis of variance (ANOVA). However, due to the limitations of the data (normalization and variance); a Kruskal Wallis one-way ANOVA on ranks was employed. Dependent on the outcome of the ANOVA, further testing was performed using a post hoc multiple comparison procedure to assess the significant differences in shell chemistry between feeding groups. This procedure was also followed to examine the differences between left and right valve chemistry followed by a T-test for each diet group. Further analysis was then employed using a Pearson Correlation and Regression Analysis to determine the association of the elemental

52

differences observed with diet as compared to valve length, valve mass, time, and elemental interactions.

Results

Growth

To minimize alterations in growth patterns due to handling and associated stress, shell-length measurement were only taken during the random sampling of the individuals for shell chemistry analysis. As such, the length and mass measurements portray the size distribution of the clams examined subsequent to each collection and not the absolute growth rate of the different feeding groups. All figures and tables subsequent to this description use median values to illustrate the trends discussed to remain consistent with statistical analyses performed and presented throughout.

The number of samples (n) does not represent all individuals originally collected; instead, the sample number is based on all individuals that were successfully analyzed during elemental analysis (ICP-OES) and used in subsequent statistical analyses reported herein. For example, the minimum shell length observed in the baseline collection is 6 mm though individuals of 4-5mm were initially collected and assessed. Furthermore, the largest shells collected for the Mixed diet group are also not represented in the summaries or analyses.

As the baseline (January) collection occurred prior to the initiation of the feeding trials, each of the diet-specific statistics represent the March – October collections. In addition, inclusion of right valve summary statistics only occurs for the baseline, March

53

and May collections due to experimental design and planned later use of the right valves collected during July and October.

An initial random subsample of clams was used to provide a baseline measurement with which all measurements taken during the experiment have been compared to determine changes over time related to the experimental variables. Thus, the changes in shell length and valve mass for each feeding group are compared to the same base line values. Figure 3.1 depicts the median shell length of each feeding group by collection month. As apparent from the figure, shell length per diet for each of the collections was variable and do not depict a consistent increasing trend for any of the treatments.



Figure 3.1: Shell length versus time by diet received.

Length	January	March	May	July	October
CTRL	7.50	7.50	6.75	7.00	7.50
ТІ	7.50	8.00	8.00	6.00	7.75
ISO	7.50	8.50	11.50	7.50	10.00
CG	7.50	6.50	8.00	8.00	11.00
СМ	7.50	9.00	8.50	8.25	12.00
Pav(609)	7.50	9.00	11.50	7.00	9.25
Pav(1209)	7.50	9.00	10.00	7.25	9.50
Mixed	7.50	7.50	10.00	9.75	12.00

Table 3.1A: Median left valve length (mm) by diet and month collected. The January collection period is equivalent to the baseline measurement with each subsequent month an experimental collection.

Table 3.1B: Median right valve length (n	nm) by diet and month collect	ed. The January collecti	on period is
equivalent to the baseline measurement	with each subsequent month	an experimental collecti	on.

Length	January	March	May
CTRL	7.50	7.50	6.75
ТІ	7.50	8.00	8.00
ISO	7.50	8.50	11.50
CG	7.50	6.50	8.00
СМ	7.50	9.00	8.50
Pav(609)	7.50	9.00	11.50
Pav(1209)	7.50	9.00	10.00
Mixed	7.50	7.50	10.00

The control group (CTRL) of starved clams (7.5 mm, 6.75 mm, 7.0 mm, and 7.5mm) did not exceed the baseline shell length (7.5mm) during the experiment. The TI fed clams exhibited a slight increase in shell length above the base line in the first collection (8.0 mm vs. 7.5 mm); however, none of the following collections exhibited a further increase – the specimens collected in May, in fact, had a median shell length below base line (6.0 mm vs. 7.5 mm). Only the Mixed diet (7.5 mm, 10 mm, 9.75 mm, and 12 mm) and CM diet (9.0 mm, 8.5 mm, 8.25 mm, and 12 mm) resulted in clams consistently above the base line shell length (7.5 mm). All remaining diet groups were variable in reference to baseline shell length, with at least one each whose median shell

length was equal to or below baseline. Table 3.1A provides the median shell length values/measurements by diet per collection month for comparison while Table 3.1B illustrates the similarity of the right value lengths for the first three collections.

The median mass of the left valves by diet over time are illustrated in Figure 3.2 and presented in Table 3.2. All diets with the exception of CG (2.41E-02 g) show an increased median mass over baseline (3.72E-02 g) during the March collection. The valve median mass of the control group is below baseline during the May collection. In the July collection, the median valve mass for the control and TI fed groups were below that of the baseline group. In the October collection, the control group was again the only experimental group whose median valve mass was below that of the baseline while general the other experimental groups valve mass followed the pattern Mixed>CM>CG>ISO>Pav1209>Pav609>TI>Baseline.



Figure 3.2: Median mass of the analyzed left valves by diet and month collected.

Mass	January	March	May	July	October
Ctrl	0.0372	0.0397	0.0270	0.0263	0.0341
ТІ	0.0372	0.0477	0.0411	0.0263	0.0539
ISO	0.0372	0.0540	0.1180	0.0419	0.0903
CG	0.0372	0.0241	0.0448	0.0479	0.1230
СМ	0.0372	0.0717	0.0455	0.0506	0.1650
Pav(609)	0.0372	0.0617	0.0891	0.0414	0.0666
Pav(1209)	0.0372	0.0688	0.0921	0.0400	0.0782
Mixed	0.0372	0.0435	0.0831	0.0744	0.1730

Table 3.2: Median mass (g) of the analyzed left valves by diet and month collected.

The mass of the right valves collected and analyzed during the first two collections post- baseline measurements follow similar trends to that of the left valves collected during the same months. The median mass of the right valves per diet and collection are depicted in Figure 3.3 and presented in Table 3.3.



Figure 3.3: Median mass of the right valves by diet and month collected.

Again, the median mass for each diet is above baseline except CG in the first collection. The only median mass below baseline during the second collection was the control group with the remaining groups showing the general ranking of ISO>Pav1209>Mixed>Pav609>CM>CG>TI>Baseline.

Mass	January	March	Мау
Ctrl	0.0372	0.0386	0.0205
ті	0.0372	0.0481	0.0409
ISO	0.0372	0.0524	0.1210
CG	0.0372	0.0254	0.0449
СМ	0.0372	0.0745	0.0454
Pav(609)	0.0372	0.0621	0.0878
Pav(1209)	0.0372	0.0720	0.0923
Mixed	0.0372	0.0440	0.0893

Table 3.3: Median mass (g) of the right valves by diet and month collected.

Shell length and valve mass by diet

The statistical analysis of the shell length measurements established that the collected valve composites for six of the experimental diets (Cm, Cg, Pav609, Pav1209, ISO, and Mixed) were significantly larger than the control group. The total collections for both the Baseline valves and those for the TI fed group were not significantly different from the control group's shell length. The shell length was not significantly different between any of the other diet comparisons, and importantly, no feeding group was significantly different from the baseline. There were, however, three comparisons which resulted in significant differences when all collected valves were included: Mixed vs. TI, Mixed vs. Baseline, and Cm vs. Baseline. Figures 3.4A and 3.4B illustrate the median shell length of the collected left and right valves, respectively, by treatment.

Tables 3.4A and 3.4B provide the summary of results for the statistical analyses performed.



Figure 3.4A: Median left valve length by treatment.



Figure 3.4B: Median right valve length by treatment.

Table 3 1 A. Regulte for the	Kruckal-Wallic anal	weie of chall lanath	among diate
	nuokai-wallio alla	yala ul anen lengun	amony ulets

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
Baseline	14	1	7.50	7.00	8.00
CTRL	61	1	7.00	6.50	7.50
ТІ	57	1	7.50	6.25	8.50
ISO	50	1	9.00	7.38	10.63
Cg	59	1	8.50	7.00	10.00
Cm	64	1	9.00	7.63	11.38
Pav(609)	59	1	8.50	7.00	10.00
Pav(1209)	54	1	9.00	7.00	9.50
Mixed	68	1	10.00	7.00	12.00
H = 75.9 with 8 degrees of freedom (P = <0.001)					

Table 3.4B: Results for post hoc pair wise comparisons of shell length between diets.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :					
Comparison	Diff of Ranks	Q	P<0.05		
Mixed vs. CTRL	167.61	6.84	Yes		
Cm vs. CTRL	164.79	6.63	Yes		
ISO vs. CTRL	132.33	4.99	Yes		
Cg vs. CTRL	112.51	4.43	Yes		
Pav(609) vs. CTRL	111.67	4.40	Yes		
Pav(1209) vs. CTRL	104.07	4.01	Yes		
Mixed vs. Baseline	127.48	3.05	No		
Mixed vs. TI	120.38	4.82	No		
Mixed vs. Pav(1209)	63.54	2.51	No		
Mixed vs. Pav(609)	55.94	2.26	No		
Mixed vs. Cg	55.10	2.23	No		
Mixed vs. ISO	35.28	1.36	No		
Mixed vs. Cm	2.82	0.12	No		
Cm vs. Baseline	124.66	2.97	No		
Cm vs. TI	117.56	4.64	No		
Cm vs. Pav(1209)	60.72	2.36	No		
Cm vs. Pav(609)	53.12	2.12	No		
Cm vs. Cg	52.28	2.08	No		
Cm vs. ISO	32.46	1.24	No		
ISO vs. Baseline	92.20	2.14	No		
ISO vs. TI	85.10	3.16	No		

Comparison	Diff of Ranks	Q	P<0.05
ISO vs. Pav(1209)	28.26	1.04	No
ISO vs. Pav(609)	20.66	0.77	No
ISO vs. Cg	19.82	0.74	No
Cg vs. Baseline	72.38	1.71	No
Cg vs. Tl	65.28	2.53	No
Cg vs. Pav(1209)	8.44	0.32	No
Cg vs. Pav(609)	0.84	0.03	No
Pav(609) vs. Baseline	71.54	1.69	No
Pav(609) vs. TI	64.44	2.50	No
Pav(609) vs. Pav(1209)	7.60	0.29	No
Pav(1209) vs. Baseline	63.94	1.50	No
Pav(1209) vs. TI	56.84	2.15	No
TI vs. CTRL	47.22	1.84	No
TI vs. Baseline	7.10	0.17	No
Baseline vs. CTRL	40.13	0.95	No

Table 3.4B (Continued)

Analyses of the left valve shell mass by experimental diet (Table 3.5) established significant differences among ten diets with subsequent ranking as follows: Mixed > CTRL, Mixed > Baseline, Mixed > TI, Cm > CTRL, Cm > Baseline, Cm > TI, ISO > CTRL, Pav(1209) > CTRL, Pav(609) > CTRL, Cg > Ctrl. No other comparisons were significant. These results are similar to those for analysis of shell length in that the Mixed, Cm, Cg, ISO, Pav609 and Pav1209 diets show significantly increased median values above the control. In addition, however, both the mixed diet and Cm diet proved to have larger median masses compared to the baseline and TI diet. Figure 3.6 depicts the median masses of each diet per collection and the summary of the statistical analyses are presented in Tables 3.5A and 3.5B.



Figure 3.5: Median left valve masses by treatment

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	Ν	Missing	Median	25%	75%
Baseline	14	1	0.037	0.0336	0.0419
CTRL	61	1	0.029	0.023	0.0405
ті	57	1	0.0452	0.0263	0.0572
ISO	50	1	0.0642	0.0414	0.102
Cg	59	1	0.0521	0.0311	0.102
Cm	64	1	0.0724	0.0421	0.13
Pav(609)	59	1	0.0524	0.0336	0.0814
Pav(1209)	54	1	0.0644	0.0367	0.0823
Mixed	68	1	0.0884	0.0361	0.139
	H = 81.0 with	8 degrees of freedo	om (P = <0.001)		

Table 3.5A: Results of analyses of left valve mass by treatment.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Mixed vs. CTRL	175.408	7.159	Yes	
Mixed vs. Baseline	143.769	3.441	Yes	
Mixed vs. TI	105.857	4.241	Yes	
Cm vs. CTRL	174.527	7.019	Yes	
Cm vs. Baseline	142.888	3.403	Yes	
Cm vs. Tl	104.976	4.147	Yes	
ISO vs. CTRL	149.806	5.644	Yes	
Pav(1209) vs. CTRL	129.437	4.981	Yes	
Pav(609) vs. CTRL	119.158	4.695	Yes	
Cg vs. CTRL	118.693	4.676	Yes	
Mixed vs. Cg	56.716	2.294	No	
Mixed vs. Pav(609)	56.25	2.275	No	
Mixed vs. Pav(1209)	45.972	1.814	No	
Mixed vs. ISO	25.602	0.988	No	
Mixed vs. Cm	0.881	0.0364	No	
Cm vs. Cg	55.835	2.226	No	
Cm vs. Pav(609)	55.369	2.207	No	
Cm vs. Pav(1209)	45.091	1.755	No	
Cm vs. ISO	24.721	0.942	No	
ISO vs. Baseline	118.167	2.748	No	
ISO vs. TI	80.255	2.976	No	
ISO vs. Cg	31.113	1.163	No	
ISO vs. Pav(609)	30.648	1.146	No	
ISO vs. Pav(1209)	20.37	0.746	No	
Pav(1209) vs. Baseline	97.798	2.292	No	
Pav(1209) vs. TI	59.885	2.267	No	
Pav(1209) vs. Cg	10.744	0.41	No	
Pav(1209) vs. Pav(609)	10.278	0.392	No	
Pav(609) vs. Baseline	87.519	2.069	No	
Pav(609) vs. TI	49.607	1.921	No	
Pav(609) vs. Cg	0.466	0.0182	No	
Cg vs. Baseline	87.054	2.058	No	
Cg vs. TI	49.142	1.903	No	
TI vs. Ctrl	69.551	2.716	No	
TI vs. Baseline	37.912	0.893	No	
Baseline vs. CTRL	31.639	0.75	No	

Figure 3.5B: Results of post hoc pairwise analyses of left valve mass and treatment.

Figure 3.6 depicts the median mass of the collected right valves per diet in each of the collections. During statistical analyses, it was determined that the valve mass of clams in each the Cm, ISO, Mixed, Pav(1209), and Pav(609) treatments was significantly different from the CTRL group (Tables 3.6A and 3.6B). Additionally, Pav(1209) vs. Cg was determined to be significant. No other comparisons were significant in terms of valve mass and diet received. The general trend that valve mass of clams fed unique diets increased compared to the control, as seen in both the valve length comparisons and left valve mass, is again supported by data for the right valve mass. The main difference from previously presented comparisons is that of Pav1209 versus Cg.





One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	Ν	Missing	Median	25%	75%
Baseline	14	1	0.0372	0.0336	0.043
CTRL	26	1	0.0305	0.0219	0.0417
ТІ	17	1	0.0479	0.0402	0.0538
ISO	10	1	0.0634	0.0405	0.11
Cg	19	1	0.0333	0.0203	0.0448
Cm	27	1	0.0671	0.0375	0.0836
Pav(609)	19	1	0.0628	0.0345	0.0809
Pav(1209)	14	1	0.0785	0.0549	0.0932
Mixed	31	1	0.0523	0.0337	0.0918
	H = 38.3 with	8 degrees of freedo	m (P = <0.001)		

Table 3.6A: Results of analysis of right valve mass comparison by treatment.

Table 3.6B: Results of post hoc pairwise comparisons of right valve shell mass by diet.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :					
Comparison	Diff of Ranks	Q	P<0.05		
Cm vs. CTRL	54.746	4.018	Yes		
ISO vs. CTRL	61.122	3.233	Yes		
Mixed vs. CTRL	49.217	3.736	Yes		
Pav(1209) vs. Cg	59.543	3.363	Yes		
Pav(1209) vs. CTRL	64.054	3.851	Yes		
Pav(609) vs. CTRL	51.456	3.422	Yes		
Baseline vs. Cg	10.697	0.604	No		
Baseline vs. CTRL	15.208	0.914	No		
Cg vs. CTRL	4.511	0.3	No		
Cm vs. Baseline	39.538	2.393	No		
Cm vs. Cg	50.235	3.368	No		
Cm vs. Mixed	5.529	0.424	No		
Cm vs. Pav(609)	3.291	0.221	No		
Cm vs. TI	15.534	1.005	No		
ISO vs. Baseline	45.915	2.177	No		
ISO vs. Cg	56.611	2.851	No		
ISO vs. Cm	6.376	0.339	No		
ISO vs. Mixed	11.906	0.644	No		
ISO vs. Pav(609)	9.667	0.487	No		
ISO vs. TI	21.91	1.081	No		
Mixed vs. Baseline	34.009	2.106	No		
Mixed vs. Cg	44.706	3.083	No		

Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. TI	10.004	0.664	No
Pav(1209) vs. Baseline	48.846	2.56	No
Pav(1209) vs. Cm	9.308	0.563	No
Pav(1209) vs. ISO	2.932	0.139	No
Pav(1209) vs. Mixed	14.837	0.919	No
Pav(1209) vs. Pav(609)	12.598	0.712	No
Pav(1209) vs. Tl	24.841	1.368	No
Pav(609) vs. Baseline	36.248	2.047	No
Pav(609) vs. Cg	46.944	2.895	No
Pav(609) vs. Mixed	2.239	0.154	No
Pav(609) vs. TI	12.243	0.733	No
TI vs. Baseline	24.005	1.322	No
TI vs. Cg	34.701	2.076	No
TI vs. CTRL	39.212	2.518	No

Table 3.6B (Continued)

Differences between the left and right valves: valve length and mass

Further analyses were performed to determine the difference between the valves within each of the feeding groups during the baseline, March and May collections in reference to length and mass. Three separate comparative analyses were performed: 1) an overall comparison of left and right valve independent of time and diet, 2) a comparison of the baseline valves and all experimental valves, and 3) comparison within each individual group.

The left and right valves were similar in length when considering all valves measured with no significant difference found (P=0.233) between neither left and right valve composites nor between the left and right valve lengths of the baseline and feeding group composite (P=0.334). The valve lengths within each feeding group were found to be virtually identical (P=1.00).

Figure 3.7 depicts the masses of both the left and right valves collected for each diet by month. The statistical comparisons revealed no significant differences between the composited right and left valves (P=0.877), the left and right valves for the experimental and base line groups (P=0.118), or the left and right valves within the specific feeding groups. Tables 3.7-3.8 provide the summary of the statistical tests associated with the comparisons of valve mass.



Figure 3.7.Median left (L) and right (R) valve mass by diet received.

One Way Analysis of Variance						
Normality Test:	Failed (P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	Ν	Missing	Median	25%	75%	
Left - all	169	1	0.0445	0.0314	0.0721	
Right All	169	1	0.0445	0.03	0.0741	
H = 0.024 with 1 degrees of freedom (P = 0.877)						

Table 3.7A: Results of analyses of all valves collected, left vs. right valve mass.

Table 3.7B: Summa	y results of ana	lyses of baseline	data versus ex	perimental com	posite.
-------------------	------------------	-------------------	----------------	----------------	---------

One Way Analysis of Variance						
Normality Test:	Failed	(P < 0.050)				
Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	N	Missing	Median	25%	75%	
Baseline-Left	14	1	0.037	0.0336	0.0419	
Baseline-Right	14	1	0.0372	0.0336	0.043	
Experimental-Left	156	1	0.045	0.0303	0.0748	
Experimental-Right	156	1	0.0454	0.0294	0.0752	
H = 5.87 with 3 degrees of freedom (P = 0.118)						

Table 3.8A: Summary of results for diet specific left and right valve comparisons.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	Ν	Missing	Median	25%	75%
Baseline-Left	14	1	0.037	0.0336	0.0419
Baseline-Right	14	1	0.0372	0.0336	0.043
CTRL-Left	26	1	0.0321	0.0229	0.0429
CTRL-Right	26	1	0.0305	0.0219	0.0417
TI-Left	17	1	0.0467	0.04	0.0557
TI-Right	17	1	0.0479	0.0402	0.0538
ISO-Left	10	1	0.0642	0.0422	0.107
ISO-Right	10	1	0.0634	0.0405	0.11
Cg-Left	19	1	0.0372	0.0192	0.0445
Cg-Right	19	1	0.0333	0.0203	0.0448
Cm-Left	27	1	0.0642	0.0395	0.0833
Cm-Right	27	1	0.0671	0.0375	0.0836
Pav(609)-Left	19	1	0.0626	0.0336	0.0814
Pav(609)-Right	19	1	0.0628	0.0345	0.0809
Pav(1209)-Left	14	1	0.0777	0.0542	0.0934
Pav(1209)-Right	14	1	0.0785	0.0549	0.0932
Mixed-Left	31	1	0.0516	0.0339	0.0908
Mixed-Right	31	1	0.0523	0.0337	0.0918
H = 74.7 with 17 degrees of freedom (P = < 0.001)					

All Pairwise Multiple Comparison Procedures (Dunn's Method) :					
Comparison	Diff of Ranks	Q	P<0.05		
Baseline-Right vs. Baseline-Left	1.192	0.0313	No		
Cg-Left vs. Cg-Right	0.278	0.00858	No		
Cm-Left vs. Cm-Right	5.577	0.207	No		
CTRL-Left vs. CTRL-Right	12.1	0.44	No		
ISO-Right vs. ISO-Left	0.167	0.00364	No		
Mixed-Right vs. Mixed-Left	2.583	0.103	No		
Pav(1209)-Right vs. Pav(1209)-Left	4.231	0.111	No		
Pav(609)-Right vs. Pav(609)-Left	0.444	0.0137	No		
TI-Right vs. TI-Left	1.25	0.0364	No		

Table 3.8B: Results of post hoc pairwise analyses of diet specific left and right valve comparisons.

Elemental shell chemistry

Due to the results from the ICP-OES and the post- processing procedures, described previously, the total number of shells (n) is specific to each experimental diet and element ratio analyzed. Only the concentrations of boron, barium, cadmium, calcium, cobalt, copper, iron, potassium, lithium, magnesium, manganese, nickel, potassium, lead, silica, strontium, and zinc were detected from each single valve sample overall, so, only these elements are reported herein. Subsequent analyses were further limited by the observed variability and the different number of quantifiable observations for each element by diet which restricted reporting of differences in cadmium, cobalt, copper, nickel and lead.

The associated chemistries are provided for all collections for the left valves while the right valve chemistries are only available for the Baseline, March and May collections. Figures 3.8 – 3.20 depict the median element/Ca for each diet by collection month. The baseline measures for P/Ca and Sr/Ca, all Cd/Ca, all Ni/Ca, and Co/Ca for the right valves are not depicted due to measurement complications or elimination during post-processing as previously described.

The provided graphics (Figures 3.8-3.20) illustrate the variability in element/calcium ratios among the different diets and collections. Visually, it is evident that many of the reported ratios decreased from baseline during the course of the experiment. It is also obvious that there is not a clear trend over time with multiple diets showing random spikes in specific ratios.

Some of the observed elemental ratios do appear to remain relatively stable (i.e., B/Ca) throughout the experimental time frame regardless of diet received, while others are extremely variable. In many cases, the left and right values do not mirror one another, though usually representative of the same clams.



Figure 3.8A: Median B/Ca versus diet received for the collected left valves by collection month. The feeding groups are labeled as follows: starvation control = CTRL, TI = *Isochrysis* sp. (CCMP1324), ISO = *Isochrysis* sp. (CCMP1611), Pav609 = *Pavlova* pinguis (CCMP609), Pav1209 = *Pavlova* sp. (CCMP1209), CG = Chaetoceros galvestonensis (CCMP186), CM = Chaetoceros mulleri (CCMP609), and Mixed = the mixed diet of all species.



Figure 3.8B: Median B/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



Figure 3.9A: Median Ba/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



Figure 3.9B: Median Ba/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



Figure 3.10: Median Co/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



Figure 3.11A: Median Cu/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



Figure 3.11B: Median Cu/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.12A: Median Fe/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.12B: Median Fe/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.13A: Median K/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.13B: Median K/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.14A: Median Li/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.14B: Median Li/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.15A: Median Mg/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.15B: Median Mg/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.16A: Median Mn/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.16B: Median Mn/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.17A: Median P/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.17B: Median P/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.18A: Median Si/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.18B: Median Si/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.19A: Median Sr/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.19B: Median Sr/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.20A: Median Zn/Ca versus diet received for the collected left valves by collection month. The diet labels are the same as those in Fig. 3.8A.



3.20B: Median Zn/Ca versus diet received for the collected right valves by collection month. The diet labels are the same as those in Fig. 3.8A.
Statistical comparison of elemental shell chemistry between left and right valves

Two levels of analysis for examination of possible differences between left and right valve shell chemistry were conducted:1) all left and right valves were compared regardless of diet received to determine any chemical differences that might be expected at a population or species level, and 2) the right and left valves specific to the diet received to determine any differences in shell chemistry resultant of experimental variables and procedures as well as determine dominant influences observed during the first left versus right valve analysis.

Analysis of all collected left and right valves resulted in only two ratios being determined significantly different in regard to valve orientation – Ni/Ca and Zn/Ca. The summary of statistical tests is presented in Tables 3.9 and 3.10. The difference in Ni/Ca between valves was based on fewer than 20 left and right valves, which were representative of only a few diets. Though a significant difference was determined (P<0.05), these results were primarily attributable to differences between the Control and Baseline with the other diets unevenly represented with regard to valve orientation. Zinc measurements were, on the other hand, represented for all diets and in each collection. A P value of <0.05 was determined from the Dunn's method pairwise analysis and established right valve Zn/Ca > left valve Zn/Ca.

When each individual diet was examined with regard to the differences between left and right valve elemental chemistry, significant differences were observed in both the Control (CTRL) and *Chaetoceros mulleri* (CM) diet groups (Table 3.11). Three ratios, B/Ca, Si/Ca and Zn/Ca, were identified as being significantly higher in the right valve compared to the left valve in the Control (P=0.001, 0.046, and 0.002 respectively).

83

Only the Ba/Ca ratio was identified as significantly different between valves (P=0.030) in the CM group, though Li/Ca (P=0.053) was borderline. The right valve was again identified as having a higher ratio.

Ni										
One Way Analysis of Variance										
Normality Test:	Passed (P = 0.831)									
Equal Variance Test:	Passed		(P =	0.256)						
Group Name	N	Missing	Mean	Std Dev	SEM					
Left	143	124	9.04E-07	4.58E-07	1.05E-07					
Right	152	137	4.74E-07	3.45E-07	8.91E-08					
Source of Variation	DF	SS MS F P								
Between Groups	1	1.55E-12	1.55E-12	9.138	0.005					
Residual	32	5.44E-12	1.70E-13							
Total	33	7.00E-12								
	Power of per	formed test with	alpha = 0.050: 0.803							
	All Pairwise Multiple (Comparison Prod	cedures (Holm-Sidak m	nethod):						
Overall significance level = 0.05										
Comparison	Diff of Means	t	Unadjusted P	Critical Level	Significant?					
Left vs. Right	4.31E-07	3.023	0.0049	0.05	Yes					

Table 3.9: Results for left versus right valve comparisons with respect to Ni/Ca.

Table 3.10: Results for left versus right valve comparisons with respect to Zn/Ca.

Zn									
	One Wa	ay Analysis of Vari	ance						
Normality Test:	Failed		(P <	0.050)					
	Kruskal-Wallis One	Way Analysis of V	ariance on Ranks						
Group	Ν	Missing	Median	25%	75%				
Left	167	35 1.65E-05 1.08E-05 3.37E-06							
Right	167	46	2.30E-05	1.35E-05	3.79E-05				
	H = 3.88 with 1 c	legrees of freedon	n. (P = 0.049)						
All Pairwise Multiple Comparison Procedures (Dunn's Method) :									
Comparison	Comparison Diff of Ranks Q P<0.05								
Left vs. Right	18.137	1.969		Yes					

All associated P-values for the statistical tests used for each diet and elemental ratio are provided in Table 3.11. The median ratio values with 95% confidence intervals are depicted in Figures 3.21 - 3.24 for the CTRL and CM examinations. As well, Tables 3.12 - 3.15 provide the statistical test summaries for the CTRL and CM left and right valve group comparisons.

	Significance of diet-specific le	it and right	valve compansons.		
BI	Left vs. Right P value	<u>TI</u>	Left vs. Right P value	ISO	Left vs. Right P value
В	0.391	В	0.609	В	0.821
Ba	0.892	Ba	0.102	Ba	0.735
Cd	Na	Cd	Na	Cd	Na
Со	0.747	Со	1.000	Со	Na
Cu	1.000	Cu	0.640	Cu	0.314
Fe	0.765	Fe	0.877	Fe	0.158
к	0.807	κ	0.954	κ	0.245
Li	0.765	Li	0.765	Li	0.885
Mg	0.977	Mg	0.721	Mg	0.229
Mn	0.840	Mn	0.369	Mn	0.209
Ni	Na	Ni	Na	Ni	Na
Р	Na	P	0.598	Р	0.994
Pb	Na	Pb	Na	Pb	Na
Si	0.887	Si	0.841	Si	0.102
Sr	Na	Sr	0.554	Sr	0.996
Zn	0.840	Zn	0.910	Zn	0.284
<u>CG</u>	Left vs. Right P value	<u>CM</u>	Left vs. Right P value	<u>Pav609</u>	Left vs. Right P value
В	0.580	В	0.309	В	0.728
Ва	0.937	Ba	0.030	Ba	0.817
Cd	Na	Cd	Na	Cd	Na
Со	Na	Со	0.410	Со	0.451
Cu	0.397	Cu	0.297	Cu	0.381
Fe	0.669	Fe	0.750	Fe	0.531
к	0.856	κ	0.489	к	0.385
Li	0.978	Li	0.053	Li	0.753
Mg	0.319	Mg	0.705	Mg	0.197
Mn	0.812	Mn	0.922	Mn	0.978
Ni	Na	Ni	Na	Ni	Na

Table 3.11: Significance of diet-specific left and right valve comparisons.

CG	Left vs. Right P value	CM	Left vs. Right P value	Pav609	Left vs. Right P value
Р	0.517	Р	0.962	Р	0.394
Pb	Na	Pb	Na	Pb	Na
Si	0.740	Si	0.421	Si	0.717
Sr	0.809	Sr	0.091	Sr	0.994
Zn	0.812	Zn	0.125	Zn	0.869
Pav1209	Left vs. Right P value	CTRL	Left vs. Right P value	Mixed	Left vs. Right P value
В	0.716	В	0.001	В	0.418
Ba	0.505	Ba	0.222	Ba	0.554
Cd	Na	Cd	Na	Cd	Na
Со	0.391	Со	0.418	Со	0.189
Cu	0.878	Cu	0.231	Cu	0.323
Fe	0.915	Fe	0.831	Fe	0.845
к	0.837	κ	0.421	κ	0.796
Li	0.864	Li	0.171	Li	0.687
Mg	0.779	Mg	0.614	Mg	0.829
Mn	0.887	Mn	0.771	Mn	0.580
Ni	Na	Ni	0.123	Ni	Na
Р	0.608	Р	0.561	Р	0.473
Pb	Na	Pb	Na	Pb	Na
Si	0.878	Si	0.046	Si	0.845
Sr	0.561	Sr	0.985	Sr	0.798
Zn	0.918	Zn	0.002	Zn	0.820

Table 3.11 (Continued)

Comparison of elemental shell chemistry by to diet

Statistical comparisons of differences in shell chemistry between experimental diets were performed using the composite of all month collections by diet on both the left and right valves. Kruskal-Wallis One way Analysis of Variance on Ranks was used, followed by appropriate post-hoc pairwise examinations to determine if there were significant differences among diets with regard to elemental shell chemistry and which elements, if any, were influenced. It is important to recognize that the comparison of right valves only included individuals from the Baseline, March and May collections, whereas, the left valve comparisons include individuals from all experimental collections

January – October as time and growth influences must be closely examined during interpretation.



Figure 3.21: Left versus right valve for the Control feeding group with regard to B/Ca.

Table 3.12: Results of the comparison	of the left and	right valves from	the Control group	with regard to
B/Ca.		-		-

CTRL-B									
t-test									
Normality Test: Failed (P < 0.050)									
Mann-Whitney Rank Sum Test									
Group	N	Missing Median 25% 75%							
Left	26	1	2.83E-05	2.40E-05	3.22E-05				
Right	26	1	5.23E-05	3.09E-05	7.90E-04				
Mann-Whitney U Statistic= 510.000									
T = 440 n(small)= 25 n(big)= 25 (P = <0.001)									



Figure 3.22: Left versus right valve for the Control feeding group with regard to Si/Ca.

Table 3.13: Results of the	comparison of the le	eft and right valves	from the Control gro	up with regard to
Si/Ca.		-	-	

CTRL-Si										
		t-test								
Normality Test: Failed (P < 0.050)										
Mann-Whitney Rank Sum Test										
Group	Ν	Missing	Median	25%	75%					
Left	26	1	1.65E-04	1.42E-04	2.30E-04					
Right	Right 26 1 1.99E-04 1.73E-04 2.47E-04									
Mann-Whitney U Statistic= 416.000										
	T = 534	1 n(small)= 25 n(b	ig) = 25 (P = 0.046)							

Inadequate sample sizes preclude analyses for Cd/Ca, Cu/Ca, Ni/Ca, and Pb/Ca.

Comparisons P/Ca and Sr/Ca are included despite baseline comparisons not being available.

Ten differences based on diet were identified for B/Ca of the left valves: Cg vs. ISO, Cg vs. Pav609, Mixed vs. ISO, Mixed vs. Pav609, TI vs. Baseline, TI vs. Cm, TI vs.

CTRL, TI vs. ISO, TI vs. Pav1209, and TI vs. Pav609 (Table 3.16B). The median values and associated 95% confidence intervals are provided in Figure 3.25, and the analyses summaries are provided in Tables 3.16 and 3.17. The analyses support the



Figure 3.23: Left versus right valve for the Control feeding group with regard to Zn/Ca.

Z1/Ca.										
CTRL-Zn										
t-test										
Normality Test: Failed (P < 0.050)										
Mann-Whitney Rank Sum Test										
Group	N	N Missing Median 25% 75%								
Left	26	2	1.71E-05	1.15E-05	3.16E-05					
Right	26	1	3.40E-05	2.44E-05	4.68E-05					
Mann-Whitney U Statistic= 454.000										
	T = 446 n(small) = 24 n(big) = 25 (P = 0.002)									

Table 3.14: Results of the comparison	of the	left and	right	valves f	from t	the	Control	group	with	regard	to
Zn/Ca.			-							-	



Figure 3.24: Left versus right valve for the CM feeding group with regard to Ba/Ca.

CM-Ba										
t-test										
Normality Test: Failed (P < 0.050)										
Mann-Whitney Rank Sum Test										
Ν	Missing	Median	25%	75%						
27	9	1.44E-05	1.23E-05	1.70E-05						
27	9	1.69E-05	1.53E-05	1.81E-05						
Mann-Whitney U Statistic= 231.000										
T = 264 n(small) = 18 n(big) = 18 (P = 0.030)										
	Failed N 27 27 27 Ma T = 264	CM-Ba t-test Failed Mann-Whitney Ran N Missing 27 9 27 9 27 9 27 9 T = 264 n(small) = 18 n(bi	CM-Ba t-test Failed (P < Failed Mann-Whitney Rank Sum Test N Missing Median 27 9 1.44E-05 27 9 1.69E-05 Mann-Whitney U Statistic= 231.000 T = 264 n(small)= 18 n(big)= 18 (P = 0.030)	CM-Ba t-test Failed (P < 0.050) Failed (P < 0.050) Mann-Whitney Rank Sum Test Sum Test N Missing Median 25% 27 9 1.44E-05 1.23E-05 27 9 1.69E-05 1.53E-05 Mann-Whitney U Statistic= 231.000 T = 264 n(small)= 18 n(big)= 18 (P = 0.030)						

Table 3.15: Results of the comparison of the left and right valves from the CM group with regard to Ba/Ca.

following ranking: Cg> ISO and Pav609, Mixed> ISO and Pav609, and finally, TI>Baseline, Cm, CTRL, ISO, Pav609, and Pav1209.

The comparison of right valve B/Ca between experimental diet groups determined that only the CTRL diet was significantly different from the Baseline, Cm,

Mixed, Pav1209, and Pav609 diets (Table 17B). The associated ranking is as follows: CTRL > Baseline, Cm, Mixed, Pav1209, and Pav609.



Figure 3.25: Left valve median B/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as follows: Baseline = baseline/January collection, starvation control = CTRL, TI = *Isochrysis* sp. (CCMP1324), ISO = *Isochrysis* sp. (CCMP1611), Pav609 = *Pavlova pinguis* (CCMP609), Pav1209 = *Pavlova sp.* (CCMP1209), *Cg* = *Chaetoceros galvestonensis* (CCMP186), Cm = *Chaetoceros mulleri* (CCMP609), and Mixed = the mixed diet of all species. The lower graph is a close up of the first to allow a closer examination not limited by the error bar range.

Table 3.16A: Results of analysis of variance of left valve B/Ca among experimental diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance									
Normality Test:	Failed	(P < 0.050)							
Kruskal-Wallis One Way Analysis of Variance on Ranks									
Group	N	Missing	Median	25%	75%				
Baseline	14	1	2.86E-05	2.71E-05	3.17E-05				
CTRL	61	1	3.05E-05	2.39E-05	5.23E-05				
ТІ	54	4	4.09E-05	3.13E-05	5.39E-05				
ISO	50	1	2.76E-05	1.92E-05	3.20E-05				
Cg	58	3	3.30E-05	2.64E-05	3.92E-05				
Cm	64	8	2.88E-05	2.44E-05	3.70E-05				
Pav609	59	7	2.84E-05	2.39E-05	3.18E-05				
Pav1209	52	1	3.14E-05	2.59E-05	3.96E-05				
Mixed	53	13	3.63E-05	2.91E-05	1.35E-04				
H = 61.1 with 8 degrees of freedom (P = <0.001)									

Table 3.16B: Results of pairwise comparison of left valve B/Ca between experimental diets. Diets are represented as previously identified in Figure 3.25.

Comparison	Diff of Ranks	0	P<0.05
Cq vs. ISO	87.071	3.6	Yes
Cg vs. Pav609	76.485	3.212	Yes
Mixed vs. ISO	123.566	4.71	Yes
Mixed vs. Pav609	112.981	4.363	Yes
TI vs. Baseline	124.238	3.241	Yes
TI vs. Cm	103.325	4.313	Yes
TI vs. CTRL	75.717	3.212	Yes
TI vs. ISO	149.516	6.041	Yes
TI vs. Pav1209	87.759	3.582	Yes
TI vs. Pav609	138.931	5.697	Yes
Baseline vs. ISO	25.278	0.658	No
Baseline vs. Pav609	14.692	0.385	No
Cg vs. Baseline	61.793	1.627	No
Cg vs. Cm	40.88	1.749	No
Cg vs. Ctrl	13.271	0.577	No
Cg vs. Pav1209	25.313	1.058	No
Cm vs. Baseline	20.913	0.552	No
Cm vs. ISO	46.191	1.918	No
Cm vs. Pav609	35.606	1.502	No
CTRL vs. Baseline	48.522	1.288	No

Comparison	Diff of Ranks	Q	P<0.05
CTRL vs. Cm	27.608	1.207	No
CTRL vs. ISO	73.8	3.113	No
CTRL vs. Pav1209	12.042	0.514	No
CTRL vs. Pav609	63.214	2.71	No
Mixed vs. Baseline	98.288	2.501	No
Mixed vs. Cg	36.495	1.426	No
Mixed vs. Cm	77.375	3.036	No
Mixed vs. Ctrl	49.767	1.98	No
Mixed vs. Pav1209	61.809	2.377	No
Pav1209 vs. Baseline	36.48	0.954	No
Pav1209 vs. Cm	15.566	0.653	No
Pav1209 vs. ISO	61.758	2.508	No
Pav1209 vs. Pav609	51.172	2.109	No
Pav609 vs. ISO	10.586	0.432	No
TI vs. Cg	62.445	2.596	No
TI vs. Mixed	25.95	0.994	No

Table 3.16B (Continued)

Table 3.17A: Results of analysis of variance of right valve B/Ca among experimental diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
	Kruskal-Wa	llis One Way Analy	sis of Variance on Ra	nks	
Group	N	Missing Median 25% 75%			
Baseline	14	2	2.79E-05	2.68E-05	2.99E-05
CTRL	29	1	4.61E-05	3.08E-05	7.77E-04
ТІ	17	7	3.42E-05	2.73E-05	3.81E-05
ISO	10	1	3.02E-05	2.40E-05	3.25E-05
Cg	19	1	3.31E-05	2.67E-05	3.87E-05
Cm	27	9	2.87E-05	2.41E-05	3.10E-05
Pav609	19	9	2.62E-05	2.22E-05	3.11E-05
Pav1209	14	1	2.35E-05	2.11E-05	2.67E-05
Mixed	31	13	2.98E-05	2.57E-05	3.18E-05
	H = 38.9	with 8 degrees of	freedom (P = <0.001)		

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
CTRL vs. Baseline	44.833	3.298	Yes	
CTRL vs. Cm	41.583	3.493	Yes	
CTRL vs. Mixed	38.194	3.208	Yes	
CTRL vs. Pav1209	71.865	5.434	Yes	
CTRL vs. Pav609	50.35	3.469	Yes	
Baseline vs. Pav1209	27.032	1.714	No	
Baseline vs. Pav609	5.517	0.327	No	
Cg vs. Baseline	23.139	1.576	No	
Cg vs. Cm	19.889	1.514	No	
Cg vs. ISO	13.111	0.815	No	
Cg vs. Mixed	16.5	1.256	No	
Cg vs. Pav1209	50.171	3.498	No	
Cg vs. Pav609	28.656	1.844	No	
Cm vs. Baseline	3.25	0.221	No	
Cm vs. Pav1209	30.282	2.111	No	
Cm vs. Pav609	8.767	0.564	No	
CTRL vs. Cg	21.694	1.822	No	
CTRL vs. ISO	34.806	2.305	No	
CTRL vs. TI	20.25	1.395	No	
ISO vs. Baseline	10.028	0.577	No	
ISO vs. Cm	6.778	0.421	No	
ISO vs. Mixed	3.389	0.211	No	
ISO vs. Pav1209	37.06	2.169	No	
ISO vs. Pav609	15.544	0.859	No	
Mixed vs. Baseline	6.639	0.452	No	
Mixed vs. Cm	3.389	0.258	No	
Mixed vs. Pav1209	33.671	2.348	No	
Mixed vs. Pav609	12.156	0.782	No	
Pav609 vs. Pav1209	21.515	1.298	No	
TI vs. Baseline	24.583	1.457	No	
TI vs. Cg	1.444	0.0929	No	
TI vs. Cm	21.333	1.373	No	
TI vs. ISO	14.556	0.804	No	
TI vs. Mixed	17.944	1.155	No	
TI vs. Pav1209	51.615	3.114	No	
TI vs. Pav609	30.1	1.708	No	

Table 3.17B: Results of pairwise comparison of right valve B/Ca between experimental diets. Diets are represented as previously identified in Figure 3.25.

The median Ba/Ca value for left valves by diet is depicted in Figure 3.26. Analysis of variance and post hoc pairwise analysis revealed 10 significant differences based on diet: Baseline vs. Cg, Baseline vs. Cm, Baseline vs. CTRL, Baseline vs. ISO, Baseline vs. Mixed, Baseline vs. Pav1209, Baseline vs. Pav609, Baseline vs. TI, ISO vs. Mixed, and TI vs. Mixed (Tables 3.18A and 3.18B). The ranking of these diet groups is as follows: Baseline > Cg, Cm, CTRL, ISO, Mixed, Pav1209, Pav609, and TI, as well as, TI and ISO > Mixed.

The comparison of Ba/Ca values between diets using the right valves resulted in four significant differencest: Baseline vs. CTRL, Baseline vs. ISO, Baseline vs. Mixed, and Baseline vs. TI. These differences support Baseline Ba/Ca > CTRL, ISO, Mixed, and TI. Summaries of the statistical results have been provided in Tables 3.19A and 3.19B.



Figure 3.26: Left valve median Ba/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance						
Normality Test:	Failed		(P <	0.050)		
Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	N	Missing Median 25% 75%				
Baseline	14	1	2.21E-05	1.83E-05	2.65E-05	
CTRL	61	1	1.36E-05	1.17E-05	1.51E-05	
ТІ	54	8	1.43E-05	1.18E-05	1.69E-05	
ISO	50	1	1.41E-05	1.31E-05	1.58E-05	
Cg	58	3	1.29E-05	1.15E-05	1.56E-05	
Cm	64	9	1.32E-05	1.18E-05	0.000015	
Pav609	59	7	0.000013	1.18E-05	1.46E-05	
Pav1209	52	1	1.33E-05	1.15E-05	1.55E-05	
Mixed	53	9	1.06E-05	7.99E-06	1.53E-05	
	H = 50.	7 with 8 degrees of	freedom (P = <0.001))		

Table 3.18A: Results of analysis of variance of left valve Ba/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.18B: Results of pairwise comparison of left valve Ba/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :					
Comparison	Diff of Ranks	Q	P<0.05		
Baseline vs. Cg	189.568	5.004	Yes		
Baseline vs. Cm	183.077	4.833	Yes		
Baseline vs. Ctrl	177.76	4.731	Yes		
Baseline vs. ISO	145.832	3.806	Yes		
Baseline vs. Mixed	253.509	6.538	Yes		
Baseline vs. Pav1209	177.587	4.653	Yes		
Baseline vs. Pav609	189.269	4.969	Yes		
Baseline vs. TI	149.946	3.886	Yes		
ISO vs. Mixed	107.677	4.221	Yes		
TI vs. Mixed	103.562	3.998	Yes		
Cg vs. Mixed	63.941	2.574	No		
Cm vs. Cg	6.491	0.277	No		
Cm vs. Mixed	70.432	2.835	No		
Cm vs. Pav609	6.192	0.261	No		
CTRL vs. Cg	11.808	0.515	No		
CTRL vs. Cm	5.317	0.232	No		
CTRL vs. Mixed	75.748	3.107	No		
CTRL vs. Pav609	11.509	0.495	No		
ISO vs. Cg	43.736	1.813	No		
ISO vs. Cm	37.245	1.544	No		

Comparison	Diff of Ranks	Q	P<0.05
ISO vs. Ctrl	31.928	1.35	No
ISO vs. Pav1209	31.755	1.292	No
ISO vs. Pav609	43.437	1.776	No
ISO vs. TI	4.114	0.163	No
Pav1209 vs. Cg	11.981	0.502	No
Pav1209 vs. Cm	5.49	0.23	No
Pav1209 vs. Ctrl	0.174	0.00742	No
Pav1209 vs. Mixed	75.922	3.004	No
Pav1209 vs. Pav609	11.683	0.483	No
Pav609 vs. Cg	0.299	0.0126	No
Pav609 vs. Mixed	64.24	2.553	No
TI vs. Cg	39.621	1.614	No
TI vs. Cm	33.13	1.35	No
TI vs. Ctrl	27.814	1.155	No
TI vs. Pav1209	27.64	1.107	No
TI vs. Pav609	39.323	1.582	No

Table 3.19A: Results of analysis of variance of right valve Ba/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
Baseline	14	2	2.36E-05	1.87E-05	2.70E-05
Ctrl	29	1	1.49E-05	1.33E-05	1.77E-05
ТІ	17	7	1.46E-05	1.01E-05	1.49E-05
ISO	10	1	1.28E-05	1.24E-05	1.71E-05
Cg	19	1	1.75E-05	1.08E-05	2.27E-05
Cm	27	9	1.69E-05	1.53E-05	1.81E-05
Pav(609)	19	9	1.44E-05	1.23E-05	1.96E-05
Pav(1209)	14	1	1.41E-05	1.25E-05	2.22E-05
Mixed	31	11	1.20E-05	1.04E-05	1.65E-05
	H = 30.4	with 8 degrees of	freedom (P = <0.001)		

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Baseline vs. CTRL	46.869	3.398	Yes	
Baseline vs. ISO	57.306	3.25	Yes	
Baseline vs. Mixed	71.383	4.89	Yes	
Baseline vs. TI	66.833	3.904	Yes	
Baseline vs. Cg	39.417	2.645	No	
Baseline vs. Cm	31.139	2.09	No	
Baseline vs. Pav1209	43.353	2.709	No	
Baseline vs. Pav609	46.383	2.709	No	
Cg vs. Ctrl	7.452	0.617	No	
Cg vs. ISO	17.889	1.096	No	
Cg vs. Mixed	31.967	2.461	No	
Cg vs. Pav1209	3.936	0.27	No	
Cg vs. Pav609	6.967	0.442	No	
Cg vs. Tl	27.417	1.739	No	
Cm vs. Cg	8.278	0.621	No	
Cm vs. CTRL	15.73	1.302	No	
Cm vs. ISO	26.167	1.603	No	
Cm vs. Mixed	40.244	3.098	No	
Cm vs. Pav1209	12.214	0.839	No	
Cm vs. Pav609	15.244	0.967	No	
Cm vs. TI	35.694	2.264	No	
CTRL vs. ISO	10.437	0.681	No	
CTRL vs. Mixed	24.514	2.094	No	
CTRL vs. TI	19.964	1.355	No	
ISO vs. Mixed	14.078	0.877	No	
ISO vs. TI	9.528	0.519	No	
Pav1209 vs. CTRL	3.516	0.262	No	
Pav1209 vs. ISO	13.953	0.805	No	
Pav1209 vs. Mixed	28.031	1.968	No	
Pav1209 vs. Pav609	3.031	0.18	No	
Pav1209 vs. TI	23.481	1.396	No	
Pav609 vs. CTRL	0.486	0.033	No	
Pav609 vs. ISO	10.922	0.595	No	
Pav609 vs. Mixed	25	1.615	No	
Pav609 vs. TI	20.45	1.144	No	
TI vs. Mixed	4.55	0.294	No	

Table 3.19B: Results of pairwise comparison of right valve Ba/Ca between diets. Diets are represented as previously identified in Figure 3.25.

Statistical comparison of diet groups as relates to Co/Ca could only be performed on the left valves (Figure 3.27). Two significant differences were found: ISO > Mixed and TI > Mixed (Tables 3.20A and 3.20B).



Figure 3.27: Left valve median Co/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

Table 3.20A: Results of analysis of variance of left valve Co/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance						
Normality Test:	Failed	(P < 0.050)				
Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	N	Missing Median 25% 75%				
Baseline	12	7	1.36E-06	1.17E-06	1.71E-06	
CTRL	59	44	9.26E-07	7.46E-07	1.39E-06	
ТІ	53	44	1.93E-06	1.45E-06	2.21E-06	
ISO	48	35	1.90E-06	1.70E-06	2.12E-06	
Cg	57	40	1.58E-06	7.36E-07	1.79E-06	
Cm	63	31	9.77E-07	8.69E-07	1.91E-06	
Pav609	58	33	1.06E-06	9.26E-07	1.94E-06	
Pav1209	52	38	1.06E-06	8.91E-07	2.38E-06	
Mixed	53	35	8.27E-07	7.05E-07	1.17E-06	
	H = 22.6	6 with 8 degrees of	freedom ($P = 0.004$)			

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
ISO vs. Mixed	59.701	3.826	Yes	
TI vs. Mixed	57.778	3.301	Yes	
Baseline vs. Cg	16.8	0.77	No	
Baseline vs. Cm	12.144	0.589	No	
Baseline vs. CTRL	32.133	1.452	No	
Baseline vs. Mixed	43.578	2.011	No	
Baseline vs. Pav1209	9.586	0.429	No	
Baseline vs. Pav609	9.24	0.44	No	
Cg vs. CTRL	15.333	1.01	No	
Cg vs. Mixed	26.778	1.847	No	
Cm vs. Cg	4.656	0.362	No	
Cm vs. CTRL	19.99	1.49	No	
Cm vs. Mixed	31.434	2.489	No	
CTRL vs. Mixed	11.444	0.764	No	
ISO vs. Baseline	16.123	0.715	No	
ISO vs. Cg	32.923	2.085	No	
ISO vs. Cm	28.267	2.005	No	
ISO vs. CTRL	48.256	2.971	No	
ISO vs. Pav1209	25.709	1.557	No	
ISO vs. Pav609	25.363	1.73	No	
ISO vs. TI	1.923	0.103	No	
Pav1209 vs. Cg	7.214	0.466	No	
Pav1209 vs. Cm	2.558	0.186	No	
Pav1209 vs. CTRL	22.548	1.415	No	
Pav1209 vs. Mixed	33.992	2.225	No	
Pav609 vs. Cg	7.56	0.561	No	
Pav609 vs. Cm	2.904	0.254	No	
Pav609 vs. CTRL	22.893	1.635	No	
Pav609 vs. Mixed	34.338	2.591	No	
Pav609 vs. Pav1209	0.346	0.0242	No	
TI vs. Baseline	14.2	0.594	No	
TI vs. Cg	31	1.754	No	
TI vs. Cm	26.344	1.629	No	
TI vs. CTRL	46.333	2.563	No	
TI vs. Pav1209	23.786	1.299	No	
TI vs. Pav609	23.44	1.407	No	

Table 3.20B: Results of pairwise comparison of left valve Co/Ca between diets. Diets are represented as previously identified in Figure 3.25.

Figure 3.28 depicts the left valve Fe/Ca median values by diet received. In all, 15 comparisons were determined to be significant as relates to diet-specific Fe/Ca: Baseline vs. Cg, Baseline vs. Cm, Baseline vs. CTRL, Baseline vs. Mixed, ISO vs. Cg, ISO vs. Cm, ISO vs. CTRL, ISO vs. Mixed, Pav1209 vs. Cg, Pav1209 vs. Cm, Pav1209 vs. CTRL, Pav1209 vs. Mixed, Pav609 vs. Cg, Pav609 vs. CTRL, and TI vs. Cg. Based on the analyses, the following rankings are supported (Table 3.21A, B): Baseline > Cg, Cm, CTRL, and Mixed; ISO > Cg, Cm, CTRL, and Mixed; Pav609 > Cg and CTRL; and finally, TI > Cg.

The assessment of Fe/Ca in the right valves (Table 3.22A, B) revealed significant differences between four diet pairs: Baseline vs. CTRL, Baseline vs. TI, ISO vs. CTRL, and Mixed vs. CTRL. The ratios of the shells from the Baseline, ISO, and Mixed treatments were higher than those from CTRL group, and the ratios from the Baseline higher than those from the TI treatment.



Figure 3.28: Left valve median Fe/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed		(P <	0.050)	
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing Median 25% 75%			
Baseline	14	1	8.01E-05	5.74E-05	7.70E-04
CTRL	61	1	4.42E-05	2.22E-05	6.41E-05
ТІ	54	7	5.00E-05	3.85E-05	7.05E-05
ISO	50	2	8.02E-05	5.55E-05	9.67E-05
Cg	58	3	3.32E-05	2.50E-05	4.29E-05
Cm	64	14	4.57E-05	2.84E-05	6.40E-05
Pav609	59	7	5.89E-05	4.75E-05	8.22E-05
Pav1209	52	5	6.66E-05	4.57E-05	9.26E-05
Mixed	53	12	4.18E-05	3.29E-05	5.67E-05
	H = 93.0	with 8 degrees of	freedom (P = <0.001)		

Table 3.21A: Results of analysis of variance of left valve Fe/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.21B: Results of pairwise comparisons of left valve Fe/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :					
Comparison	Diff of Ranks	Q	P<0.05		
Baseline vs. Cg	189.99	5.161	Yes		
Baseline vs. Cm	127.154	3.422	Yes		
Baseline vs. CTRL	136.821	3.747	Yes		
Baseline vs. Mixed	129.398	3.406	Yes		
ISO vs. Cg	177.836	7.543	Yes		
ISO vs. Cm	115	4.768	Yes		
ISO vs. Ctrl	124.667	5.393	Yes		
ISO vs. Mixed	117.244	4.619	Yes		
Pav1209 vs. Cg	144.858	6.109	Yes		
Pav1209 vs. Cm	82.021	3.382	Yes		
Pav1209 vs. CTRL	91.688	3.943	Yes		
Pav1209 vs. Mixed	84.265	3.303	Yes		
Pav609 vs. Cg	130.375	5.647	Yes		
Pav609 vs. CTRL	77.205	3.414	Yes		
TI vs. Cg	87.347	3.684	Yes		
Baseline vs. ISO	12.154	0.326	No		
Baseline vs. Pav1209	45.133	1.207	No		
Baseline vs. Pav609	59.615	1.611	No		
Baseline vs. TI	102.643	2.744	No		

Comparison	Diff of Ranks	Q	P<0.05
Cm vs. Cg	62.836	2.694	No
Cm vs. CTRL	9.667	0.423	No
Cm vs. Mixed	2.244	0.0892	No
Ctrl vs. Cg	53.17	2.386	No
ISO vs. Pav1209	32.979	1.346	No
ISO vs. Pav609	47.462	1.986	No
ISO vs. TI	90.489	3.694	No
Mixed vs. Cg	60.592	2.46	No
Mixed vs. CTRL	7.423	0.307	No
Pav1209 vs. Pav609	14.483	0.603	No
Pav1209 vs. TI	57.511	2.336	No
Pav609 vs. Cm	67.538	2.857	No
Pav609 vs. Mixed	69.782	2.799	No
Pav609 vs. TI	43.028	1.791	No
TI vs. Cm	24.511	1.011	No
TI vs. Ctrl	34.177	1.47	No
TI vs. Mixed	26.755	1.049	No

Table 3.21B (Continued)

Table 3.22A: Results of analysis of variance of right valve Fe/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
	Kruskal-Wal	lis One Way Analy	sis of Variance on Rai	nks	
Group	N	Missing	Median	25%	75%
Baseline	14	2	9.88E-05	5.51E-05	5.04E-04
CTRL	29	1	2.10E-05	1.67E-05	3.46E-05
ТІ	17	7	2.49E-05	1.86E-05	4.84E-05
ISO	10	1	7.83E-05	3.86E-05	1.01E-04
Cg	19	1	3.75E-05	2.89E-05	5.45E-05
Cm	27	9	3.73E-05	2.54E-05	6.75E-05
Pav609	19	9	3.18E-05	2.47E-05	5.65E-05
Pav1209	14	1	3.19E-05	2.34E-05	5.03E-05
Mixed	31	11	4.16E-05	2.97E-05	6.51E-05
H = 39.1 with 8 degrees of freedom (P = <0.001)					

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Baseline vs. CTRL	74.488	5.4	Yes	
Baseline vs. TI	65.117	3.804	Yes	
ISO vs. CTRL	57.683	3.765	Yes	
Mixed vs. CTRL	39.221	3.351	Yes	
Baseline vs. Cg	45.75	3.07	No	
Baseline vs. Cm	44.194	2.966	No	
Baseline vs. ISO	16.806	0.953	No	
Baseline vs. Mixed	35.267	2.416	No	
Baseline vs. Pav1209	51.532	3.22	No	
Baseline vs. Pav609	52.317	3.056	No	
Cg vs. CTRL	28.738	2.379	No	
Cg vs. Pav1209	5.782	0.397	No	
Cg vs. Pav609	6.567	0.416	No	
Cg vs. TI	19.367	1.228	No	
Cm vs. Cg	1.556	0.117	No	
Cm vs. CTRL	30.294	2.508	No	
Cm vs. Pav1209	7.338	0.504	No	
Cm vs. Pav609	8.122	0.515	No	
Cm vs. TI	20.922	1.327	No	
ISO vs. Cg	28.944	1.773	No	
ISO vs. Cm	27.389	1.678	No	
ISO vs. Mixed	18.461	1.15	No	
ISO vs. Pav1209	34.726	2.003	No	
ISO vs. Pav609	35.511	1.933	No	
ISO vs. TI	48.311	2.63	No	
Mixed vs. Cg	10.483	0.807	No	
Mixed vs. Cm	8.928	0.687	No	
Mixed vs. Pav1209	16.265	1.142	No	
Mixed vs. Pav609	17.05	1.101	No	
Mixed vs. TI	29.85	1.928	No	
Pav1209 vs. CTRL	22.956	1.711	No	
Pav1209 vs. Pav609	0.785	0.0467	No	
Pav1209 vs. TI	13.585	0.808	No	
Pav609 vs. CTRL	22.171	1.505	No	
Pav609 vs. TI	12.8	0.716	No	
TI vs. CTRL	9.371	0.636	No	

Table 3.22B: Results of pairwise comparisons of right valve Fe/Ca between diets. Diets are represented as previously identified in Figure 3.25.

Comparison of the diet groups with regard to K/Ca in the left valves (Figure 3.29) initially revealed significant differences (P=0.011) within the dataset (Table 3.23A); however, no pairwise comparisons revealed significant differences between the individual diets (Table 3.23B). One comparison, CTRL vs. Baseline, did result in a Difference of Ranks of 111 and associated Q value of 3.04, which was assessed further and the labeled insignificance regarded as questionable. Comparison of diets with regard to K/Ca in the collected right valves did result in two significant pairwise comparisons (Tables 3.24A,B): Baseline > Cg and Pav1209 > Cg.



Figure 3.29: Left valve median K/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance						
Normality Test:	Failed		(P <	0.050)		
Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	N	N Missing Median 25% 75%				
Baseline	14	1	3.40E-03	3.72E-06	4.72E-03	
CTRL	61	1	2.38E-04	2.25E-05	1.49E-03	
TI	54	13	4.80E-04	3.18E-04	8.12E-04	
ISO	50	1	5.78E-04	2.83E-04	8.34E-04	
Cg	58	4	5.34E-04	2.85E-04	8.46E-04	
Cm	64	7	7.20E-04	3.61E-04	1.27E-03	
Pav609	59	5	6.84E-04	4.40E-04	9.13E-04	
Pav1209	52	5	4.58E-04	3.33E-04	1.21E-03	
Mixed	53	15	1.00E-03	4.29E-04	1.26E-03	
H = 19.9 with 8 degrees of freedom (P = 0.011)						

Table 3.23A: Results of analysis of variance of left valve K/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.23B: Results of pairwise comparisons of left valve K/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Baseline vs. Cm	45.128	1.23	No	
Baseline vs. Pav609	55.499	1.505	No	
Baseline vs. Pav1209	69.951	1.87	No	
Baseline vs. Mixed	42.462	1.107	No	
Baseline vs. CTRL	111.162	3.044	No	
Baseline vs. TI	92.535	2.435	No	
Baseline vs. ISO	76.686	2.059	No	
Baseline vs. Cg	89.295	2.421	No	
Cm vs. Pav609	10.37	0.457	No	
Cm vs. Pav1209	24.823	1.055	No	
Cm vs. CTRL	66.033	2.991	No	
Cm vs. TI	47.407	1.939	No	
Cm vs. ISO	31.558	1.357	No	
Cm vs. Cg	44.167	1.948	No	
Pav609 vs. Pav1209	14.452	0.607	No	
Pav609 vs. CTRL	55.663	2.486	No	
Pav609 vs. TI	37.036	1.498	No	
Pav609 vs. ISO	21.187	0.9	No	
Pav609 vs. Cg	33.796	1.471	No	
Pav1209 vs. CTRL	41.211	1.772	No	

Comparison	Diff of Ranks	Q	P<0.05
Pav1209 vs. TI	22.584	0.885	No
Pav1209 vs. ISO	6.735	0.276	No
Pav1209 vs. Cg	19.344	0.812	No
Mixed vs. Cm	2.667	0.107	No
Mixed vs. Pav609	13.037	0.516	No
Mixed vs. Pav1209	27.489	1.056	No
Mixed vs. CTRL	68.7	2.776	No
Mixed vs. TI	50.073	1.863	No
Mixed vs. ISO	34.224	1.326	No
Mixed vs. Cg	46.833	1.853	No
TI vs. CTRL	18.627	0.77	No
ISO vs. CTRL	34.476	1.5	No
ISO vs. TI	15.849	0.627	No
ISO vs. Cg	12.609	0.535	No
Cg vs. CTRL	21.867	0.977	No
Cg vs. TI	3.24	0.131	No

Table 3. 23B (Continued)

Table 3.24A: Results of analysis of variance of right valve K/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed		(P <	0.050)	
	Kruskal-Wal	llis One Way Analy	sis of Variance on Ra	nks	
Group	N	Missing	Median	25%	75%
Baseline	14	2	3.57E-03	1.32E-03	4.85E-03
Ctrl	29	2	1.42E-03	1.87E-04	2.41E-03
ТІ	17	7	1.02E-03	4.79E-04	1.44E-03
ISO	10	2	1.35E-03	9.65E-04	1.54E-03
Cg	19	1	3.35E-04	1.88E-04	1.31E-03
Cm	27	6	7.16E-04	4.16E-04	2.04E-03
Pav(609)	19	6	1.59E-03	8.92E-04	2.03E-03
Pav(1209)	14	1	2.17E-03	1.36E-03	4.01E-03
Mixed	31	10	1.26E-03	8.67E-04	1.52E-03
H = 23.3 with 8 degrees of freedom (P = 0.003)					

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Baseline vs. Cg	51.278	3.322	Yes	
Pav(1209) vs. Cg	59.406	3.94	Yes	
Baseline vs. Cm	33.143	2.211	No	
Baseline vs. CTRL	31.148	2.167	No	
Baseline vs. ISO	26.333	1.393	No	
Baseline vs. Mixed	22.429	1.496	No	
Baseline vs. Pav(609)	15.026	0.906	No	
Baseline vs. TI	39.133	2.206	No	
Cm vs. Cg	18.135	1.363	No	
Cm vs. TI	5.99	0.376	No	
Ctrl vs. Cg	20.13	1.597	No	
Ctrl vs. Cm	1.995	0.165	No	
Ctrl vs. TI	7.985	0.521	No	
ISO vs. Cg	24.944	1.417	No	
ISO vs. Cm	6.81	0.396	No	
ISO vs. CTRL	4.815	0.289	No	
ISO vs. TI	12.8	0.651	No	
Mixed vs. Cg	28.849	2.168	No	
Mixed vs. Cm	10.714	0.838	No	
Mixed vs. CTRL	8.72	0.723	No	
Mixed vs. ISO	3.905	0.227	No	
Mixed vs. TI	16.705	1.05	No	
Pav1209 vs. Baseline	8.128	0.49	No	
Pav1209 vs. Cm	41.271	2.823	No	
Pav1209 vs. Ctrl	39.276	2.809	No	
Pav1209 vs. ISO	34.462	1.851	No	
Pav1209 vs. Mixed	30.557	2.09	No	
Pav1209 vs. Pav(609)	23.154	1.425	No	
Pav1209 vs. TI	47.262	2.712	No	
Pav609) vs. Cg	36.252	2.404	No	
Pav609 vs. Cm	18.117	1.239	No	
Pav609 vs. Ctrl	16.123	1.153	No	
Pav609 vs. ISO	11.308	0.607	No	
Pav609 vs. Mixed	7.403	0.506	No	
Pav609 vs. TI	24.108	1.384	No	
TI vs. Cg	12.144	0.743	No	

Table 3.24B: Results of pairwise comparisons of right valve K/Ca between diets. Diets are represented as previously identified in Figure 3.25.

Figure 3.30 illustrates the left valve median Li/Ca values by diet. Statistical comparison of the diet groups suggests significant differences between 11 of the groups (Tables 3.25A, B): Baseline vs. Cg, Baseline vs. Cm, Baseline vs. CTRL, Baseline vs. ISO, Baseline vs. Mixed, Baseline vs. Pav1209, Baseline vs. Pav609, TI vs. Cg, TI vs. Cm, TI vs. ISO, and TI vs. Pav1209. The determined relationships are as follows: Baseline> Cg, Cm, CTRL, ISO, Mixed, Pav1209, Pav609; TI > Cg, Cm, ISO, and Pav1209.

The statistical comparison of the diet groups with regard to right valve Li/Ca (Tables 3.26A, B) identified six significant differences: Baseline vs. Cg, Baseline vs. Cm, Baseline vs. CTRL, Baseline vs. Mixed, Baseline vs. Pav1209, Tl vs. Cg. The relationships established are as follows: Baseline > Cg, Cm, CTRL, Mixed, Pav1209 and Tl> Cg.



Figure 3.30: Left valve median Li/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.24.

One Way Analysis of Variance					
Normality Test:	Failed		(P <	0.050)	
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	N Missing Median 25% 75%			
Baseline	14	1	4.03E-06	3.86E-06	9.15E-04
Ctrl	61	1	2.86E-06	2.20E-06	3.30E-06
ТІ	54	12	3.24E-06	2.84E-06	4.19E-06
ISO	50	1	2.61E-06	1.94E-06	3.06E-06
Cg	58	5	2.71E-06	1.95E-06	3.25E-06
Cm	64	8	2.82E-06	2.08E-06	3.32E-06
Pav(609)	59	6	2.95E-06	1.87E-06	3.89E-06
Pav(1209)	52	5	2.68E-06	2.34E-06	3.27E-06
Mixed	53	14	2.78E-06	2.45E-06	3.13E-06
H = 48.1 with 8 degrees of freedom (P = <0.001)					

Table 3.25A: Results of analysis of variance of left valve Li/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.25B: Results of pairwise comparisons of left valve Li/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Baseline vs. Cg	183.868	4.989	Yes	
Baseline vs. Cm	160.536	4.379	Yes	
Baseline vs. CTRL	154.383	4.238	Yes	
Baseline vs. ISO	196.204	5.281	Yes	
Baseline vs. Mixed	157.692	4.135	Yes	
Baseline vs. Pav1209	171.617	4.599	Yes	
Baseline vs. Pav609	158.566	4.302	Yes	
TI vs. Cg	102.392	4.162	Yes	
TI vs. Cm	79.06	3.253	Yes	
TI vs. ISO	114.728	4.582	Yes	
TI vs. Pav1209	90.141	3.565	Yes	
Baseline vs. TI	81.476	2.156	No	
Cg vs. ISO	12.336	0.523	No	
Cm vs. Cg	23.332	1.022	No	
Cm vs. ISO	35.668	1.531	No	
Cm vs. Pav1209	11.081	0.47	No	
CTRL vs. Cg	29.485	1.314	No	
CTRL vs. Cm	6.152	0.278	No	
CTRL vs. ISO	41.821	1.824	No	
CTRL vs. Mixed	3.309	0.135	No	

Table 3.25B (Continued)			
Comparison	Diff of Ranks	Q	P<0.05
CTRL vs. Pav1209	17.234	0.743	No
CTRL vs. Pav609	4.183	0.186	No
Mixed vs. Cg	26.176	1.042	No
Mixed vs. Cm	2.843	0.114	No
Mixed vs. ISO	38.512	1.507	No
Mixed vs. Pav1209	13.925	0.54	No
Mixed vs. Pav609	0.874	0.0348	No
Pav1209 vs. Cg	12.251	0.513	No
Pav1209 vs. ISO	24.587	1.011	No
Pav609 vs. Cg	25.302	1.094	No
Pav609 vs. Cm	1.97	0.0863	No
Pav609 vs. ISO	37.638	1.595	No
Pav609 vs. Pav1209	13.051	0.547	No
TI vs. CTRL	72.907	3.043	No
TI vs. Mixed	76.216	2.878	No
TI vs. Pav609	77.09	3.134	No

Table 3.26A: Results of analysis of variance of right valve Li/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
Baseline	14	2	3.96E-06	3.80E-06	4.71E-04
CTRL	29	2	2.88E-06	2.33E-06	3.68E-06
ТІ	17	7	5.72E-06	2.79E-06	7.15E-06
ISO	10	2	3.09E-06	3.02E-06	3.65E-06
Cg	19	1	2.40E-06	1.16E-06	3.11E-06
Cm	27	9	2.62E-06	2.55E-06	3.03E-06
Pav609	19	7	2.97E-06	2.56E-06	3.21E-06
Pav1209	14	1	2.79E-06	2.47E-06	3.26E-06
Mixed	31	13	2.76E-06	2.50E-06	2.83E-06
	H = 41.0	with 8 degrees of	freedom (P = <0.001)		

All Pairwise Multiple Com	parison Procedures (Dunn's Meth	od) :	
Comparison	Diff of Ranks	Q	P<0.05
Baseline vs. Cg	74.306	5.06	Yes
Baseline vs. Cm	64.25	4.375	Yes
Baseline vs. CTRL	50.398	3.687	Yes
Baseline vs. Mixed	67.528	4.598	Yes
Baseline vs. Pav1209	57.122	3.621	Yes
TI vs. Cg	54.622	3.515	Yes
Baseline vs. ISO	27.708	1.541	No
Baseline vs. Pav609	48.333	3.005	No
Baseline vs. TI	19.683	1.167	No
Cm vs. Cg	10.056	0.766	No
Cm vs. Mixed	3.278	0.25	No
CTRL vs. Cg	23.907	1.994	No
CTRL vs. Cm	13.852	1.155	No
CTRL vs. Mixed	17.13	1.429	No
CTRL vs. Pav1209	6.724	0.505	No
ISO vs. Cg	46.597	2.783	No
ISO vs. Cm	36.542	2.182	No
ISO vs. Ctrl	22.69	1.43	No
ISO vs. Mixed	39.819	2.378	No
ISO vs. Pav1209	29.413	1.661	No
ISO vs. Pav609	20.625	1.147	No
Mixed vs. Cg	6.778	0.516	No
Pav1209 vs. Cg	17.184	1.198	No
Pav1209 vs. Cm	7.128	0.497	No
Pav1209 vs. Mixed	10.406	0.726	No
Pav609 vs. Cg	25.972	1.769	No
Pav609 vs. Cm	15.917	1.084	No
Pav609 vs. CTRL	2.065	0.151	No
Pav609 vs. Mixed	19.194	1.307	No
Pav609 vs. Pav1209	8.788	0.557	No
TI vs. Cm	44.567	2.868	No
TI vs. CTRL	30.715	2.106	No
TI vs. ISO	8.025	0.429	No
TI vs. Mixed	47.844	3.079	No
TI vs. Pav1209	37.438	2.259	No
TI vs. Pav609	28.65	1.698	No

Table 3.26B: Results of pairwise comparisons of right valve Li/Ca between diets. Diets are represented as previously identified in Figure 3.25.

Comparison of left valve Mg/Ca between experimental diets (Tables 3.27A, B) revealed no significant differences, probably as a consequence of high variability (Figure 3.31). Comparison of right valve Mg/Ca with regard to diet (Tables 3.28A, B) revealed significant differences between three of the diets: CTRL < Baseline, CTRL < Cg, and CTRL < Mixed.



Figure 3.31: Left valve median Mg/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

Table 3.27A: Results of analysis of variance of left valve Mg/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
Baseline	12	2	1.29E-03	1.13E-03	1.43E-03
CTRL	61	1	7.14E-04	5.33E-04	1.80E-03
ТІ	54	6	9.57E-04	8.38E-04	1.16E-03
ISO	50	1	9.50E-04	7.58E-04	1.17E-03

Table 3.27A (Continued)

Group	Ν	Missing	Median	25%	75%
Cg	58	3	1.02E-03	6.92E-04	1.20E-03
Cm	64	7	1.08E-03	8.40E-04	1.57E-03
Pav609	59	6	9.20E-04	7.60E-04	1.07E-03
Pav1209	52	3	9.41E-04	7.54E-04	1.15E-03
Mixed	53	12	1.13E-03	8.10E-04	1.40E-03
H = 20.0 with 8 degrees of freedom (P = 0.010)					

Table 3.27B: Results of pairwise comparisons of left valve Mg/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :						
Comparison	Diff of Ranks	Q	P<0.05			
Baseline vs. Cg	89.582	2.137	No			
Baseline vs. Cm	34.109	0.816	No			
Baseline vs. CTRL	105.983	2.544	No			
Baseline vs. ISO	90.294	2.134	No			
Baseline vs. Mixed	44.21	1.028	No			
Baseline vs. Pav1209	86.784	2.051	No			
Baseline vs. Pav609	94.826	2.255	No			
Baseline vs. Tl	78.975	1.863	No			
Cg vs. CTRL	16.402	0.72	No			
Cg vs. ISO	0.712	0.0297	No			
Cg vs. Pav609	5.245	0.223	No			
Cm vs. Cg	55.473	2.406	No			
Cm vs. CTRL	71.875	3.186	No			
Cm vs. ISO	56.185	2.365	No			
Cm vs. Mixed	10.101	0.404	No			
Cm vs. Pav1209	52.675	2.217	No			
Cm vs. Pav609	60.718	2.609	No			
Cm vs. TI	44.866	1.878	No			
ISO vs. CTRL	15.689	0.668	No			
ISO vs. Pav609	4.533	0.188	No			
Mixed vs. Cg	45.372	1.803	No			
Mixed vs. CTRL	61.774	2.5	No			
Mixed vs. ISO	46.084	1.785	No			
Mixed vs. Pav1209	42.574	1.649	No			
Mixed vs. Pav609	50.617	1.995	No			
Mixed vs. TI	34.765	1.34	No			
Pav1209 vs. Cg	2.798	0.117	No			
Pav1209 vs. CTRL	19.2	0.818	No			

Table 3.27 D (Continued	Table 3.27E	3 (Continued)
-------------------------	-------------	---------------

Comparison	Diff of Ranks	Q	P<0.05
Pav1209 vs. ISO	3.51	0.142	No
Pav1209 vs. Pav609	8.043	0.333	No
Pav609 vs. CTRL	11.157	0.485	No
TI vs. Cg	10.607	0.44	No
TI vs. CTRL	27.008	1.144	No
TI vs. ISO	11.319	0.457	No
TI vs. Pav1209	7.809	0.315	No
TI vs. Pav609	15.851	0.652	No

Table 3.28A: Results of analysis of variance of right valve Mg/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
Baseline	14	4	1.30E-03	1.14E-03	1.47E-03
CTRL	29	1	1.90E-03	1.69E-03	2.71E-03
ТІ	17	4	1.69E-03	1.30E-03	2.26E-03
ISO	10	2	1.54E-03	1.30E-03	1.78E-03
Cg	19	1	1.30E-03	1.07E-03	1.58E-03
Cm	27	6	1.72E-03	1.41E-03	2.13E-03
Pav609	19	6	1.50E-03	1.24E-03	1.75E-03
Pav1209	14	1	1.58E-03	1.23E-03	1.68E-03
Mixed	31	10	1.26E-03	1.15E-03	1.52E-03
	H = 27 5	with 8 degrees of	freedom (P = < 0.001)		

Table 3.28B: Results of pairwise comparisons of right valve Mg/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :						
Comparison	Diff of Ranks	Q	P<0.05			
CTRL vs. Baseline	51.864	3.352	Yes			
CTRL vs. Cg	46.575	3.67	Yes			
CTRL vs. Mixed	48.036	3.962	Yes			
Cg vs. Baseline	5.289	0.319	No			
Cg vs. Mixed	1.46	0.108	No			
Cm vs. Baseline	37.543	2.326	No			
Cm vs. Cg	32.254	2.391	No			
Cm vs. ISO	10.893	0.624	No			
Cm vs. Mixed	33.714	2.601	No			

Comparison	Diff of Ranks	Q	P<0.05
Cm vs. Pav1209	15.297	1.032	No
Cm vs. Pav609	16.374	1.105	No
CTRL vs. Cm	14.321	1.181	No
CTRL vs. ISO	25.214	1.497	No
CTRL vs. Pav1209	29.618	2.101	No
CTRL vs. Pav609	30.695	2.177	No
CTRL vs. TI	14.234	1.01	No
ISO vs. Baseline	26.65	1.338	No
ISO vs. Cg	21.361	1.197	No
ISO vs. Mixed	22.821	1.308	No
ISO vs. Pav1209	4.404	0.233	No
ISO vs. Pav609	5.481	0.29	No
Mixed vs. Baseline	3.829	0.237	No
Pav1209 vs. Baseline	22.246	1.259	No
Pav1209 vs. Cg	16.957	1.109	No
Pav1209 vs. Mixed	18.418	1.243	No
Pav1209 vs. Pav(609)	1.077	0.0654	No
Pav609 vs. Baseline	21.169	1.198	No
Pav609 vs. Cg	15.88	1.039	No
Pav609 vs. Mixed	17.341	1.17	No
TI vs. Baseline	37.631	2.13	No
TI vs. Cg	32.342	2.116	No
TI vs. Cm	0.0879	0.00593	No
TI vs. ISO	10.981	0.582	No
TI vs. Mixed	33.802	2.28	No
TI vs. Pav1209	15.385	0.934	No
TI vs. Pav609	16.462	0.999	No

Table 3.28B (Continued)

The Mn/Ca median values of the collected left valves for each diet group are depicted in Figure 3.32. Fifteen of the diet group comparisons were significant (Tables 3.29A, B) with regard to Mn/Ca associated with the collected left valves - Baseline vs. CTRL, Cg vs. CTRL, Cm vs. CTRL, ISO vs. Cg, ISO vs. Cm, ISO vs. CTRL, Mixed vs. Cg, Mixed vs. Ctrl, Pav1209 vs. Cg, Pav1209 vs. Ctrl, Pav609 vs. Cg, Pav609 vs. CTRL, TI vs. Cg, TI vs. Cm, TI vs. CTRL. The relationships determined are as follows:

CTRL< Baseline, Cg, Cm, ISO, Mixed, Pav1209, Pav609, TI; Cg< ISO, Mixed, Pav1209, Pav609, TI; Cm<ISO, TI.

Comparison of the right valve Mn/Ca between diet groups revealed 12 significant differences (Tables 3.30A, B): Baseline vs. Cg, Baseline vs. CTRL, Cm vs. Cg, Cm vs. CTRL, ISO vs. Cg, ISO vs. CTRL, Mixed vs. Cg, Mixed vs. CTRL, Mixed vs. Pav 609, Mixed vs. Pav1209, TI vs. Cg, TI vs. CTRL. The subsequent ranking of these diet comparisons is as follows: Cg < Baseline, Cm, ISO, Mixed, TI; CTRL < Baseline, Cm, ISO, Mixed, TI; Mixed > Pav609, Pav1209.



Figure 3.32: Left valve median Mn/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance						
Normality Test:	Failed	(P < 0.050)				
Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	N	Missing	Median	25%	75%	
Baseline	14	2	5.01E-05	3.18E-05	2.00E-04	
CTRL	61	1	1.74E-05	1.20E-05	2.21E-05	
ТІ	54	5	8.61E-05	6.26E-05	1.13E-04	
ISO	50	1	8.96E-05	6.89E-05	1.11E-04	
Cg	58	3	3.25E-05	2.71E-05	4.02E-05	
Cm	64	7	5.20E-05	4.13E-05	7.28E-05	
Pav609	59	6	6.48E-05	5.15E-05	7.64E-05	
Pav1209	52	4	6.80E-05	5.04E-05	8.40E-05	
Mixed	53	11	6.98E-05	5.68E-05	8.27E-05	
	H = 231 with 8 degrees of freedom (P = <0.001)					

Table 3.29A: Results of analysis of variance of left valve Mn/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.29B: Results of pairwise comparisons of left valve Mn/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :						
Comparison	Diff of Ranks	Q	P<0.05			
Baseline vs. CTRL	180.983	4.659	Yes			
Cg vs. CTRL	79.458	3.465	Yes			
Cm vs. CTRL	176.584	7.773	Yes			
ISO vs. Cg	199.64	8.274	Yes			
ISO vs. Cm	102.513	4.284	Yes			
ISO vs. CTRL	279.097	11.801	Yes			
Mixed vs. Cg	147.371	5.855	Yes			
Mixed vs. CTRL	226.829	9.179	Yes			
Pav1209 vs. Cg	130.213	5.367	Yes			
Pav1209 vs. CTRL	209.671	8.815	Yes			
Pav609 vs. Cg	125.581	5.312	Yes			
Pav609 vs. CTRL	205.038	8.855	Yes			
TI vs. Cg	186.354	7.723	Yes			
TI vs. Cm	89.227	3.729	Yes			
TI vs. Ctrl	265.812	11.239	Yes			
Baseline vs. Cg	101.526	2.594	No			
Baseline vs. Cm	4.399	0.113	No			
Cm vs. Cg	97.127	4.183	No			
ISO vs. Baseline	98.114	2.48	No			
ISO vs. Mixed	52.269	2.024	No			
Table 3.29B (Continued)						
-------------------------	---------------	-------	--------	--	--	
Comparison	Diff of Ranks	Q	P<0.05			
ISO vs. Pav1209	69.426	2.783	No			
ISO vs. Pav609	74.059	3.042	No			
ISO vs. TI	13.286	0.535	No			
Mixed vs. Baseline	45.845	1.14	No			
Mixed vs. Cm	50.244	2.012	No			
Mixed vs. Pav1209	17.158	0.661	No			
Mixed vs. Pav609	21.79	0.859	No			
Pav1209 vs. Baseline	28.688	0.724	No			
Pav1209 vs. Cm	33.087	1.375	No			
Pav1209 vs. Pav609	4.632	0.189	No			
Pav609 vs. Baseline	24.055	0.613	No			
Pav609 vs. Cm	28.454	1.214	No			
TI vs. Baseline	84.828	2.144	No			
TI vs. Mixed	38.983	1.509	No			
TI vs. Pav1209	56.141	2.251	No			
TI vs. Pav609	60.773	2.497	No			

Table 3.30A: Results of analysis of variance of right valve Mn/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
Baseline	14	2	5.43E-05	3.56E-05	2.32E-04
Ctrl	29	1	1.83E-05	1.23E-05	2.49E-05
ТІ	17	4	5.60E-05	4.94E-05	7.79E-05
ISO	10	1	6.15E-05	4.54E-05	6.95E-05
Cg	19	1	2.46E-05	1.97E-05	3.16E-05
Cm	27	8	5.04E-05	3.63E-05	6.00E-05
Pav (609)	19	7	3.80E-05	2.89E-05	4.73E-05
Pav(1209)	14	1	3.51E-05	2.76E-05	4.47E-05
Mixed	31	11	6.67E-05	6.22E-05	6.99E-05
	H = 91.5	with 8 degrees of	freedom (P = <0.001)		

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Baseline vs. Cg	54.694	3.518	Yes	
Baseline vs. CTRL	71.738	4.984	Yes	
Cm vs. Cg	48.83	3.559	Yes	
Cm vs. CTRL	65.874	5.313	Yes	
ISO vs. Cg	60.333	3.543	Yes	
ISO vs. Ctrl	77.377	4.841	Yes	
Mixed vs. Cg	74.878	5.525	Yes	
Mixed vs. CTRL	91.921	7.527	Yes	
Mixed vs. Pav609	49.433	3.245	Yes	
Mixed vs. Pav1209	50.638	3.407	Yes	
TI vs. Cg	67.701	4.459	Yes	
TI vs. Ctrl	84.745	6.053	Yes	
Baseline vs. Cm	5.864	0.381	No	
Baseline vs. Pav609	29.25	1.718	No	
Baseline vs. Pav1209	30.455	1.824	No	
Cg vs. CTRL	17.044	1.352	No	
Cm vs. Pav 609	23.386	1.52	No	
Cm vs. Pav1209	24.591	1.638	No	
ISO vs. Baseline	5.639	0.307	No	
ISO vs. Cm	11.503	0.681	No	
ISO vs. Pav609	34.889	1.897	No	
ISO vs. Pav1209	36.094	1.995	No	
Mixed vs. Baseline	20.183	1.325	No	
Mixed vs. Cm	26.047	1.949	No	
Mixed vs. ISO	14.544	0.869	No	
Mixed vs. TI	7.177	0.483	No	
Pav609 vs. Cg	25.444	1.637	No	
Pav609 vs. CTRL	42.488	2.952	No	
Pav609 vs. Pav1209	1.205	0.0722	No	
Pav1209 vs. Cg	24.239	1.597	No	
Pav1209 vs. CTRL	41.283	2.949	No	
TI vs. Baseline	13.006	0.779	No	
TI vs. Cm	18.87	1.257	No	
TI vs. ISO	7.368	0.407	No	
TI vs. Pav609	42.256	2.531	No	
TI vs. Pav1209	43.462	2.656	No	

Table 3.30B: Results of pairwise comparisons of right valve Mn/Ca between diets. Diets are represented as previously identified in Figure 3.25.

The median P/Ca values for the collected left valves and associated diet are depicted in Figure 3.33. No baseline data are available for this elemental ratio. Ten diet comparisons with regard to P/Ca were determined significant (Tables 3.31A, B): ISO vs. Cg, ISO vs. Cm, ISO vs. CTRL, ISO vs. Mixed, ISO vs. Pav1209, ISO vs. Pav609, ISO vs. TI, TI vs. Cg, TI vs. Cm, and TI vs. Mixed. The subsequent ranking of these diet groups is as follows: ISO> Cg, Cm, CTRL, Mixed, Pav1209, Pav609, TI and TI> Cg, Cm and Mixed.

Analysis of variance related to the right valve P/Ca per diet also identified significant differences within the dataset (P=0.006, Table 3.32A). The post hoc pairwise analysis, however, revealed no significant difference between any of the individual diet groups (Table 3.32B).



Figure 3.33: Left valve median P/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed	(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
CTRL	61	6	5.61E-04	3.02E-04	1.37E-03
ті	53	13	8.73E-04	6.10E-04	1.15E-03
ISO	50	7	1.15E-03	9.35E-04	2.09E-03
Cg	57	10	5.38E-04	3.62E-04	8.93E-04
Cm	62	18	5.49E-04	4.80E-04	6.66E-04
Pav609	58	18	5.83E-04	4.97E-04	7.82E-04
Pav1209	52	12	7.95E-04	5.63E-04	9.05E-04
Mixed	53	12	5.11E-04	4.41E-04	7.38E-04
	H = 81.7	with 7 degrees of	freedom (P = <0.001)		

Table 3.31A: Results of analysis of variance of left valve P/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.31B: Results of pairwise comparisons of left valve P/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
ISO vs. Cg	142.666	6.682	Yes	
ISO vs. Cm	150.024	6.915	Yes	
ISO vs. Ctrl	133.924	6.502	Yes	
ISO vs. Mixed	153.753	6.962	Yes	
ISO vs. Pav1209	96.195	4.328	Yes	
ISO vs. Pav609	133.42	6.003	Yes	
ISO vs. TI	72.22	3.249	Yes	
TI vs. Cg	70.446	3.237	Yes	
TI vs. Cm	77.805	3.52	Yes	
TI vs. Mixed	81.533	3.626	Yes	
Cg vs. Cm	7.359	0.347	No	
Cg vs. Mixed	11.087	0.513	No	
Cm vs. Mixed	3.728	0.17	No	
CTRL vs. Cg	8.741	0.435	No	
CTRL vs. Cm	16.1	0.787	No	
CTRL vs. Mixed	19.828	0.95	No	
Pav1209 vs. Cg	46.471	2.135	No	
Pav1209 vs. Cm	53.83	2.435	No	
Pav1209 vs. Ctrl	37.73	1.794	No	
Pav1209 vs. Mixed	57.558	2.56	No	
Pav1209 vs. Pav609	37.225	1.645	No	

Table 3.31B (Continued)

Comparison	Diff of Ranks	Q	P<0.05
Pav609 vs. Cg	9.246	0.425	No
Pav609 vs. Cm	16.605	0.751	No
Pav609 vs. Ctrl	0.505	0.024	No
Pav609 vs. Mixed	20.333	0.904	No
TI vs. Ctrl	61.705	2.935	No
TI vs. Pav1209	23.975	1.06	No
TI vs. Pav609	61.2	2.705	No

Table 3.32A: Results of analysis of variance of right valve P/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance						
Normality Test:	Failed		(P < 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	N	Missing	Median	25%	75%	
CTRL	26	1	8.23E-04	6.38E-04	1.39E-03	
ТІ	17	7	8.75E-04	4.76E-04	1.56E-03	
ISO	10	1	7.96E-04	6.94E-04	1.33E-03	
Cg	19	1	9.91E-04	6.06E-04	1.88E-03	
Cm	27	9	5.60E-04	4.65E-04	6.12E-04	
Pav609	19	9	6.51E-04	4.49E-04	8.70E-04	
Pav1209	14	1	9.23E-04	6.63E-04	1.10E-03	
Mixed	31	11	6.03E-04	4.66E-04	9.19E-04	
	H = 20.0) with 7 degrees of	freedom ($P = 0.006$)		•	

Table 3.32B: Results of pairwise comparisons of right valve P/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
CTRL vs. Cm	33.791	3.066	No	
CTRL vs. Mixed	24.73	2.312	No	
CTRL vs. Pav609	24.98	1.873	No	
CTRL vs. TI	4.48	0.336	No	
Cg vs. CTRL	3.042	0.276	No	
Cg vs. Cm	36.833	3.099	No	
Cg vs. ISO	0.278	0.0191	No	
Cg vs. Mixed	27.772	2.398	No	
Cg vs. Pav609	28.022	1.993	No	
Cg vs. Pav1209	2.722	0.21	No	

Comparison	Diff of Ranks	Q	P<0.05
Cg vs. TI	7.522	0.535	No
ISO vs. CTRL	2.764	0.199	No
ISO vs. Cm	36.556	2.512	No
ISO vs. Mixed	27.494	1.921	No
ISO vs. Pav609	27.744	1.694	No
ISO vs. Pav1209	2.444	0.158	No
ISO vs. TI	7.244	0.442	No
Mixed vs. Cm	9.061	0.782	No
Mixed vs. Pav609	0.25	0.0181	No
Pav609 vs. Cm	8.811	0.627	No
Pav1209 vs. CTRL	0.32	0.0262	No
Pav1209 vs. Cm	34.111	2.629	No
Pav1209 vs. Mixed	25.05	1.972	No
Pav1209 vs. Pav609	25.3	1.687	No
Pav1209 vs. TI	4.8	0.32	No
TI vs. Cm	29.311	2.085	No
TI vs. Mixed	20.25	1.467	No
TI vs. Pav609	20.5	1.286	No

Table 3.32B (Continued)

The median Si/Ca values by diet for the left valves collected are depicted in Figure 3.34. Significant differences were revealed in ten diet group comparisons with regard to left valve Si/Ca (Tables 3.33A, B), and the following ranks established: Mixed < Baseline, CTRL, ISO, Pav1209, Pav609, TI; Pav1209 > Cg, Cm; Pav609 > Cg, Cm.

The comparisons of right valves collected with regard to Si/Ca revealed significant differences in nine comparisons (Tables 3.34A, B): Baseline vs. Cg, Baseline vs. Cg, Baseline vs. Cm, Baseline vs. Mixed, Baseline vs. Pav609, Baseline vs. Pav1209, Ctrl vs. Cg, CTRL vs. Mixed, CTRL vs. Pav609, and CTRL vs. Pav1209. Based on these assessments, the subsequent rankings of the diet groups are as follows: Baseline > Cg, Cm, Mixed, Pav609, Pav1209 and CTRL > Cg, Mixed, Pav609, and Pav1209.



Figure 3.34: Left valve median Si/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed		(P <	0.050)	
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
Baseline	10	1	4.17E-04	2.93E-04	5.26E-04
CTRL	61	1	3.13E-04	1.66E-04	5.09E-04
ТІ	54	7	3.64E-04	2.28E-04	7.26E-04
ISO	50	1	3.21E-04	2.13E-04	4.98E-04
Cg	58	3	1.68E-04	1.24E-04	2.36E-04
Cm	64	8	1.64E-04	9.96E-05	2.69E-04
Pav609	59	7	6.27E-04	4.74E-04	7.29E-04
Pav1209	52	4	6.22E-04	4.17E-04	8.29E-04
Mixed	53	12	1.23E-04	9.46E-05	1.95E-04
	H = 109	with 8 degrees of f	reedom (P = <0.001)		

Table 3.33A: Results of analysis of variance of left valve Si/Ca among diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Baseline vs. Mixed	146.965	3.313	Yes	
CTRL vs. Mixed	113.37	4.642	Yes	
ISO vs. Mixed	111.609	4.375	Yes	
Pav1209 vs. Cg	149.294	6.271	Yes	
Pav1209 vs. Cm	137.324	5.793	Yes	
Pav1209 vs. Mixed	171.874	6.706	Yes	
Pav609 vs. Cg	148.677	6.378	Yes	
Pav609 vs. Cm	136.707	5.89	Yes	
Pav609 vs. Mixed	171.258	6.804	Yes	
TI vs. Mixed	137.492	5.338	Yes	
Baseline vs. Cg	124.384	2.87	No	
Baseline vs. Cm	112.415	2.597	No	
Baseline vs. CTRL	33.594	0.78	No	
Baseline vs. ISO	35.356	0.809	No	
Baseline vs. TI	9.473	0.216	No	
Cg vs. Mixed	22.581	0.908	No	
Cm vs. Cg	11.969	0.523	No	
Cm vs. Mixed	34.55	1.395	No	
CTRL vs. Cg	90.789	4.035	No	
CTRL vs. Cm	78.82	3.52	No	
CTRL vs. ISO	1.762	0.0759	No	
ISO vs. Cg	89.028	3.76	No	
ISO vs. Cm	77.059	3.269	No	
Pav1209 vs. Baseline	24.91	0.569	No	
Pav1209 vs. CTRL	58.504	2.507	No	
Pav1209 vs. ISO	60.266	2.462	No	
Pav1209 vs. Pav609	0.617	0.0256	No	
Pav1209 vs. Tl	34.383	1.39	No	
Pav609 vs. Baseline	24.293	0.558	No	
Pav609 vs. CTRL	57.887	2.535	No	
Pav609 vs. ISO	59.649	2.486	No	
Pav609 vs. TI	33.766	1.392	No	
TI vs. Cg	114.911	4.8	No	
TI vs. Cm	102.942	4.318	No	
TI vs. CTRL	24.122	1.027	No	
TI vs. ISO	25.883	1.052	No	

Table 3.33B: Results of pairwise comparisons of left valve Si/Ca between diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance					
Normality Test:	Failed		(P <	0.050)	
Kruskal-Wallis One Way Analysis of Variance on Ranks					
Group	N	Missing	Median	25%	75%
Baseline	10	1	3.68E-04	2.94E-04	5.58E-04
CTRL	29	1	2.03E-04	1.76E-04	3.10E-04
ТІ	17	8	1.50E-04	1.11E-04	1.90E-04
ISO	10	1	1.40E-04	1.26E-04	2.56E-04
Cg	19	1	7.78E-05	5.92E-05	1.72E-04
Cm	27	9	8.49E-05	5.32E-05	2.90E-04
Pav 609	19	9	3.34E-05	9.35E-06	1.18E-04
Pav1209	14	1	3.39E-05	2.02E-05	1.05E-04
Mixed	31	11	1.06E-04	7.17E-05	1.32E-04
	H = 46.9	with 8 degrees of f	freedom (P = <0.001)		

Table 3.34A: Results of analysis of variance of right valve Si/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.34B: Results of pairwise comparisons of right valve Si/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :							
Comparison	Diff of Ranks	Q	P<0.05				
Baseline vs. Cg	59.833	3.775	Yes				
Baseline vs. Cm	54.778	3.456	Yes				
Baseline vs. Mixed	58.456	3.751	Yes				
Baseline vs. Pav609	77.056	4.319	Yes				
Baseline vs. Pav1209	79.786	4.739	Yes				
CTRL vs. Cg	39.385	3.358	Yes				
CTRL vs. Mixed	38.007	3.344	Yes				
CTRL vs. Pav609	56.607	3.958	Yes				
CTRL vs. Pav1209	59.338	4.554	Yes				
Baseline vs. CTRL	20.448	1.374	No				
Baseline vs. ISO	36.778	2.009	No				
Baseline vs. TI	40.222	2.198	No				
Cg vs. Pav609	17.222	1.125	No				
Cg vs. Pav1209	19.953	1.412	No				
Cm vs. Cg	5.056	0.391	No				
Cm vs. Mixed	3.678	0.292	No				
Cm vs. Pav609	22.278	1.455	No				
Cm vs. Pav1209	25.009	1.77	No				
CTRL vs. Cm	34.329	2.927	No				
CTRL vs. ISO	16.329	1.098	No				

Table 3.34B (Continued)							
Comparison	Diff of Ranks	Q	P<0.05				
CTRL vs. TI	19.774	1.329	No				
ISO vs. Cg	23.056	1.455	No				
ISO vs. Cm	18	1.136	No				
ISO vs. Mixed	21.678	1.391	No				
ISO vs. Pav609	40.278	2.258	No				
ISO vs. Pav1209	43.009	2.555	No				
ISO vs. TI	3.444	0.188	No				
Mixed vs. Cg	1.378	0.109	No				
Mixed vs. Pav609	18.6	1.237	No				
Mixed vs. Pav1209	21.331	1.542	No				
Pav 609 vs. Pav1209	2.731	0.167	No				
TI vs. Cg	19.611	1.237	No				
TI vs. Cm	14.556	0.918	No				
TI vs. Mixed	18.233	1.17	No				
TI vs. Pav609	36.833	2.065	No				
TI vs. Pav1209	39.564	2.35	No				

Again there was no baseline data for the examination of Sr enrichment. With regard to comparison of Sr/Ca in the left valves (Figure 3.35), significant differences were revealed to exist between four diet groupings: Cm vs. ISO, Cm vs. Pav1209, Cm vs. Pav609, and Mixed vs. Pav1209. The subsequent ranking of these comparisons (Tables 35A, B) is as follows: Cm > ISO, Pav609, Pav1209 and Mixed > Pav1209. In terms of right valve comparisons of Sr/Ca by diet (Tables 36A, B), only the TI vs. CG comparison proved significant, and the TI group revealed to have a higher Sr/Ca than that from the Cg group.



Figure 3.35: Left valve median Sr/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance							
Normality Test:	Failed	(P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%		
CTRL	61	1	4.80E-03	4.48E-03	5.46E-03		
ТІ	54	5	5.20E-03	4.54E-03	6.30E-03		
ISO	50	1	4.77E-03	4.19E-03	5.84E-03		
Cg	58	3	4.99E-03	4.11E-03	6.50E-03		
Cm	64	7	5.73E-03	4.51E-03	7.48E-03		
Pav609	59	1	4.91E-03	4.34E-03	5.36E-03		
Pav1209	52	10	4.56E-03	4.06E-03	5.21E-03		
Mixed	53	6	5.41E-03	4.59E-03	6.68E-03		
	H = 28.9 with 7 degrees of freedom (P = <0.001)						

Table 3.35A: Results of analysis of variance of left valve Sr/Ca among diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :							
Comparison	Diff of Ranks	Q	P<0.05				
Cm vs. ISO	73.511	3.131	Yes				
Cm vs. Pav1209	102.218	4.171	Yes				
Cm vs. Pav609	71.485	3.18	Yes				
Mixed vs. Pav1209	90.414	3.533	Yes				
Cg vs. CTRL	12.385	0.55	No				
Cg vs. ISO	25.006	1.056	No				
Cg vs. Pav1209	53.713	2.175	No				
Cg vs. Pav609	22.98	1.013	No				
Cm vs. Cg	48.505	2.129	No				
Cm vs. CTRL	60.889	2.731	No				
Cm vs. Mixed	11.804	0.497	No				
Cm vs. TI	23.47	1	No				
CTRL vs. ISO	12.621	0.544	No				
CTRL vs. Pav1209	41.329	1.704	No				
CTRL vs. Pav609	10.595	0.477	No				
ISO vs. Pav1209	28.707	1.133	No				
Mixed vs. Cg	36.701	1.533	No				
Mixed vs. CTRL	49.086	2.091	No				
Mixed vs. ISO	61.707	2.508	No				
Mixed vs. Pav609	59.681	2.523	No				
Mixed vs. TI	11.666	0.474	No				
Pav609 vs. ISO	2.026	0.0866	No				
Pav609 vs. Pav1209	30.733	1.259	No				
TI vs. Cg	25.035	1.057	No				
TI vs. CTRL	37.42	1.612	No				
TI vs. ISO	50.041	2.055	No				
TI vs. Pav1209	78.748	3.107	No				
TI vs. Pav609	48.015	2.053	No				

Table 3.35B: Results of pairwise comparisons of left valve Sr/Ca between diets. Diets are represented as previously identified in Figure 3.25.

The final elemental ratio analyzed was Zn/Ca. In the comparisons of the left valves (Figure 3.36), only two significant differences between diets were revealed – ISO vs. Baseline and ISO vs. Cg. In both cases, the left valve Zn/Ca was lower in the ISO diet group (Tables 3.37A, B). In the comparisons of the right valves by diet with regard

to Zn/Ca (Tables 3.38A, B), four significant differences between diets were revealed and the following ranking established: Cg and CTRL > Mixed and Pav1209.

One Way Analysis of Variance							
Normality Test:	Failed	(P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing Median 25% 75%					
CTRL	29	1	4.89E-03	4.33E-03	5.86E-03		
ТІ	17	3	6.40E-03	5.05E-03	7.42E-03		
ISO	10	1	4.79E-03	4.25E-03	5.88E-03		
Cg	19	1	3.70E-03	3.24E-03	5.04E-03		
Cm	27	7	4.99E-03	4.57E-03	5.51E-03		
Pav (609)	19	2	4.95E-03	4.19E-03	5.28E-03		
Pav(1209)	14	11	5.12E-03	4.90E-03	5.69E-03		
Mixed	31	6	4.81E-03	4.38E-03	5.82E-03		
H = 21.0 with 7 degrees of freedom (P = 0.004)							

Table 3.36A: Results of analysis of variance of right valve Sr/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.36B	: Results of	of pairwise	comparisons	of right	valve	Sr/Ca	between	diets.	Diets a	are	represent	ted
as previously	y identified	I in Figure 3	3.25.									

All Pairwise Multiple Comparison Procedures (Dunn's Method) :							
Comparison	Diff of Ranks	Q	P<0.05				
TI vs. Cg	61.722	4.461	Yes				
CTRL vs. Cm	1.507	0.133	No				
CTRL vs. Cg	33.329	2.841	No				
CTRL vs. Mixed	6.167	0.577	No				
CTRL vs. Pav609	8.137	0.682	No				
CTRL vs. ISO	5.94	0.399	No				
Cm vs. Cg	31.822	2.523	No				
Cm vs. Mixed	4.66	0.4	No				
Cm vs. Pav609	6.629	0.518	No				
Cm vs. ISO	4.433	0.284	No				
TI vs. CTRL	28.393	2.234	No				
TI vs. Cm	29.9	2.21	No				
TI vs. Mixed	34.56	2.667	No				
TI vs. Pav609	36.529	2.607	No				
TI vs. Pav1209	19	0.769	No				
TI vs. ISO	34.333	2.07	No				

Table 3.36B (Continued)
---------------	------------

Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. Cg	27.162	2.263	No
Mixed vs. Pav609	1.969	0.161	No
Pav609 vs. Cg	25.193	1.919	No
Pav1209 vs. CTRL	9.393	0.398	No
Pav1209 vs. Cm	10.9	0.453	No
Pav1209 vs. Cg	42.722	1.764	No
Pav1209 vs. Mixed	15.56	0.656	No
Pav1209 vs. Pav609	17.529	0.721	No
Pav1209 vs. ISO	15.333	0.592	No
ISO vs. Cg	27.389	1.728	No
ISO vs. Mixed	0.227	0.015	No
ISO vs. Pav609	2.196	0.137	No



Figure 3.36: Left valve median Zn/Ca values compared to diet. Error bars depict the 95% confidence intervals and diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance							
Normality Test:	Failed	(P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%		
Baseline	14	2	2.00E-05	1.61E-05	2.80E-05		
CTRL	58	17	9.74E-06	3.23E-06	1.96E-05		
ТІ	35	21	1.28E-05	9.36E-06	2.58E-05		
ISO	25	11	5.27E-06	-2.54E-04	1.18E-05		
Cg	39	7	2.45E-05	4.28E-06	8.13E-05		
Cm	45	27	1.12E-05	7.03E-06	1.59E-05		
Pav609	39	17	6.96E-06	2.59E-06	1.98E-05		
Pav1209	20	5	1.07E-05	9.01E-06	2.02E-05		
Mixed	29	10	1.49E-05	9.21E-06	2.37E-05		
	H = 20.	2 with 8 degrees o	f freedom ($P = 0.010$))			

Table 3.37A: Results of analysis of variance of left valve Zn/Ca among diets. Diets are represented as previously identified in Figure 3.25.

Table 3.37B: Results of pairwise comparisons of left valve Zn/Ca between diets. Diets are represented as previously identified in Figure 3.25.

All Pairwise Multiple Comparison Procedures (Dunn's Method) :							
Comparison	Diff of Ranks	Q	P<0.05				
Baseline vs. ISO	72.357	3.398	Yes				
Cg vs. ISO	57.42	3.311	Yes				
Baseline vs. Cg	14.938	0.815	No				
Baseline vs. Cm	37.056	1.837	No				
Baseline vs. CTRL	46.378	2.611	No				
Baseline vs. Mixed	20.816	1.043	No				
Baseline vs. Pav1209	25.367	1.21	No				
Baseline vs. Pav609	44.136	2.272	No				
Baseline vs. TI	32.714	1.536	No				
Cg vs. Cm	22.118	1.387	No				
Cg vs. CTRL	31.441	2.463	No				
Cg vs. Mixed	5.878	0.375	No				
Cg vs. Pav1209	10.429	0.616	No				
Cg vs. Pav609	29.199	1.948	No				
Cg vs. TI	17.777	1.025	No				
Cm vs. CTRL	9.322	0.609	No				
Cm vs. ISO	35.302	1.83	No				
Cm vs. Pav609	7.081	0.412	No				
CTRL vs. ISO	25.979	1.551	No				
Mixed vs. Cm	16.24	0.912	No				

Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. CTRL	25.562	1.702	No
Mixed vs. ISO	51.541	2.704	No
Mixed vs. Pav1209	4.551	0.243	No
Mixed vs. Pav609	23.321	1.376	No
Mixed vs. TI	11.898	0.624	No
Pav1209 vs. Cm	11.689	0.618	No
Pav1209 vs. Ctrl	21.011	1.286	No
Pav1209 vs. ISO	46.99	2.336	No
Pav1209 vs. Pav609	18.77	1.036	No
Pav1209 vs. TI	7.348	0.365	No
Pav609 vs. CTRL	2.242	0.157	No
Pav609 vs. ISO	28.221	1.525	No
TI vs. Cm	4.341	0.225	No
TI vs. CTRL	13.664	0.816	No
TI vs. ISO	39.643	1.938	No
TI vs. Pav609	11.422	0.617	No

Table 3.37B (Continued)

Table 3.38A: Results of analysis of variance of right valve Zn/Ca among diets. Diets are represented as previously identified in Figure 3.25.

One Way Analysis of Variance														
Normality Test:	Normality Test: Failed (P < 0.050)													
Kruskal-Wallis One Way Analysis of Variance on Ranks														
Group N Missing Median 25% 75%														
Baseline	14	2	2.10E-05	1.41E-05	2.62E-05									
CTRL	27	1	3.35E-05	2.30E-05	4.63E-05									
ТІ	17	12	1.50E-05	1.25E-05	2.11E-05									
ISO	10	5	1.70E-05	1.30E-05	2.60E-05									
Cg	19	1	7.23E-05	3.03E-05	8.46E-05									
Cm	24	9	2.03E-05	1.79E-05	2.67E-05									
Pav609	19	9	2.43E-05	1.51E-05	6.42E-05									
Pav1209	14	1	1.32E-05	9.40E-06	2.02E-05									
Mixed	29	11	1.25E-05	9.75E-06	2.17E-05									
	H = 43.9	with 8 degrees of f	freedom (P = <0.001)											

All Pairwise Multiple Co	mparison Procedures (Dunn's M	ethod) :	
Comparison	Diff of Ranks	Q	P<0.05
Cg vs. Mixed	63.667	5.401	Yes
Cg vs. Pav1209	60.158	4.674	Yes
CTRL vs. Mixed	42.47	3.917	Yes
CTRL vs. Pav1209	38.962	3.244	Yes
Baseline vs. Cm	1.117	0.0815	No
Baseline vs. ISO	8.85	0.47	No
Baseline vs. Mixed	21.528	1.634	No
Baseline vs. Pav1209	18.019	1.273	No
Baseline vs. TI	14.45	0.768	No
Cg vs. Baseline	42.139	3.197	No
Cg vs. Cm	43.256	3.499	No
Cg vs. CTRL	21.197	1.955	No
Cg vs. ISO	50.989	2.852	No
Cg vs. Pav609	32.889	2.358	No
Cg vs. TI	56.589	3.166	No
Cm vs. ISO	7.733	0.423	No
Cm vs. Mixed	20.411	1.651	No
Cm vs. Pav1209	16.903	1.261	No
Cm vs. TI	13.333	0.73	No
CTRL vs. Baseline	20.942	1.697	No
CTRL vs. Cm	22.059	1.924	No
CTRL vs. ISO	29.792	1.725	No
CTRL vs. Pav609	11.692	0.889	No
CTRL vs. TI	35.392	2.05	No
ISO vs. Mixed	12.678	0.709	No
ISO vs. Pav1209	9.169	0.493	No
ISO vs. TI	5.6	0.25	No
Pav609 vs. Baseline	9.25	0.611	No
Pav609 vs. Cm	10.367	0.718	No
Pav609 vs. ISO	18.1	0.934	No
Pav609 vs. Mixed	30.778	2.207	No
Pav609 vs. Pav1209	27.269	1.833	No
Pav609 vs. TI	23.7	1.224	No
Pav1209 vs. Mixed	3.509	0.273	No
TI vs. Mixed	7.078	0.396	No
TI vs. Pav1209	3.569	0.192	No

Table 3.38B: Results of pairwise comparisons of right valve Zn/Ca between diets. Diets are represented as previously identified in Figure 3.25.

Correlation analysis: time, growth, and element association

Because the dataset being analyzed was shown to have different degrees of normality dependent upon the variables being examined, two separate correlation analyses were conducted: 1) Pearson Product Moment Correlation and 2) Spearman' Correlation. These tests were used to further examine the association of elemental composition (element/Ca ratios) with 1) time (month of collection), 2) growth (shell length and mass), and 3) other elements present.

Table 3.39A provides the Correlation Coefficients, associated P-value, and Sample size (n) for each correlation determined significant with regard to element ratio association with time, shell length and valve mass using the Pearson's correlation procedure. Ranking of the strength of association was based on weak <0.3, moderate 0.3-0.7, and strong >0.7. The Ba/Ca, Co/Ca, K/Ca, Li/Ca, Mg/Ca, and Zn/Ca ratios all showed a weak to moderate association with time with the only positive correlation revealed for Co/Ca (Table 3.39A, B). Barium/calcium, Co/Ca, Mg/Ca, and Ni/Ca were associated with length and mass with directionality element specific (Table 3.39B). The Ni/Ca associations, as with the prior analyses, however, were based on a small sample size and considered indeterminate.

The Spearman's correlation procedure was performed after the Pearson correlation analysis on the same dataset (Table 3.40). A different strategy for classifying level of association was used: moderate association was assumed 0.4-0.6 and weak and strong either side of the moderate. Those comparisons showing significant association with time were length, mass, B/Ca, Ba/Ca, Co/Ca, Cu/Ca, Fe/Ca, K/Ca, Mg/Ca, Mn/Ca, P/Ca, Si/Ca, and Zn/Ca. Associations with length were shown to

exist in variable degrees with mass, Ba/Ca, Co/Ca, Fe/Ca, K/Ca, Li/Ca, Mg/Ca, Ni/Ca, P/Ca, Sr/Ca, and Zn/Ca. Lastly, associations with mass were revealed for Ba/Ca, Co/Ca, Fe/Ca, K/Ca, Li/Ca, Mg/Ca, Ni/Ca, P/Ca, Sr/Ca, and Zn/Ca.

After consideration of the descriptive statistics, data plots, and results of the correlation analyses, the most appropriate and best fit function was determined. Table 3.41 identifies the best fit identified between monotonic and linear relationships by variable. It is important to understand that these associations and descriptions are specific to the dataset being used and overall results of the present study.

Further analyses were performed to determine if any associations were evident between the different elemental ratios examined during the course of this study. Again, a Pearson correlation and Spearman correlation were performed due to the mixed nature of the data set. Many of the comparisons show a moderate to strong association within the context of the Pearson correlation procedure with Fe, Mg, Mn, Si, and Sr being most often seemingly associated with the other elemental ratios. The majority of the associations identified was positive and summarized in Table 3.42. For comparison, the results of the Spearman correlations are provided in Table 3.43. It is evident that there is more often an association between the ratios than not. This interpretation, however, is specific to the findings of the present study.

Discussion

The dominant focus of past research has been aimed at determining chemical signatures in bivalve shells that are capable of being used as environmental proxies. Researchers, however, have frequently noted apparent confounded trends in these

Table 3.39A: Pearson correlation analysis results of significance for element/calcium ratio associations with time, shell length, and valve mass. The correlation coefficient (Corr Coef), the P value (P), and the number of samples examined (n) are provided. The shading variations indicate level of association based on the correlation coefficient (light shade = weak, medium shade = moderate, dark shade = strong).

		Length	Mass	Ва	Co	К	Li	Mg	Ni	Zn
	Corr Coef	0.253	0.304	-0.301	0.666	-0.277	-0.216	-0.319		-0.285
	Р	1.30E- 07	1.79E- 10	2.90E-09	2.96E-18	8.68E-08	3.35E-05	3.23E-10		3.70E-04
Time	n	422	422	373	132	362	362	370		152
	Corr Coef		0.906	-0.261	0.295			0.112	-0.564	
	Р		6.03E- 181	4.07E-08	2.56E-04			2.12E-02	2.19E-03	
Length	n		480	428	149			425	27	
	Corr Coef			-0.227	0.345			0.0961	-0.511	
	Р			2.00E-06	1.69E-05			4.77E-02	6.49E-03	
Mass	n			428	149			425	27	

Table 3.39B: Direction of association for initial Pearson correlations

	time	length	mass	Ва	Со	к	Li	Mg	Ni	Zn
time		+	+	-	+	-	-	-	-	-
length			+	-	+			+	-	
mass				-	+			+	-	

		Len	mass	В	Ва	Со	Cu	Fe	К	Li	Mg	Mn	Ni	Р	Si	Sr	Zn
	сс	0.216	0.252	0.174	-0.186	0.63	-0.439	0.356	-0.415		-0.435	0.269		0.17	0.502		-0.687
	P Value	0	0	0	0	0	0	0	0		0	0		0.003	0		0
time	n	422	422	374	373	132	237	361	362		370	373		303	365		152
	СС		0.957		-0.305	0.235		-0.153	0.44	0.311	0.381		-0.626	0.154		0.645	-0.174
	P Value		0		0	0.004		0.0017	0	0	0		0	0.004		0	0.0182
length	n		480		428	149		416	416	415	425		27	350		420	184
	СС				-0.306	0.258		-0.14	0.447	0.296	0.371		-0.611	0.185		0.641	-0.18
	P Value				0	0.002		0.0042	0	0	0		0	0		0	0.0144
mass	n				428	149		416	416	415	425		27	350		420	184

Table 3.40 Spearman's correlation analysis results of significance for element/calcium ratio associations with time, shell length, and valve mass.

Table 3.41 Best fit correlation function for data description.

Linear	Ba/Ca	Co/Ca	Li/Ca	Mg/Ca	Ni/Ca					
Monotonic	length	mass	B/Ca	Cu/Ca	Fe/Ca	K/Ca	P/Ca	Si/Ca	Sr/Ca	Zn/Ca

Table 3.42: Significant element/calcium ratio correlation results – Pearson correlation. Weak associations have been highlighted in light grey; moderate in a shade darker and the strong associations the darkest shade. The correlation coefficient (Corr Coef), the P value (P), and the number of samples examined (n) are provided in order.

	Co/Ca	Fe/Ca	K/Ca	Li/Ca	Mg/Ca	Mn/Ca	P/Ca	Si/Ca	Sr/Ca	Zn/Ca
	-0.175	0.997	0.925		0.83	0.985	-0.231	0.999	0.983	
	0.0326	0	2.92E-		1.54E-	6.818E-	0.00001	0	1.35E-	
B/Ca	149	413	408		418	420	348	417	395	
			0.217							0.314
			9.91E-06							0.00001 41
Ba/Ca			409							184
							0.516	0.268	0.251	-0.499
							2.71E-09	0.00102	0.00322	0.00001 46
Co/Ca							117	147	136	68
			0.915	0.84	0.829	0.989	0.118	0.999	0.997	
			5.15E- 159	5.90E- 108	3.99E- 105	0	0.0289	0	0	
Fe/Ca			401	400	410	413	344	411	383	
					0.905	0.894	0.227	0.914	0.959	
					1.30E- 154	4.97E- 145	0.00002 99	7.16E- 160	4.19E- 212	
K/Ca					412	413	333	405	386	
						0.718			0.554	
						1.24E-66			2.36E-32	
Li/Ca						413			385	
						0.809	0.233	0.823	0.842	
						2.79E-99	0.00001 15	5.67E- 104	8.58E- 108	
Mg/Ca						423	346	417	396	
							0.116	0.995	0.99	
							0.0302	0	0	
Mn/Ca							347	418	397	
P/Ca										
									0.995	-0.16
									0	0.0312
Si/Ca									391	181

Table 3.43 Significant element/calcium ratio correlation results – Spearman correlation. Weak associations have been highlighted in light grey; moderate in a shade darker and the strong associations the darkest shade. The correlation coefficient, The P value, and the number of samples examined provided in order (Top to bottom) for each comparison.

	Ba/Ca	Co/Ca	Cu/Ca	Fe/Ca	K/Ca	Li/Ca	Mg/ Ca	Mn/ Ca	Ni/Ca	P/Ca	Si/Ca	Sr/Ca	Zn/Ca
	-0.186	0.167			-0.102	0.425			0.4	-0.166	0.186	0.412	
	0	0.040			0.039	0			0.038	0.001	0		
D /0	0	0.042			3	0			4	9	0	0	
B/Ca	424	149			408	409			27	348	417	395	
			0.162	0.294				0.15				-0.31	0.353
			1	0				1				0	0
Ba/Ca			267	412	ļ			419				394	184
				0.291		0.449		0.185		0.59	0.371	0.267	
				0		0		0.024		0	0	0.001	
Co/Ca				143		149		148		117	147	136	
					0.192	0.25	0.417	-0.24			-0.194		0.563
					0.001	0	0	0			0.001		0
0/0					9	0	0	070			0		100
Cu/Ca					259	260	268	2/2	0.405	0.445	264	0.000	132
					-0.356	0.002	-0.276	0.617	0.465	0.115	0.723	-0.288	-0.17
					0	3	0	0	8	9	0	0	9
Fe/Ca					401	400	410	413	26	344	411	383	184
		ļ			ļ	0.386	0.818	-0.194		0.317	-0.331	0.382	0.434
						0	0	0		0	0	0	0
K/Ca						408	412	413		333	405	386	182
							0.523			0.108		0.589	0.363
							0			0.049 6		0	0
Li/Ca							410			334		385	179
								-0.143		0.313	-0.306	0.388	0.573
								0.003		0	0	0	
Mg/C										0	0	0	0
a								423		346	417	396	181
										0.143	0.372	-0.14	
Mn/C										8	0	2	
а			<u> </u>							347	418	397	
		 	 	 	 								0.224
													0.003
P/Ca													169
													-0.43
													0
Si/Ca													181

	Ba/Ca	Co/Ca	Cu/Ca	Fe/Ca	K/Ca	Li/Ca	Mg/ Ca	Mn/ Ca	Ni/Ca	P/Ca	Si/Ca	Sr/Ca	Zn/Ca
													-0.159
													0.047 3
Sr/Ca													156

Table 3.43 (Continued)

signatures due to biologic factors (Rosenberg, 1980; Wheeler, 1992; Carre et al., 2006; Strasser et al., 2008). Research specific to biologic controls and influences on shell chemistry is limited despite the assertion that elemental composition is highly dependent on these factors (Wheeler, 1992).

The results presented in this chapter elaborate on the potential influences of specific algal diets on the elemental shell chemistry of *Mercenaria mercenaria* under controlled conditions. Overall, the differences observed in this study can be categorized as influenced by pre experimental factors, seemingly biologic/metabolic factors not assessed, assumed metabolic factors due to stress or nutritional variance, or directly associated with diet received. All factors aside from pre experimental influences can be related to the study design and project variables which ultimately link the observations to diet; however, direct influences and indirect influences have been regarded as separate due to project scope and reliance on previous findings to explain observations outside the analytical design.

Shell length and mass

In general diets composed of multiple algal species have been shown to be superior to those composed of single algal species (Romberger and Epifanio, 1981; Albentosa et al., 1993; Brown et al., 1997). Diatoms, however, have been shown to be rich in fatty acids (Brown et al., 1997) and support healthy growth rates and survival of cultured *Argopecten irradians* (Milke et al., 2006), though Prymnesiophytes such as *Pavlova* and *Isochrysis* species have also been shown to be relatively rich in fatty acids (Brown et al., 1997). In fact, Prymnesiophytes were shown to have the highest percentage of saturated fatty acids which were suggested to be more beneficial to *C. gigas* (the pacific oyster) larval growth compared to polyunsaturated fatty acids (Thompson et al., 1993). The results of the present study seemingly supported the noted benefit of mixed algal diets and the correlation with growth rates and diatom inclusion; however, previous observations associated with Prymnesiophyte diets were not duplicated. Replication of results determined for different species of bivalve is not necessarily expected due to different requirements of the animal being examined (Brown, 2002) as well nutritional value of the microalgae fed can change due to differences in culture conditions (Enright et al., 1986).

In my experiment, clams fed the Mixed diet or *Chaetoceros mulleri* (CCMP1316)[Cm] diet exhibited the most consistent increase in shell length. The *Chaetoceros galvestonensis* (CCMP186)[Cg] fed group exhibited the next most consistent shell length increase following the second collection which was suspected to be a consequence of algal strain culture problems prior to the March collection. The *Isochrysis species* (CCMP1611) [ISO], *Pavlova species* (CCMP1209)[Pav1209], and *Pavlova pinguis* (CCMP609)[Pav609] fed clams exhibited moderate to poor shell length increase with *Isochrysis* sp. (CCMP1324)[TI] fed clams and the control group [CTRL] exhibiting virtually no growth during the experiment. All feeding groups except for the *Isochrysis* sp. (CCMP1324)[TI] fed group differed significantly from the control group in

terms of overall shell length. Regarding the valves used in the elemental chemistry analyses, median collected shell length never significantly exceeded the baseline measurements. When the same comparisons were made using all valves collected throughout the experiment, however, the Mixed and *Chaetoceros mulleri* diets were found to promote significant growth over baseline.

The valve mass followed the same trends as the shell length in regard to the overall mass increase of the left valves collected. The right valves collected and subsequently analyzed for the purposes of this experiment exhibited a different trend where ISO>Pav1209>Mixed>Pav609>Cm>Cg>Tl; however, the right valves did not differ significantly from the left and were only specifically measured during the first two trials, thus did not reflect changes occurring through the entire experiment. Statistically, the left valve mass for both the mixed and Cm diet groups exceeded those of the control and baseline valves. The remaining groups, excluding TI, significantly differed only from the control group.

Overall, only the Cm and Mixed diet groups exhibited significant growth throughout the experiment. This is an important consideration. It has been suggested variable growth or shifts in shell morphology could possibly affect shell deposition and elemental shell chemistry of the valves of bivalve molluscs (Rhoades and Lutz, 1980) with one of the associated factors of shell form being food supply (Stanley, 1970). Additionally, past research has identified potential associations between growth and element incorporation, i.e. Strontium (Gilken et al., 2005), Manganese (Carre et al.; 2006; Strasser et al., 2006), and Magnesium (Carriker, 1996).

Left versus right valve elemental chemistry

Due to the observed variation between left and right shell mass, as well as previous observations in the eastern oyster having increased elemental concentration in the right valve (Carriker et al., 1996), element/Ca ratios were compared between the left and right valves of the experimental clams. A step-wise approach was taken during analysis to develop a more comprehensive picture of the elemental dynamics between the shell valves.

In the previous chapter, no significant differences were found between the right and left valve groups of Argopecten irradians. This result did not support the original hypothesis, but was suggested as a function of the more equivalved nature of the scallops as compared to observations made by Carriker et al. (1996) in reference to the eastern oyster, Crassostrea virginica. The other possibilities explaining the results being different than expected were, 1. the use of element/Ca ratios to normalize concentration for the examination as compared to the previously identified oyster study which examined straight elemental concentrations or 2. the analytical methodology was insufficient to identify any differences between valves if present. In the current study, Mercenaria mercenaria, an equivalved bivalve, was examined using similar analytical procedures which provided for somewhat different results from that of the scallop study. Both Ni/Ca and Zn/Ca differed significantly between the left and right valves; however, the nickel ratio comparison was based, overall, on only a few diets and not representative of all clams analyzed. A notable difference in the experimental design was the addition of a starvation control in the present clam study which contributed an additional influence for consideration – starvation and associated stress.

When the left and right valves were examined specific to diet received, two diet groups exhibited significant differences with respect to four different element/Ca ratios. The right valves from the Control were determined to have a higher ratio value with respect to B/Ca, Si/Ca and Zn/Ca as compared to the left valve. The right valves of the Cm group had a higher ratio with respect to Ba/Ca as compared to the left. This similar to the observations of Carriker et al. (1996), enrichment was specific to the right valve in each case.

The enrichment of elements or of specific elements in the right valve could be due to a genetic disposition such as remnant biologic controls for torsion (personal communication with Sandra E. Shumway) or potentially, metabolic differences between the mantle tissues associated with each valve. The difference between the valves being a function of metabolic change is partially supported by the results of the control group. Three separate elemental ratios differed between the left and right valves compared to only one in the Cm group and zero in the other groups. Additionally, the differences in the diet specific comparison - Zn/Ca. The control group valves were most probably the dominant influence of the Zn/Ca result from the composite analysis. It is believed that an increased differences between the left and right valve is a function of stress related metabolic changes.

With respect to the difference observed between the valves of the Cm feeding group in the present study, metabolic stress or changes do not seem as suspect when taking into consideration experimental procedure and the comparisons of shell length and mass discussed previously. The specific diet received, however, could potentially

explain the difference. Barium is released during phytoplankton decay (Stecher and Kogut, 1999) and Ba is often linked to diatoms (Vander Putten et al., 2000) as it can substitute for Si in the frustules. A minor fraction of Ba in the water column is transformed to barite crystals but most remains labile (Ganeshram et al., 2003). It is proposed that Ba/Ca was enriched in the right valve due to exposure of one valve to settled barite or increased dissolved barium as might occur at the sediment-water interface (Tabouret et al., 2012).

Elemental shell composition differences as a function of diet

Assimilation of material from ingested food particles is directly related to the food itself, i.e., size, biochemical composition, quantity and cell wall (Bayne, 1993; Reinfelder and Fisher, 1991). Furthermore, the partitioning of an element within an algal cell has been directly related to assimilation efficiencies of specific elements studied in *Mytilus edulis* shells and soft tissues (Wang and Fischer, 1996). It follows that elements are more readily available for assimilation from certain algal cells than in others and selective feeding could potentially limit the elements internalized providing for a mechanism of diet associated elemental shell signatures. Additionally, elemental composition has been shown to differ among algal taxa and between oceanic and neritic strains (Martin and Knauer, 1973; Lee and Morel, 1995; Sunda and Huntsman, 1995; Ho et al., 2003) thus increasing potential for diet associated signatures.

With this in mind, the algal cultures used in this experiment were selected based on multiple criteria. First all genus- species pairs obtained had been isolated from as different areas and climates as possible. Second, one of the Genus species pair was

used commonly in aquaculture settings. Third, one species was isolated from inshore waters. Fourth, one pair were diatom species. The final criterion was that the algae would grow in the laboratory and in outdoor cultures. As identified by the Provasoli-Guillard National Center for Marine Algae and Microbiota of Bigelow Laboratory for Ocean Sciences, the *Pavlova species* (CCMP1209) was isolated from an unknown tropical site and *Pavlova pinguis* (CCMP609) the Sargasso Sea; *Isochrysis* sp. (CCMP1324) was isolated from the South Pacific off the Society Islands and Isochrysis *species* (CCMP186) was isolated from the Gulf of Mexico off St. Petersburg, FL while *Chaetoceros mulleri* (CCMP1316) from the North Pacific off Hawaii.

Of the 29 elements originally assessed, only 16 were detected in sufficient concentrations for use in the analyses. All valves collected from each experimental diet were initially assessed for differences in B/Ca, Ba/Ca, Cd/Ca, Co/Ca, Cu/Ca, Fe/Ca, K/Ca, Li/Ca, Mg/Ca, Mn/Ca, Ni/Ca, P/Ca, Pb/Ca, Si/Ca, Sr/Ca and Zn/Ca. Due to sample size limitations created for specific elements during analysis and post processing procedures, Cd/Ca, Cu/Ca, Ni/Ca, and Pb/Ca were eliminated from most of the analyses conducted.

As within the scallop experiments discussed in Chapter 2, the elemental data were highly variable both across diet groups as well as within diet groups. This, however, was similar to results previously reported with cultured bivalves (Carriker, 1996; Strasser et al., 2008), as well as, cultured foraminifera (Hintz et al., 2006). This consistent problem with cultured molluscs and other taxa further complicates analysis and, especially, evaluation of elemental trends in the shell.

I found that, in hard clams fed unique diets, the boron concentration with respect to calcium in the shell was higher in both the Cg and Mixed groups compared to the ISO and Pav609 groups and higher in the TI diet group as compared to the baseline, Cm, Control, ISO, Pav609 and Pav1209 groups. Boron has been associated with salinity (Schopf, 1980) and isotopes have been used as proxies for environmental pH (Rollion-Bard et al., 2003). Salinity was held constant among all feeding groups and therefore was considered non-influential with regard to differences revealed in my study. The growth parameters assessed also do not lend any evidence that this ratio was directly influenced by growth rates observed during the feeding trials, though, upon further consideration and examination of the right valves of the Control group, it was noted that the significant increases in B/Ca occurred in the two groups associated with the lowest level of growth. Furthermore, the increased ratio in the Cg group over two compared groups could account for influence of the clams in the specific collection (March) exhibiting poor growth. The increased ratio over the same two diets in the mixed group did contradict this association. It was previously identified in Rosenberg (1980) that effects of nutrition on boron concentrations not assessed could complicate associations between boron and environmental factors. I examined specific dietary factors in the current study with seemingly confounded results. As such, based on the deviation of trends observed, it is suggested that B/Ca is influenced by multiple factors with substitution, pre experimental conditions, and stress plausible explanations for the differences observed.

The examination of Ba/Ca suggested that Baseline values tended to be higher than or similar to those of the majority of diets in both the left and right values.

Commonly, as in Stecher et al. (1996), Thebault et al. (2009), Putten et al. (2000), and Lazareth et al. (2007), barium is associated with phytoplankton bloom dynamics (diatoms mostly). This association was further developed by Tabouret et al. (2012) when concluding Ba enrichment was most likely due to incorporation of dissolved Ba such as might be available following an algal bloom (Stecher and Kogut 1999) or at the water-sediment interface. The conclusion made in reference to the observations in the present study is that the higher Ba/Ca ratios are due to influences prior to the start of the feeding experiment and potentially a consequence of the brown water culture methodology immediately prior to the investigation.

The analysis of Co/Ca in the left valves of the different feeding groups showed higher ratios in both the ISO and TI groups than in the Mixed diet group. Research performed by Ho et al 2003 suggested the examined Coccolithophores to have a higher Co quota than that of diatoms though this ultimately suggested being due to the origin of the isolates – the Coccolithophores being oceanic and the Diatoms being neritic. The differences in metal requirements between oceanic and neritic algal species was examined in Brand et al. 1983 where it was suggested oceanic species are associated with higher Co quotas than neritic species. Because all diet groups were seemingly similar based on the statistical analyses besides the shown difference between the Mixed diet and the two Prymesiophycea species, Co/Ca was interpreted as mostly stable during the course of this study and without experimental influence; though, sample size attributable to this result and interpretation may not be fully representative of all changes.

The results obtained from statistical analysis of Fe/Ca are interesting. Specific to the left valve comparisons, the Baseline, ISO and Pav1209 groups are all associated with higher Fe/Ca values than Cg, Cm, CTRL, and Mixed diet groups. The Pav609 group had a higher ratio value associated as compared to Cg and CTRL, while, the TI associated ratio was higher than Cg. In respect to the right valve analysis, the ISO, Baseline and Mixed diet groups were associated with a higher Fe/Ca than the control, furthermore; Baseline Fe/Ca was higher than the TI associated value. Iron has been labeled as a major component in shell deposition and otherwise linked to shell development, proteins and pigments (Almeida et al. 1996). The differences between diet groups in this study appear more attributable to the diet received versus related factors such as growth. The consistent enrichment in the Pavlovophycea and Coccolithophyceae (Prymnesiophycidae) Bacillariophycea over the (Coscinodiscophyceae) diet containing Pavlovophycea or in а the and Coccolithophyceae suggests the possibility of Fe enrichment by ingestion of particular algae. This would follow the results of Ho et al 2003 which demonstrate a slightly higher average Fe/P in Prymesiophycea species examined than in the Bacillariophycea species. Confounding factors, however, might be loss of Ca in the Pavlovophycea diets, or loss of iron in the control group giving the impression pre experimental causes were not dominant.

The findings in reference to right valve associated K/Ca are similar to those observed during the scallop research presented in Chapter 2. The K/Ca ratio value is higher in both the Baseline and Pav1209 groups as compared to Cg but all remaining diets similar. The left valve comparisons did not identify any significant differences

despite the Baseline value appearing larger than all other diet group medians and the control seemingly smaller which would support a gradual decrease in element enrichment. Carriker et al. (1996) concluded that differences in the potassium concentrations observed were possibly from ingestion of sedimentary particles containing potassium though specific examination of the algal cells being ingested was not included in the scope of the research. Conversely, Ho et al. (2003) suggested K is in higher concentration in algal cells than in seawater and that diatoms have a much higher K quota than the other taxonomic classes examined. The results of the current study do not support enrichment of potassium in clam shell by dietary intake but do suggest pre experimental factors might have influenced the observed ratios.

The analysis of Li/Ca was similar in both the left and right valve assessments. Baseline values were significantly larger as compared to the majority of the experimental diets. The ratio for TI was also significantly larger than Cg in valve assessments as well as Cm, ISO, and Pav1209 for the left valve assessment. Research conducted on Li/Ca in *Arctica islandica* concluded patterns of the elemental ratio were most likely associated with calcification rate or river inputs (Thebault et al., 2009). Thebault and Chauvaud (2011) examined Li/Ca signatures in *Pecten maximus* with the conclusion that growth, potentially temperature and increased Lithium due to diatom blooms were responsible for enrichment. In the present study, the differences observed seem more indicative of the preserved signature from pre experimental growth similar to baseline with potential loss or signature dilution due to new growth.

The left values of each feeding group were found not to differ significantly with respect to Mg/Ca; however, the right value comparison resulted in three significant

comparisons between the control, baseline, Cg, and mixed diet groups with the control group being associated with the highest ratio value. Magnesium has been proposed to fluctuate with temperature in certain species of bivalve (Rucker and Valentine, 1961; Dodd, 1965; Rosenburg, 1980); though, recent research suggests temperature or salinity have minor to no influence on Mg/Ca for specific bivalves (Strasser et al., 2008; Carre et al., 2006). In *Crassostrea virginica* (Carriker, 1996) and certain gastropods (Foster and Cravo, 2003) Mg has been shown to increase with size and through ontogeny. Ho et al. (2003) suggests that of the five taxonomic classes of algae examined in their study, diatoms had the largest Mg/P quota while the others were mostly similar. Assimilation of Magnesium, however, was suggested from the dissolved phase in Poigner et al. (2012). Because temperature was not a variable in the present study and enrichment was not specific to the diatom diets, growth rates, metabolic control, or calcium replacement may be plausible explanations.

The results of the diet comparisons with regard to Mn/Ca for the collected left valves established that the control group ratio was lower than all the other groups. The Cg group associated ratio was determined smaller than the ratios for Isochrysis species, the mixed diet, and both Pavlova species; and following the developing trend, the Cm group ratio was deemed smaller that of both Isochrysis groups.

The same analysis using the collected right valve ratios established that Mn/Ca was smaller in the Cg and CTRL groups than that of the baseline, Cm, ISO, Mixed, and TI groups. It was also determined the mixed diet associated ratio was larger than that of both Pavlova species.

Defining environmental associations for manganese has been problematic. Blanchard and Chasteen (1976) surmised the amount of substitution of Ca²⁺ by Mn²⁺ was correlated with tidal level, though did not consider the oxygenation of the environment (Rosenburg, 1980). Crisp (1975) attempted to correlate the Mn concentration to salinity, while Strasser et al. (2008) correlated Mn/Ca levels with that of seawater concentration, potentially confounded by biological activity. Carre et al. (2006) suggested positive Mn/Ca association with growth rates while Strasser et al. (2006) showed negative correlation. Though, Poigner et al. (2012) suggests manganese is assimilated most often through its dissolved phase, the results of the current study in conjunction with the trends visualized in Figure 3.31 seem to support the possibility that manganese enrichment was influenced by the diet received, more specifically the Prymnesiophyceae diets, which was supported in Ho et al. 2003 where it was shown average Mn/P concentrations in the Coccolithophores were greater than those in other phyla inspected, including diatom species. These results, however, do not support any differences between oceanic and neritic species (as suggested in Brand et al., 1983 and Ho et al., 2003) being translated to the shells.

The comparisons of P/Ca between diet groups only resulted in significant differences with respect to the left valve. It was determined P/Ca was increased in the ISO group over all other diets and in TI over Cg, Cm, and mixed diets. This suggests a fairly stable association with increased P/Ca in the two Isochrysis species. Ho et al. (2003) points out the major nutrients C, N, P, and S are variable in algae but the average quotas of the organic biomass being similar to that of Redfield et al. (1963). This current observation could be attributed to an increased P concentration in the shell
from feeding solely or selectively upon specific cells in conjunction with an otherwise relatively stable signature.

The Si/Ca ratio was higher in both *Pavlova* species as compared to Cg and Cm in the left collected valves, additionally; the mixed diet had an associated Si/Ca lower than the remaining diets. When the right valves of the feeding groups were compared, baseline Si/Ca was higher than in the diatom species, Pavlova species and mixed diet treatments, in addition, the Si/Ca in the control treatment higher than those associated with Cg, Mixed, Pav609 and Pav1209 feeding groups. Carriker et al. (1996) concluded that the oysters maintained in the natural environment received silica through seston not incorporated in the laboratory habitat. Considering this, the differences in Si/Ca observed are most seemingly due to pre experimental conditions with a potential secondary dietary influence.

Four comparisons with regard to Sr/Ca between feeding groups were deemed significant and established Cm group left valves were Sr enriched compared to ISO, Pav609, and Pav1209 and Mixed diet group valves enriched compared to Pav1209. When the right valves were compared, only the TI was determined enriched over the Cg group. These results do not consistently align with the description in Ho et al. (2003) where Coccolithophores were determined to have a higher Sr quota than studied diatoms and other algal phyla. As such, there is no clear association with any experimental variable nor is there an obvious indication of an artifact effect from the rearing procedure. This, however, is a common problem in determining factors influencing Sr in shells. Researchers have documented associations with temperature (Dodd, 1965), growth rates (Gilken et al., 2005; Hamer and Jenkins, 2007), and

ontogeny (Dodd, 1970; Crisp, 1975); however, there are equal accounts of opposing influence as well as results indicating no influence when considering the same factors. Poigner et al. (2012) concluded that Strontium was predominantly assimilated in particulate phase; however, the current results do not support a dietary influence on Strontium incorporation. The results do suggest multiple controlling factors with one potentially being shell growth and growth rate.

Zinc to calcium inspections revealed two significant comparisons with regard to the left shell and four with regard to the right. The baseline and Cg group were determined significantly enriched compared to the ISO group left valves. The right valve comparison established that the Cg and control were associated with higher Zn/Ca than were the Mixed and Pav1209 diet groups. As with strontium, there is not a clear indication of what contributed to the limited differences observed. Many researchers have paired zinc concentrations with environmental concentrations for purpose of contaminant surveys (Fang and Shen, 1984; Martincie et al., 1984; Puente et al., 1996; Markich, 2002; etc.). Zinc has been, however, recognized as a dietary element metabolically transformed and passed through food web interactions (Windisch 2001, Wang 2002). In the present research, the differences observed were not seemingly attributable to a contaminant source as the association was specific to diet groups. The differences observed in the current study, though, do not support, definitively, a specific algal diet contributing to relative zinc concentration.

Correlation analyses

Correlation analyses revealed associations between B, Fe, K, Mg, Mn, Si and Sr as well as Fe and Li. In relation to the current project, both Fe and Mn were determined to be potentially related to the Prymnesiophyceae diets thus a correlation between the two is logical if influenced by the same variable. Similarly, Thebault and Chauvaud (2011) suggested Li was enriched due to diatom blooms which would indicate potentially similar influences between Fe and Li though different pathways/mechanisms are suspected based on previous research discussed. Because the original assessments of B, K, Mg and Sr were uncertain as to the dominant influences surrounding enrichment, the relationship is still unclear. The correlation analyses performed with respect to Time, Length and Mass do support influence of pre experimental factors (K and Mg), size and early growth rates (Sr and Mg), or undetermined biological influences (B and Mg).

Iron, manganese and phosphorus showed no strong associations with time or growth. Because the majority of potential influences identified in the literature were held constant or consistent among trials and they seemingly acted in varied association with one another, diet still appears to be a plausible primary influence.

Silica did show moderate association with time and strontium was strongly correlated with growth. Neither Silica nor Strontium showed a clear association with the diets or assessed experimental parameters. Strontium, however, was determined to show a potential relationship with growth and pre-experimental growth rates due to the diet groups associated with the highest Sr/Ca ratios. The positive association between Si and time does not agree with the conclusions made when comparing direct dietary

influence; this however, does substantiate the possibility of two or more influences and subsequent confounded results.

The Ba, K, Li, Mg and Zn associated ratios were all negatively associated with time. The original assessments of Ba, K, Li, and Mg with regard to algal diet influence suggested the relative concentration of these elements was more strongly associated with pre experimental conditions or related factors. The determined overall decrease through time does not preclude this determination; however, exact reasoning for the decrease is seemingly element specific and dependent on the influence of the experimental feeding group. The correlation between zinc and time is potentially attributable to an experimental complication. As previously noted, Windisch (2001) and Wang (2002) demonstrated zinc to be metabolically influenced and associated with food web transfer. An association between Zn/Ca with specific algal diets was not demonstrated in this study; however, production of feeding cultures was ultimately limited by available space and laboratory equipment. Similar to the differences between valves with regard to relative zinc concentrations in the starvation control, the negative association to time is likely a consequence of limited food availability and associated metabolic influence.

The initial interpretation made concerning Co/Ca was that it appeared stable across diets and did not seem influenced by the experimental variables. Here it was strongly associated with time and moderately with mass and shell length. These associations, however, are very specific to particular diet groups and do not represent overall changes in this elements concentrations.

Conclusion

Once all results were considered, three ratios were directly associated with diet received: Fe/Ca, Mn/Ca, and P/Ca. In all three cases, the Prymnesiophycidae (the two Isochrysis diets) had a significant impact on the relative elemental concentrations, as did the Pavlovophycea diets with regard to Fe/Ca. The Mg/Ca and Sr/Ca ratios were both influenced by growth or more specifically growth rates; however, it should be noted that pre experimental growth was most attributable to this conclusion and biologic factors not assessed also seemingly contributed to differences observed. With regard to Zn/Ca and B/Ca, the results favor differences observed being most attributable to stress and assumed metabolic shifts; while the remaining elemental ratios examined, Ba/Ca, K/Ca, Li/Ca and Si/Ca, were determined to be primarily influenced by factors associated with pre experimental conditions not encompassed in the experimental scope and design.

Manganese, copper, zinc, and cadmium have been shown to be primarily associated with the organic matrix in oyster shells (Carriker et al. 1980) as was Iron (Almeida et al. 1998) and magnesium in mussel shells (Lorens and Bender 1980). This association of specific elements with the organic matrix alone suggests a high level of biologic control. In the present study, iron, manganese, and phosphorous were revealed to be influenced by diet. It has been suggested that the organic matrix is responsible for shell strength (Addadi et al. 2006) and that the organic matrix potentially influences elemental content and structure of the shell (Watabe et al. 2001, Takesue and van Green 2004, Morse et al. 2007, and Johnstone 2008). If diet associated elements are incorporated into the organic matrix and can be manipulated by dietary

changes as demonstrated, the diet of cultured bivalves could potentially be implicated in differences in shell strength and structure between wild and aquacultured *Pecten maximus* as described in Gresfrude and Strand (2006). This demonstrated diet influence also demands consideration of diet composition when conducting experiments aimed at describing influences on shell dynamics or determining cause and effect relationships.

Though diet was shown to directly affect the shell chemistry of *Mercenaria mercenaria*, resolution of the algal contribution was limited to taxonomic class with regard to only a few elements, and algal isolate origin related differences were not apparently translated to the shells as had been hypothesized. It also is evident that other contributing factors will potentially complicate detection of diet associated signals when outside a controlled environment (higher food concentrations, multiple influences on the same element, contamination, multiple algal species with similar elemental quotas, age influence, etc.). As such, applications or use of diet related elemental signatures is currently limited. It is believed that diet associated signatures can eventually be used as shell markers or in determination of food web interactions related to eutrophication, ocean acidification, or other environmental impacts capable of changing plankton composition; however, before these applications can be pursued, further research and development of alternative analytical techniques are needed.

4. Can phosphorous normalization indicate dietary influence on elemental chemistry in bivalve shells?

Introduction

The productivity and species composition of marine phytoplankton communities are controlled by a number of trace metal nutrients (iron, zinc, cobalt, manganese, copper, and cadmium) as well as major nutrients (nitrogen, phosphorus, and silicon) in biologically available forms (Sunda, 2012). Trace metals can play important roles in regulating the species composition of phytoplankton communities because of large differences in cellular trace metal concentrations and growth requirements among species (Brand et al., 1983; Sunda and Huntsman, 1995; Crawford et al., 2003; Ho et al., 2003). The species composition will ultimately affect consumers, namely bivalves as related to present research, as it defines the available food source.

As presented in Chapters 2 and 3, the types of algae ingested by marine bivalve molluscs can influence the elemental composition of the shell with differences in cellular elemental concentrations, assimilation controls, or metabolic influences being plausible mechanisms for the differences. The influences of individual algal diets, however, were not completely evident. An analytical process with more resolution to determine the contribution of an algal species appears to be necessary to allow ecological/trophic interpretations of the bivalve diets by use of shell chemistries.

Elemental composition of algae is most often analyzed by normalizing to phosphorous (referred to as element quotas) following the protocol of Redfield (1934, 1958) and facilitating the comparison of individual organisms and natural plankton samples independent of cell volume (Ho et al., 2003). These quotas can be highly variable among algal taxa (Figure 4.1), with a range of 14-57% RSE (Ho et al., 2003; Quigg et al., 2003).

In studies of shell chemistry, elemental composition is commonly normalized to calcium to offset the spatial variability within the shell. I hypothesize that normalizing element concentrations to phosphorous or phosphorous and calcium will enhance resolution to determine species-specific influence of algae on elemental shell dynamics, provided that phosphorous signatures are stable.

Although different parameters can influence phosphorous composition in algal cells versus marine bivalve molluscan shells, a finding in the previous chapter with regard to P/Ca indicated a potential opportunity, as did findings that phosphorous concentrations in the shells of freshwater bivalves were similar within the same water shed (Jurkiewicz-Karnkowska, 2002). Certain elements can be good environmental proxies based on their relative stability in shell material from different individuals until an environmental flux occurs (Tabouret et al., 2012). In my study, I observed that P/Ca in the experimental clam shells remained fairly consistent with the exception of particular diets. This chapter examines the use of (element/P)/Ca ratios and (element/Ca)/ (P/Ca) (mathematically equivalent to element/P) to test the utility for analyzing shell dynamics with respect to diet.



Figure 4.1: Comparison of average elemental quotas among different algal taxa from Ho et al. (2003)

Methods

The general procedures for mariculture of both the algal cultures and clams were presented in Chapter 3, as were the ICP-OES analysis methods. This chapter uses information collected during the experiments described in Chapter 3 to test the hypothesis that a modified data-analysis procedure could be used to determine the influence of algal diets and ultimately used in field diagnostics.

The elemental concentration findings from the ICP-OES analysis of *Mercenaria mercenaria* valves subsampled from different feeding experiments were transformed to new elemental ratios in the form of (element/P)/Ca and (element/Ca)/(P/Ca) [referred to herein as shell quotas]. The general trends were statistically compared using a Kruskal-Wallis analysis of ranks and subsequent post hoc analysis to determine the differences in elemental shell chemistry between feeding groups. The results were then compared to previous findings discussed in Chapter 3 to evaluate utility of the proposed alternative ratios.

Boron, barium, cobalt, iron, potassium, lithium, magnesium, manganese, silica, strontium, and zinc concentrations from left shells were analyzed using both (element/P)/Ca and element/P. No baseline comparisons could be made because ICP data for phosphorous were not usable from the baseline shell analyses.

Results

Significant differences in boron, when normalized to phosphorous and calcium were found when comparing shells from several dietary treatments (Tables 4.1 and A1). These comparisons (Figure 4.2) revealed the (B/P)/Ca ratios in shells of clams fed ISO

were lower than all other treatments except for the Pav609, while the shells from the Mixed diet treatment were higher than the Cm, Ctrl, and Pav609 diet treatments.



Figure 4.2: Median (B/P)/Ca ratios versus diet received. The letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A1).

One Way Analysis of Variance						
Normality Test:	Failed		(P <	0.050)		
	Kruskal-Wa	lis One Way Analy	sis of Variance on Ra	nks		
Group	N	Missing	Median	25%	75%	
CTRL	61	6	1.41E-10	8.26E-11	2.29E-09	
ТІ	56	16	1.77E-10	1.14E-10	4.72E-10	
ISO	50	7	7.62E-11	2.81E-11	1.21E-10	
Cg	58	11	1.82E-10	1.32E-10	3.24E-10	
Cm	62	18	1.71E-10	1.26E-10	2.21E-10	
Pav609	58	18	1.39E-10	1.1E-10	1.71E-10	
Pav1209	54	14	1.68E-10	1.3E-10	2.3E-10	
Mixed	65	26	3.85E-10	1.93E-10	2.54E-09	
	H = 75.2	with 7 degrees of	freedom (P = <0.001)			

Table 4.1. Results	of Kruskal-Wallis and	lvsis for	(B/P)/Ca by	/ diet
		119313 101		y uiei.

When B/P data were compared among the diets (Figure 4.3), results (P< 0.05, Tables 4.2 and A2) were similar to those of (B/P)/Ca. The ratio for the ISO diet was lower than for all other experimental diets, while the Mixed diet ratio was significantly higher than the *Pavlova* diets, but no others.



Figure 4.3: Median B/P ratios in *M. mercenaria* shells compared to diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A2).

One Way Analysis of Variance						
Normality Test:	Failed		(P <	0.050)		
	Kruskal-Wa	llis One Way Analy	sis of Variance on Ra	nks		
Group	N	Missing	Median	25%	75%	
CTRL	61	6	4.39E-02	2.38E-02	1.49E-01	
ТІ	56	13	5.34E-02	3.38E-02	7.78E-02	
ISO	50	7	2.70E-02	9.15E-03	3.37E-02	
Cg	58	9	5.36E-02	3.43E-02	9.91E-02	
Cm	62	18	4.79E-02	4.00E-02	5.63E-02	
Pav609	58	18	4.55E-02	3.44E-02	5.39E-02	
Pav1209	54	12	4.47E-02	3.16E-02	6.61E-02	
Mixed	68	20	6.49E-02	4.72E-02	1.04E-01	
	H = 76.8	with 7 degrees of	freedom (P = <0.001)			

Table 4.2: Results of Kruskal-Wallis analysis for (B/P) by diet.

The results for barium with respect to phosphorous and calcium followed similar trends as for boron (P< 0.05; Tables 4.3 and A3): The ratios for the ISO group were lower than for all other diets, while the ratios for the Mixed diet were higher than for all diet treatments except Cg and Cm (*Chaetoceros*) diets (Figure 4.4).

The shell Ba/P ratio differed significantly (P<0.05) between seven diets. Again, ISO was determined to have the lowest ratio as compared to the other diets (Figure 4.5, Tables 4.4 and A4). All other experimental diets were found to be similar.



Figure 4.4 Median (Ba/P)/Ca ratios with associated 95% confidence intervals in Mercenaria shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A3).

The analysis of (Co/P)/Ca revealed somewhat different relationships among the dietary treatments, (P=0.02, Table 4.5). In this case, the ratio in the CTRL group was significantly lower than in the Pav1209 group (Figure 4.6, Table A5). The analysis of Co/P did not reveal any significant differences among diets (P=0.118).

One Way Analysis of Variance						
Normality Test:	Failed		(P <	0.050)		
	Kruskal-Wa	llis One Way Analy	sis of Variance on Ra	nks		
Group	N	Missing	Median	25%	75%	
CTRL	61	6	5.06E-06	2.41E-06	8.35E-06	
ТІ	56	16	5.18E-06	3.3E-06	8.71E-06	
ISO	50	7	2.9E-06	1.42E-06	3.84E-06	
Cg	58	11	5.58E-06	4.39E-06	7.8E-06	
Cm	62	18	6.06E-06	4.82E-06	7.5E-06	
Pav609	58	18	5.71E-06	4.39E-06	6.29E-06	
Pav1209	54	14	5.35E-06	4.01E-06	5.92E-06	
Mixed	65	24	8.79E-06	5.46E-06	1.54E-05	
	H = 80.3	with 7 degrees of	freedom ($P = < 0.001$)			

Table 4.3: Results of Kruskal-Wallis analysis for (Ba/P)/Ca by diet.



Figure 4.5: Median Ba/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A4).

Table 4.4: Res	sults of Kruskal-\	Nallis analv	sis for (Ba/P) I	bv diet.
		r and analy			oy ulot.

One Way Analysis of Variance							
Normality Test:	Failed	Failed (P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%		
CTRL	61	6	2.52E-02	1.09E-02	4.16E-02		
ТІ	56	18	1.56E-02	1.23E-02	2.52E-02		

Table 4.4 (Continued)						
Group	Ν	Missing	Median	25%	75%	
ISO	50	7	1.05E-02	6.94E-03	1.47E-02	
Cg	58	9	2.86E-02	1.29E-02	3.52E-02	
Cm	62	18	2.49E-02	2.19E-02	3.08E-02	
Pav609	58	18	2.26E-02	1.78E-02	2.61E-02	
Pav1209	54	12	1.66E-02	1.45E-02	2.62E-02	
Mixed	68	18	1.70E-02	1.37E-02	2.44E-02	
H = 57.8 with 7 degrees of freedom (P = <0.001)						



Figure 4.6: Median (Co/P)/Ca ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A5).

One Way Analysis of Variance						
Normality Test:	Failed		(P <	< 0.050)		
	Kruskal-Wa	allis One Way Anal	ysis of Variance on Ra	anks		
Group	N	Missing	Median	25%	75%	
CTRL	54	40	3.91E-12	3.09E-12	5.01E-12	
ТІ	56	49	6.53E-12	5.71E-12	8.92E-12	
ISO	48	36	4.85E-12	3.13E-12	6.99E-12	
Cg	58	46	6.61E-12	4.77E-12	1.00E-11	
Cm	58	34	6.01E-12	4.49E-12	1.14E-11	
Pav609	58	39	5.35E-12	4.77E-12	6.35E-12	

Table 4.5: Results of Kruskal-Wallis	analysis for	(Co/P)/Ca b	v diet.
	analyoid for t		<i>y</i> aiot.

Table 4.5 (Continued)

Group	N	Missing	Median	25%	75%	
Pav1209	54	44	9.11E-12	5.64E-12	1.33E-09	
Mixed	65	47	6.87E-12	5.37E-12	9.09E-12	
H = 16.6 with 7 degrees of freedom (P = 0.020)						



Figure 4.7: Median Co/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. No significant differences among diets were determined.

One Way Analysis of Variance						
Normality Test:	Failed		(P <	: 0.050)		
	Kruskal-Wa	llis One Way Analy	sis of Variance on Ra	nks		
Group	N	Missing	Median	25%	75%	
CTRL	54	40	1.35E-03	9.23E-04	1.95E-03	
ТІ	56	48	2.07E-03	1.55E-03	2.63E-03	
ISO	48	36	1.14E-03	9.06E-04	1.94E-03	
Cg	58	46	1.97E-03	1.41E-03	2.30E-03	
Cm	58	34	1.77E-03	1.56E-03	1.95E-03	
Pav609	58	39	1.65E-03	1.54E-03	2.05E-03	
Pav1209	54	43	1.63E-03	1.06E-03	2.22E-03	
Mixed	68	45	2.08E-03	1.43E-03	2.41E-03	
	H = 11.5	5 with 7 degrees of	freedom ($P = 0.118$)			

Table 4.6: Results of Kruskal-Wallis analysis for (Co/P) by diet.

The analyses of the iron shell quota as compared to calcium revealed the ratios for the ISO and Cg treatments to be significantly lower than for those of Cm, Mixed and Pav1209 treatments, while the ratios for the CTRL treatment to be lower than only the Mixed and Pav1209 treatments (Figure 4.8,Tables 4.7 and A6). The only significant differences in the Fe/P ratio were found between the Pav609 group and both the Cg and ISO groups (Figure 4.9, Tables 4.8 and A7).

Comparison of median (K/P)/Ca values for each experimental diet (Figure 4.10) indicated that the shells from the Mixed diet had significantly more potassium incorporated than did the shells from the ISO and CTRL groups (P < 0.05, Tables 4.9 and A8). The comparisons based on K/P provided similar results (Figure 4.11), with Mixed and Cm treatment ratios revealed as significantly higher than those from the ISO treatment (Tables 4.10 and A9).



Figure 4.8: Median (Fe/P)/Ca ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A6).

One Way Analysis of Variance						
Normality Test:	Failed		(P <	: 0.050)		
	Kruskal-Wa	allis One Way Anal	lysis of Variance on Ra	anks		
Group	N	Missing	Median	25%	75%	
CTRL	61	6	1.4E-10	7.41E-11	4.41E-10	
ТІ	56	18	2.41E-10	1.58E-10	3.83E-10	
ISO	50	8	1.65E-10	1.2E-10	2.56E-10	
Cg	58	11	1.69E-10	1.31E-10	2.5E-10	
Cm	58	15	2.88E-10	1.95E-10	5.14E-10	
Pav609	58	18	3.27E-10	1.66E-10	4.74E-10	
Pav1209	54	14	4.09E-10	2.28E-10	6.54E-10	
Mixed	65	26	3.86E-10	2.5E-10	8.16E-10	
	H = 53.7	7 with 7 dearees of	freedom. (P = <0.001)		

Table 4.7: Results of Kruskal-Wallis analysis for (Fe/P)/Ca by diet.



Figure 4.9: Median Fe/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A7).

One Way Analysis of Variance								
Normality Test:	Failed		(P < 0.050)					
	Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%			
CTRL	61	6	4.84E-02	2.72E-02	1.89E-01			
ТІ	56	16	6.52E-02	3.47E-02	9.29E-02			
ISO	50	8	5.18E-02	3.98E-02	7.09E-02			
Cg	58	10	5.30E-02	4.23E-02	7.38E-02			
Cm	58	15	7.70E-02	5.04E-02	1.35E-01			
Pav609	58	18	1.02E-01	5.29E-02	1.53E-01			
Pav1209	54	12	9.24E-02	4.65E-02	1.32E-01			
Mixed	68	20	7.17E-02	5.75E-02	9.40E-02			
H = 26.4 with 7 degrees of freedom (P = <0.001)								

Table 4.8: Results of Kruskal-Wallis analysis for (Fe/P) by diet.



Figure 4.10: Median (K/P)/Ca ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A8).

One Way Analysis of Variance								
Normality Test:	Failed	(P < 0.050)						
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing Median 25% 75%						
CTRL	61	6	5.55E-10	1.61E-10	5.71E-09			
ТІ	56	25	2.7E-09	9.98E-10	4.11E-09			
ISO	50	7	1.83E-09	5.38E-10	3.54E-09			
Cg	58	12	3.09E-09	2.37E-09	4.35E-09			
Cm	62	18	2.78E-09	1.86E-09	6.02E-09			
Pav609	58	18	3.29E-09	1.48E-09	5.31E-09			
Pav1209	54	16	2.34E-09	1.49E-09	7.14E-09			
Mixed	65	29	4.88E-09	1.39E-09	9.66E-09			
H = 23.9 with 7 degrees of freedom (P = 0.001)								



Figure 4.11: Median K/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A9).

One Way Analysis of Variance							
Normality Test:	Failed		(P < 0.050)				
	Kruskal-Wa	allis One Way Anal	ysis of Variance on Ra	nks			
Group	N	Missing	Median	25%	75%		
CTRL	61	6	1.76E-01	7.07E-02	1.98E+00		
ТІ	56	24	6.66E-01	3.18E-01	1.11E+00		
ISO	50	7	5.82E-01	1.80E-01	1.00E+00		
Cg	58	10	9.87E-01	6.48E-01	1.30E+00		
Cm	62	18	8.66E-01	6.56E-01	1.68E+00		
Pav609	58	18	1.01E+00	5.66E-01	1.63E+00		
Pav1209	54	16	6.71E-01	4.63E-01	2.17E+00		
Mixed	68	24	1.32E+00	7.13E-01	2.19E+00		
H = 24.9 with 7 degrees of freedom (P = <0.001)							

Table 4.10: Results of Kruskal-Wallis analysis for (K/P) by diet.

Comparison of median (Li/P)/Ca values between feeding groups (Figure 4.12, Tables 4.11 and A10) revealed that ratios from the ISO diet were significantly lower from all other treatments except those fed the Pav609 diet. The Mixed diet ratios were also revealed as significantly higher than those from the Pav609 diet. Comparing Li/P (Figure 4.13) only revealed that the ratios from the ISO diet group were significantly lower than those from all other diet groups (Tables 4.12 and A11).

The comparisons of magnesium concentrations as compared to phosphorous and calcium (Figure 4.14) by diet revealed that the mixed diet produced the highest ratio, being significantly higher than ratios from all diets except Cm. And again, the ISO diet produced a ratio significantly lower than most other diets, with the exception being Pav609. All other diets resulted in intermediate (Mg/P)/Ca ratios (Tables 4.13 and A12).

The shell Mg/P quota comparisons among diets (Figure 4.15, Tables 4.14 and A13) revealed once again that the ISO treatment produced the lowest shell ratios, significantly lower than all diets except the TI and the *Pav1209* diets. At the other end of the spectrum, the CTRL group had the highest Mg/P ratio, though not significantly 175

higher than those from the Mixed, *Chaetoceros (Cg and Cm)*, and Pav609 treatments. Overall, the ranking supported was ISO < Cg, Cm, CTRL, Mixed, Pav609; CTRL > ISO, Pav1209, TI; Cm> TI; and Mixed > ISO and TI.



Figure 4.12: Median (Li/P)/Ca ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A10).

One Way Analysis of Variance							
Normality Test:	Failed	Failed (P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing Median 25% 75%					
CTRL	61	6	1.72E-11	8.48E-12	2.43E-11		
ТІ	56	24	1.59E-11	9.91E-12	2.02E-11		
ISO	50	7	7.65E-12	2.66E-12	1.19E-11		
Cg	58	13	1.43E-11	1.14E-11	1.94E-11		
Cm	62	18	1.41E-11	9.82E-12	2.32E-11		
Pav609	58	18	1.18E-11	9.21E-12	1.74E-11		
Pav1209	54	14	1.37E-11	1.09E-11	1.78E-11		
Mixed	65	28	2.04E-11	1.40E-11	6.09E-11		
H = 49.0 with 7 degrees of freedom (P = <0.001)							

Table 4.11: Results of Kruskal-Wallis analysis for (Li/P)/Ca by diet.



Figure 4.13: Median Li/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A11).

One Way Analysis of Variance							
Normality Test:	Failed		(P < 0.050)				
	Kruskal-Wa	llis One Way Analy	sis of Variance on Rai	nks			
Group	N	Missing	Median	25%	75%		
CTRL	61	6	5.10E-03	2.60E-03	9.20E-03		
ТІ	56	23	3.88E-03	3.15E-03	5.72E-03		
ISO	50	7	2.38E-03	8.71E-04	3.46E-03		
Cg	58	10	4.40E-03	3.52E-03	6.14E-03		
Cm	62	18	4.09E-03	3.51E-03	5.31E-03		
Pav609	58	18	3.69E-03	3.50E-03	4.75E-03		
Pav1209	54	15	3.86E-03	3.35E-03	4.59E-03		
Mixed	68	24	4.66E-03	3.54E-03	6.53E-03		
H = 54.1 with 7 degrees of freedom (P = <0.001)							

Table 4.12: Results of Kruskal-Wallis analysis for (Li/P) by diet.

As depicted in Figure 4.16, the comparisons of manganese concentrations as compared to phosphorous and calcium among diets revealed the Mixed diet as the highest ratio, differing close to an order of magnitude from the lowest ratios in the CTRL group (Tables 4.15 and A14). The ratios from the Cm, Ti, and Pav1209 groups were identified as statistically similar to that of the Mixed group with the remaining diet groups falling between the end values. The ratios in the CTRL treatment were significantly lower than in all other treatments.



Figure 4.14: Median (Mg/P)/Ca ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A12).

One Way Analysis of Variance								
Normality Test:	Failed		(P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Missing Median 25% 75%					
CTRL	61	6	5.4E-09	3.92E-09	7.33E-09			
ТІ	56	18	4.93E-09	3.69E-09	7.53E-09			
ISO	50	7	2.97E-09	1.11E-09	4.92E-09			
Cg	58	11	5.22E-09	4.06E-09	5.98E-09			
Cm	62	18	6.52E-09	4.49E-09	1.06E-08			
Pav609	58	18	4.34E-09	3.66E-09	5.95E-09			
Pav1209	54	14	4.47E-09	3.65E-09	6.24E-09			
Mixed	65	26	8.22E-09	4.79E-09	1.69E-08			
H = 58.4 with 7 degrees of freedom (P = <0.001)								

Table 4.13: Results of Kruskal-Wallis ana	alysis for	(Mg/P)/Ca by	y diet.
---	------------	--------------	---------



Figure 4.15: Median Mg/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A13).

One Way Analysis of Variance							
Normality Test:	Failed		(P < 0.050)				
	Kruskal-W	allis One Way Ana	lysis of Variance on Ra	anks			
Group	N	Missing Median 25% 75%					
CTRL	61	6	2.17E+00	1.73E+00	2.40E+00		
ТІ	56	16	1.09E+00	9.36E-01	1.34E+00		
ISO	50	7	8.92E-01	3.54E-01	1.27E+00		
Cg	58	9	1.58E+00	1.28E+00	1.96E+00		
Cm	62	18	1.78E+00	1.37E+00	2.39E+00		
Pav609	58	18	1.44E+00	1.20E+00	1.82E+00		
Pav1209	54	13	1.39E+00	1.16E+00	1.53E+00		
Mixed	68	21	1.68E+00	1.28E+00	2.43E+00		
H = 81.0 with 7 degrees of freedom (P = <0.001)							

Table 4.14: Results of Kruskal-Wallis analysis for (Mg/P) by diet.

The comparison of Mn/P ratios between experimental diets (Figure 4.17) supported similar interpretations as those for the (Mn/P)/Ca comparisons. The ratios from CTRL group were significantly lower than all other treatments. The ratios for the ISO and Cg groups were revealed as similar, while the ratios for the Mixed, Cm, Ti, Pav609, and Pav1209 revealed to be similar (Tables 4.16 and A15).



Figure 4.16: Median (Mn/P)/Ca ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A14).

Table 4.15: Results of Kruskal-Wallis ana	lysis for	(Mn/P))/Ca by	/ diet

One Way Analysis of Variance								
Normality Test:	Failed	Failed (P < 0.050)						
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Missing Median 25% 75%					
CTRL	61	6	6.87E-11	5.13E-11	1.2E-10			
ТІ	56	17	4.23E-10	2.63E-10	7.1E-10			
ISO	50	7	1.75E-10	1.31E-10	3.71E-10			
Cg	58	11	2E-10	1.13E-10	3.03E-10			
Cm	62	18	4.08E-10	2.44E-10	6.27E-10			
Pav609	58	18	3.59E-10	1.96E-10	4.77E-10			

Table 4.15 (Continued)

Group	N	Missing	Median	25%	75%	
Pav1209	54	14	3.42E-10	2.61E-10	5.25E-10	
Mixed 65 26 6.26E-10 4.18E-10 1.01E-09						
H = 152 with 7 degrees of freedom (P = <0.001)						



Figure 4.17: Median Mn/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A15).

One Way Analysis of Variance							
Normality Test:	Failed	Failed (P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%		
CTRL	61	6	2.26E-02	1.63E-02	4.66E-02		
ТІ	56	15	9.62E-02	6.39E-02	1.46E-01		
ISO	50	7	5.46E-02	4.22E-02	9.31E-02		
Cg	58	9	4.76E-02	3.36E-02	1.01E-01		
Cm	62	18	1.17E-01	6.43E-02	1.60E-01		
Pav609	58	18	1.13E-01	6.16E-02	1.61E-01		
Pav1209	54	12	8.57E-02	5.04E-02	1.31E-01		
Mixed	68	20	1.12E-01	9.05E-02	1.50E-01		
	H = 122	with 7 degrees of f	reedom (P = <0.001)				

Table 4.16: Results of Kruskal-Wallis a	analysis for (Mn/P) by diet.
---	--------------------	------------

The (Si/P)/Ca ratios were highest in *Pavlova* diets with TI and Mixed diets ranking next highest and the remaining diet groups being somewhat similar (Figure 4.18). The ISO diet resulted in the lowest (Si/P)/Ca, which along with the Cg diet yielded ratios significantly lower than the *Pavlova* diets. The ratios for the Cm and CTRL were also revealed to be significantly lower than those of the Pav1209 group (Tables 4.17 and A16).

In comparing Si/P shell content between diet groups, again the highest ratios were in the two *Pavlova* diet groups, followed by TI and the CTRL, while the remaining diets were similar (Figure 4.19). The ratios from the ISO group were the lowest but not significantly different from those of the Cg, Cm, and Mixed groups (Tables 4.18 and A17).



4.18: Median (Si/P)/Ca ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A16).

One Way Analysis of Variance						
Normality Test:	Failed	ed (P < 0.050)				
Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	N	Missing	Median	25%	75%	
CTRL	61	6	1.08E-09	6.65E-10	3.23E-09	
ТІ	56	18	1.9E-09	1.14E-09	3.43E-09	
ISO	50	7	7.35E-10	4.27E-10	1.25E-09	
Cg	58	11	1.04E-09	5.39E-10	1.67E-09	
Cm	62	18	1.08E-09	6.37E-10	2.61E-09	
Pav609	58	18	3.26E-09	7.75E-10	4.54E-09	
Pav1209	54	14	3.45E-09	2.5E-09	4.81E-09	
Mixed	65	24	1.47E-09	7.88E-10	3.13E-09	
	H – 49 7	with 7 degrees of	freedom (P = < 0.001)			

4.17: Result of Kruskal-Wallis analysis for (Si/P)/Ca by diet



4.19: Median Si/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A17).

With respect to (Sr/P)/Ca, the highest concentration of Sr was found in the mixed diet group, followed by the *Chaetoceros* diet groups, then by the TI, CTRL, Pav609 and Pav1209 groups, with the ISO group significantly lowest (Figure 4.20). The statistical analyses (Tables 4.19 and A18) also revealed the ratios from the Mixed group were 183

significantly higher than that those of the CTRL, Pav609, and Pav1209 groups but similar to that of the *Chaetoceros* and TI groups.

One Way Analysis of Variance							
Normality Test:	Failed		(P <	: 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%		
CTRL	61	6	3.74E-01	2.21E-01	1.40E+00		
TI	56	16	4.67E-01	3.11E-01	7.57E-01		
ISO	50	7	2.40E-01	1.41E-01	3.37E-01		
Cg	58	9	3.03E-01	1.96E-01	4.83E-01		
Cm	62	18	2.96E-01	1.82E-01	5.79E-01		
Pav609	58	18	9.86E-01	4.16E-01	1.35E+00		
Pav1209	54	12	9.57E-01	2.91E-01	1.24E+00		
Mixed	68	19	2.53E-01	1.91E-01	3.92E-01		
H = 18.8 with 7 degrees of freedom (P = <0.001)							

4.18: Results of Kruskal-Wallis analysis for (Si/P) by diet

The Mixed diet group was associated with the highest concentration of strontium when Sr/P was compared among diet treatments, but not significantly different from that of the Cg, Cm, Pav609, or CTRL diet groups. The ratios from the ISO group were once again revealed as significantly lower than those from all other treatments (Figure 4.21, Tables 4.20 and A19).

Zinc as compared to phosphorus and calcium was significantly higher in Mixed diet group than in the CTRL and Pav609 groups (Figure 4.22, Tables 4.21 and A20). The ratio from the ISO group was revealed to be significantly lower than those from the Cg, Cm, Mixed, and Pav1209 groups. All other diets were revealed to be similar with respect to (Zn/P)/Ca.

Median Zn/P by diet (Figure 4.23) very closely resembled that of (Zn/P)/Ca. Again Cg group had the highest ratio, though not significantly different from the other 184 diet groups aside from ISO. The ratio from the ISO group was the lowest but revealed to only be significantly different from the Cg and Mixed groups (Tables 4.22 and A21).



Figure 4.20: Median (Sr/P)/Ca ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A18).

|--|

One Way Analysis of Variance							
Normality Test:	Failed		(P <	0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%		
CTRL	61	6	2.46E-08	1.38E-08	4.09E-08		
ТІ	56	19	2.2E-08	1.79E-08	4.72E-08		
Table 4.36 Continued							
Group	N	Missing	Median	25%	75%		
ISO	50	7	1.43E-08	6.61E-09	2.07E-08		
Cg	58	11	3.07E-08	2.19E-08	3.96E-08		
Cm	62	22	2.93E-08	2.13E-08	3.98E-08		
Pav609	58	18	2.5E-08	1.93E-08	3.25E-08		
Pav1209	54	23	2.41E-08	2.01E-08	2.67E-08		
Mixed	65	28	4.07E-08	2.81E-08	7.58E-08		
	H = 65.3	with 7 degrees of f	freedom (P = <0.001)				



Figure 4.21: Median Sr/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A19).

One Way Analysis of Variance							
Normality Test:	Failed		(P < 0.050)				
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%		
CTRL	61	6	8.42E+00	4.33E+00	1.66E+01		
ТІ	56	17	7.27E+00	4.81E+00	9.22E+00		
ISO	50	7	4.28E+00	2.19E+00	5.72E+00		
Cg	58	9	9.26E+00	7.41E+00	1.26E+01		
Cm	62	22	9.00E+00	7.90E+00	1.07E+01		
Pav609	58	18	8.64E+00	6.34E+00	9.73E+00		
Pav1209	54	21	7.07E+00	5.56E+00	8.47E+00		
Mixed	68	22	9.97E+00	8.19E+00	1.48E+01		
H = 80.3 with 7 degrees of freedom. (P = <0.001)							

Table 4.20: Results of Kruskal-Wallis analysis for (Sr/P) by diet



Figure 4.22: Median (Zn/P)/Ca ratios with associated 95% confidence intervals in Mercenaria shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A20).

One Way Analysis of Variance							
Normality Test:	Failed		(P <	: 0.050)			
Kruskal-Wallis One Way Analysis of Variance on Ranks							
Group	N	Missing	Median	25%	75%		
CTRL	53	16	4.72E-11	3.48E-11	7.70E-11		
ТІ	37	25	5.67E-11	3.26E-11	1.75E-10		
ISO	25	11	1.91E-11	-3.1E-10	4.54E-11		
Cg	39	7	1.1E-10	2.76E-11	2.48E-10		
Cm	45	27	7.60E-11	2.05E-11	5E-10		
Pav609	39	17	4.57E-11	1.60E-11	1.06E-10		
Pav1209	20	5	7.65E-11	5.31E-11	4.22E-08		
Mixed	29	10	9.60E-11	7.38E-11	4E-08		
	H = 32.3	with 7 degrees of	freedom (P = <0.001))			

Table 4.21: Results of Kruskal-Wallis analysis for (Zn/P)/Ca by diet.



Figure 4.23: Median Zn/P ratios with associated 95% confidence intervals in *Mercenaria* shells by diet. Letters indicate the similarities and differences revealed by Dunn's post hoc analyses (Table A21).

One Way Analysis of Variance							
Normality Test:	Failed		(P ·	< 0.050)			
	Kruskal-Wallis One Way Analysis of Variance on Ranks						
Group	N	Missing	Median	25%	75%		
CTRL	53	16	1.67E-02	1.33E-02	2.52E-02		
ТІ	37	25	1.87E-02	7.18E-03	5.94E-02		
ISO	25	11	5.02E-03	-1.08E-01	1.34E-02		
Cg	39	7	3.48E-02	9.85E-03	5.61E-02		
Cm	45	27	1.88E-02	1.17E-02	2.43E-02		
Pav609	39	17	1.28E-02	4.23E-03	3.63E-02		
Pav1209	20	5	1.62E-02	1.11E-02	2.00E-02		
Mixed	29	10	2.46E-02	1.97E-02	3.21E-02		
	H = 21.	3 with 7 degrees of	f freedom (P = 0.003)			

Table 4.22: Results of Kruskal-Wallis analysis for (Zn/P) by diet

Discussion

In general, problems associated with comparing the elemental concentrations to phosphorus include a) variance in P concentrations potentially not being uniform and attributable to the same factor in each group, b) high P concentrations contributing to unrelated minimal ratios with specific diets, and c) potential confounding factors outside controlled environments as identified in previous studies. Analyses using both the (element/P)/Ca and element/P did, however, produced results considered to be of interest with regard to six elements, either due to their similarity to elemental trends observed in algae quotas described by Ho et al (2003), Sunda and Huntsman (1995), and Brand et al. (1983), or mirroring trends observed in the results described in Chapters 2 and 3. Iron, potassium, magnesium, manganese, silica, and zinc were found to most possibly be directly associated with the diet received or assumed metabolic influences. Observed relative strontium concentrations, again, support an association with growth rate. Conversely, analysis of boron, barium, cobalt, and lithium appeared to be associated with other environmental factors or factors not associated with experimental variables.

Ho et al. (2003) presented analyses of the cellular content of 15 phytoplankton species with regard to C, N, P, S, K, Mg, Ca, Sr, Fe, Mn, Zn, Cu, Co, Cd and Mo (Figure 4.1). In general, they found that K concentrations are higher than in seawater and Mg lower, except in the case of diatoms where both Mg and K are relatively high. The major nutrients C, N, P, and S are variable with the average quotas of the organic biomass being similar to that of Redfield et al. (1963). The trace metals followed the general pattern of Fe>Mn>Zn>Cu>Co=Cd>Mo. The coccolithophores examined by Ho et al. (2003) had higher Mn, Co, and Cd quotas compared to the diatoms. This difference was explained by the possible difference between metal requirements of oceanic and neritic species, as the oceanic diatom examined had low Fe, Mn, and Cu quotas and higher

Co and Cd quotas compared to the coastal diatoms. Table 4.44 provides a summary of the quota comparisons by algal class as adapted from Ho et al. (2003).

	Chloro	Prasino	Dino	Соссо	Diatoms			
S/P	5	3	2	4	1			
K/P	5	2	4	3	1			
Mg/P	3	4	2	5	1			
Sr/P	5	3	2	1	4			
Fe/P	1	2	3	4	5			
Mn/P	5	2	3	1	4			
Zn/P	1	2	3	5	4			
Cu/P	2	1	1	5	4			
Co/P	4	3	2	1	3			
Cd/P	4	2	3	1	5			
Mo/P	5	2	1	3	4			

Table 4.23: Interpreted ranking of algae element quotas as depicted in Ho et al. (2003). 1=highest concentration and 5 the lowest.

In the present research, two species each representative of three taxonomic classes (Pavlovacea, Prymesiophycea, and Bacillariophycea) were used as diet variables. *Pavlova* sp. (CCMP1209, Pav1209) was isolated from an unknown tropical site and *Pavlova pinguis* (CCMP609) from the Sargasso Sea. *Isochrysis* sp.1 (CCMP1324, TI) was isolated from the South Pacific off the Society Islands while Isochrysis sp.2 (CCMP1611, ISO) from Chesapeake Bay. Finally, *Chaetoceros galvestonensis* (CCMP186, Cg) was isolated from the Gulf of Mexico off St. Petersburg, FL, and *C. mulleri* (CCMP1316, Cm) from the North Pacific off Hawaii. Originally, two species of *Tetraselmis* were proposed for inclusion in the study, however, trouble maintaining these cultures ultimately lead to their removal from the feeding trials.

Iron has been suggested as a major constituent of shell deposition (Almeida et al. 1998) as well as suggested to be associated with sediment concentrations during 190
experiments with Crassostrea virginica by Carriker et al. (1996). The hypothesis that metabolic/biologic controls or ingestion of particulates could be responsible for iron concentrations observed in the shells of studied bivalves was based upon these previous findings. The Fe/P ratios appeared to suggest a link between Pavlova spp. and the highest concentrations of iron, though the trend was not significant. The comparisons in Ho et al. (2003), unfortunately, did not include specific Pavlovacea species, thus direct comparison is not fully possible as to the potential relationship of iron with this class of algae. However, an increased iron guota and hypothesized subsequent contribution similar to what was expected for the Coccolithophores/Prymnesiophycea is based on similarities between the classes. Of particular interest, analyses of both (Fe/P)/Ca and Fe/P, revealed the ratios from the Cg, ISO, and the CTRL groups as the lowest among the feeding groups, though again not significantly different from the majority of the remaining diet groups. Both the Cg and ISO were neritic isolates and hypothesized to be associated with iron enrichment of the shells based on findings from Sunda and Huntsman (1997), which evidently is the opposite of the current trends observed.

Potassium concentrations were expected to be higher in the shells of clams fed diatom-rich diets based on Ho et al. (2003) as compared to other classes of algae examined. Shells in the *Chaetoceros* treatments did appear to be more enriched with potassium than those from the *Isochrysis spp*. treatments, though not significantly so. The shells from the *Pavlova spp*. treatments exhibited more similar ratios to the *Chaetoceros spp*. treatments. Significant differences, however, were found only between the ratios from the Mixed group and those from the ISO and CTRL groups with

respect to (K/P)/Ca and both the Cm and Mixed groups from ISO with respect to K/P. As such, the observed trends do not support the hypothesis that trends in potassium as normalized to phosphorous can be attributable to specific classes of algae but rather indicate potential species specific influences confounded by nutritional value of the diet and potentially differences between inshore algal strains and nearshore/offshore strains.

Magnesium/phosphorous ratios presented a marginally significant trend with diet. Median ratios for both *Chaetoceros spp.* were higher than those for *Isochrysis spp.* with ratios in shells from the Cm diet significantly higher than both *Isochrysis spp.* When (Mg/P)/Ca ratios were considered, direct dietary influence was not apparent, in fact, the trends indicate growth rate as the most likely dominant influence. These findings were interpreted as suggesting multiple factors being associated with magnesium concentrations. Foster et al. (2008) demonstrated that magnesium was associated with the organic partitions of aragonitic shells and, as such, was related to compositional changes in the organic matter or extra pallial fluid contributing to changes in incorporation rate. In context of the present study, both growth rate and element availability had the potential to influence the composition of the organic matter and therefore element incorporation.

Shells of clams fed the neritic species had lower relative manganese concentrations than those fed the oceanic species, or diets composed of both, as did the CTRL group. This trend, however, was significant only when using Mn/P ratios, though the algal species of unknown origin was not significantly different from the known neritic species. These findings did support the hypothesis that oceanic algal isolates would be associated with higher manganese shell concentrations than the

neritic isolates. Of interest is the low value in the CTRL group, as this indicates potential for metabolic/stress associated changes or the influence of the pre-experimental brownwater diet composed only of neritic algal species. Though associations between manganese and biologic influences have been suggested (Strasser et al., 2008) alternative factors cannot be dismissed. Blanchard and Chasteen (1976) surmised that the amount of substitution of Ca²⁺ by Mn²⁺ was correlated with tidal level, though did not consider the oxygenation of the environment (Rosenberg, 1980). Crisp (1975) attempted to correlate the Mn concentration to salinity, while Strasser et al. (2008) correlated Mn/Ca levels with that of seawater concentration, potentially confounded by biological activity. Carre et al. (2006) suggested positive Mn/Ca association with growth rates while Strasser et al. (2006) found a negative correlation. Most researchers, however, have not considered dietary influence or stress-related influences potentially associated with their observations.

The silica ratios in the shells from clams fed *Pavlova spp.* were consistently higher than in shells from those fed diatoms. Interestingly, shells of clams fed ISO were among the least Si enriched treatments, including when compared to TI. Again, multiple factors are likely associated with these findings, however, the trends do indicate that diet can influence the Si concentration of clam shells.

Zinc-associated ratios appeared to have a similar trend to that observed by Ho et al. (2003) in that Zn concentrations in the shells from diatom treatments were higher than for the Coccolithophores/ Prymnesiophytes treatments. Furthermore, the trends for both Zn ratios were similar, with the shells from the Mixed diet, one or both the diatom diets, and Pav1209 significantly more enriched than those from the ISO group.

The Mixed diet ranked higher for both Zn ratios as compared to the control and Pav609 groups. Based on previous findings associating zinc with dietary factors and metabolic transformations (Windisch, 2001), incorporation of zinc specific to diet as well as metabolic differences amongst clams in the different feeding groups did both likely contribute to the results.

Conclusion

Because the ratios from the ISO group were consistently the lowest in the study, it is suspected that factors not assessed and potential contamination of these shells influenced the results and ultimately complicated interpretations. The results of this study, thusly, are inconclusive with respect to the influence of diet on elemental composition of the shell of *M. mercenaria*. The (Element/P)/Ca and Element/P ratios, while not necessarily capable of being a primary line of evidence, have potential utility in support of findings from standard normalized comparisons, as well as in revealing possible secondary and tertiary influences on elemental shell composition.

5. Temporal changes in the elemental shell chemistry of *Mercenaria mercenaria* during starvation-induced stress and death

Introduction

The composition of fossil shells is generally determined by some combination of the physiochemical environment inhabited by the organism, biologic controls of skeletal growth, and diagenetic alterations (Gomez-Alday and Elorza, 2001). Shell alteration begins while the organism is living and factors that cause shell weathering are among the same factors associated with changes in the shell-deposition metabolism (Rosenberg, 1980). Thus, knowledge of initial shell chemistry is vital in making interpretations as to chemical alterations and subsequent reconstruction of environments (Rosenberg, 1980).

Shell dissolution can occur during periods of anaerobiosis in bivalve molluscs (Crenshaw, 1980) which are characterized by decreased pH and increased succinic acid (Crenshaw and Neff, 1969). Environmental stresses can also cause shell dissolution and ultrastructural changes (Davies and Sayre, 1970; Wilbur, 1972). As discussed in chapters 2 and 3, environmental stresses can ultimately lead to changes in organismal metabolism and changes in chemical composition.

The focus of this chapter is the description of changes in elemental chemistry observed in the shells of starved individuals and that of the dead individuals. The goal is to provide a chronological view of changes potentially occurring in bivalve populations with regard to shell chemistry during events leading to death of the organism and shortly after death.

Methods

The shells analyzed during this research were originally cultured as part of the control group for the feeding experiments described in Chapter 3. A subsample of valves was collected at the start of the starvation period to provide a baseline to compare elemental changes through time with subsequent collections in March, May, July and October. The collected clams were examined during valve separation and notes taken to document the visible changes in health of the animals by use of mantle retraction, tissue occupancy of the shell, gaping, tissue deterioration, and the lack of tissue.

The shells were treated identically as described in Chapter 3 and analyzed for elemental composition using the ICP-OES. Each elemental concentration was again transformed to a ratio as compared to calcium for statistical analysis.

The element/Ca levels were initially graphed by collection month to determine the general trends associated with each element. The data were then examined by collection month using analysis of variance or Kruskall-Wallis one-way analysis of variance on ranks followed by an appropriate post hoc analysis as in the previous chapters.

Results

No dead clams were observed until July though mantle retraction, small tissue volumes and some intermediate gaping was observed during March with increased signs of stress noted in both May and July. The July collection was the first time empty valves and deteriorating tissue were observed. All the animals in the October collection were dead, with the majority of shells void of tissue and only a few with small remnants of decomposing tissue apparent.

Due to unsuccessful analysis of specific elements for some of the collection groups, the analyses were confined to B, Ba, Co, Fe, K, Li, Mg, Mn, Si, and Zn. The results herein thus focus on these elements as compared to Ca and changes in those ratios over the course of the experiment.

The B/Ca ratios by collection month (Figure 5.1, Tables 5.1 and B1) revealed a higher relative boron concentration in July compared to all other collections. The only other significant difference was observed between the March and October collections.

The Ba/Ca ratios (Figure 5.2, Tables 5.2 and B2) indicate significant loss of Ba from the shells between January and March. The ratios thereafter, however, did not significantly change. Of note was the apparent decrease in deviation from the median values through the course of the experiment.

Cobalt concentrations as compared to calcium again were associated with a general downward trend to the July collection with a significant increase observed in October (Figure 5.3). The Co/Ca ratios in the shells from July were significantly lower than those from the baseline (January) and March collections (Tables 5.3 and B3).

There is a large degree of uncertainty as to the amount of change between January and July as the May median is based upon of a single clam.



Figure 5.1: Median B/Ca compared to month collected with associated 95% confidence intervals.

One Way Analysis of Variance								
Normality Test:	Failed	(P < 0.050)						
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Missing Median 25% 75%					
January	14	1	2.86E-05	2.71E-05	3.17E-05			
March	16	1 2.92E-05 2.78E-05 4.87E-						
Мау	11	1	1 2.43E-05 2.19E-05					
July	21	1	6.70E-04	3.80E-05	7.91E-04			
October	16	1 2.20E-05 2.14E-05 2.60E-05						
	H = 46.0	with 4 degrees of	freedom (P = <0.001)					

Table 5.1: Summary of analysis of variance on ranks for B/Ca.

The Fe/Ca ratios exhibited a significant decrease between the baseline (January) and both the March and May collections followed by a significant increase between the May and October collections (Figure 5.4, Tables 5.4 and B4). The ratios of the shells from the March and May collections were not significantly different, nor were the ratios

of the shells from the July and October collections. Furthermore, the ratios from the baseline collection and those from the July and October collections were not significantly different.



Figure 5.2: Median Ba/Ca compared to month collected with associated 95% confidence intervals.

One Way Analysis of Variance								
Normality Test:	Failed		(P <	0.050)				
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Median	25%	75%			
January	14	1	2.21E-05	1.83E-05	2.66E-05			
March	16	1	1.49E-05	1.29E-05	2.02E-05			
Мау	11	1	1 1.28E-05 1.14E-05 1.49E					
July	21	1	1.17E-05	1.13E-05	1.41E-05			
October	16	1	1.36E-05	1.29E-05	1.51E-05			
	H = 33.1	with 4 degrees of f	freedom (P = <0.001)					

Table 5.2: Summary of analysis of variance on ranks for Ba/Ca.



Figure 5.3: Median Co/Ca compared to month collected with associated 95% confidence intervals.

One Way Analysis of Variance								
Normality Test:	Passed	(P = 0.742)						
Equal Variance Test:	Passed	(P = 0.182)						
Group Name	N	Missing	Mean	Std Dev	SEM			
January	12	7	1.44E-06	3.59E-07	1.61E-07			
March	13	8	1.26E-06	2.05E-07	9.18E-08			
Мау	11	10	7.60E-07	0.00E+00	0.00E+00			
July	20	14	6.99E-07	1.39E-07	5.68E-08			
October	14	11	2.00E-06	3.33E-07	1.92E-07			
Source of Variation	DF	SS	MS	F	Р			
Between Groups	3	3.67E-12	1.22E-12	18.32	<0.001			
Residual	15	1.00E-12	6.68E-14					
Total	18	4.68E-12						
	Power of per	formed test with alpha	= 0.050: 1.000					

Table 5.3: Summary of analysis of variance for Co/Ca.

The median K/Ca ratios over time (Figure 5.5) revealed a significant decrease between both the baseline (January) and March collections, and between the March and July collections (Tables 5.5 and B5). A further decrease was revealed between the

May and July collections. Again, the variability observed decreased through the experiment.

The temporal comparison of Li/Ca (Figure 5.6) revealed a gradual decrease in relative Li concentrations of the shells. The Li/Ca ratios of the clam shells from the January collection were higher than those from the July and October collections (Tables 5.6 and B6). The ratios of the shells from the March and May collections were also revealed to be higher than those from October. The variability of Li/Ca ratios by collection again decreased through the experiment.



Figure 5.4: Median Fe/Ca compared to month collected with associated 95% confidence intervals.

One Way Analysis of Variance								
Normality Test:	Failed	(P < 0.050)						
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Median	25%	75%			
January	14	1 8.01E-05 5.73E-05 7.70E-04						
March	16	1	2.01E-05	1.28E-05	4.12E-05			

Table 5.4: Summary of analysis of variance on ranks for Fe/Ca.

Table 5.4 (Continued)							
Group	Ν	Missing	Median	25%	75%		
Мау	11	1	2.16E-05	1.90E-05	2.39E-05		
July	21	1	6.17E-05	4.61E-05	7.70E-05		
October	16	1	6.25E-05	4.53E-05	7.00E-05		
H = 41.4 with 4 degrees of freedom (P = <0.001)							





One Way Analysis of Variance									
Normality Test:	Failed		(P < 0.050)						
Kruskal-Wallis One Way Analysis of Variance on Ranks									
Group	N	Missing	Median	25%	75%				
January	14	1	3.40E-03	3.72E-06	4.72E-03				
March	16	1	2.62E-03	1.57E-03	3.23E-03				
Мау	11	1	1 1.15E-03 9.58E-04 1						
July	21	1	1.51E-05	9.83E-06	2.25E-05				
October	16	1	1 2.05E-04 1.58E-04 2.46E-04						
	H = 41.2	with 4 degrees of f	reedom (P = <0.001)						

Table 5.5: Summary of analysis of variance on ranks for K/Ca.



Figure 5.6: Median Li/Ca compared to month collected with associated 95% confidence intervals.

One Way Analysis of Variance								
Normality Test:	Failed		(P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Median	25%	75%			
January	14	1	4.03E-06	3.86E-06	9.14E-04			
March	16	1 3.29E-06 2.91E-06 4.29E-0						
Мау	11	1 3.16E-06 2.92E-06 3.60E-06						
July	21	1 2.83E-06 2.16E-06 3.32E-04						
October	16	1 2.19E-06 2.01E-06 2.27E-06						
H = 37.5 with 4 degrees of freedom (P = <0.001)								

Table 5.6: Summary of analysis of variance on ranks for Li/Ca.

The Mg/Ca trend revealed an increase in relative Mg concentration between the baseline and March collections followed by a decrease in Mg concentration (Figure 5.7). The Mg/Ca ratios were significantly higher in the shells from the March collection compared to those of the January, July and October collections; while Mg/Ca ratios of the shells from the May collection were significantly higher than those of the July and

October collections (Tables 5.7 and B7). Of note, the only significant differences from baseline ratios were those from the March collection.





The Mn/Ca ratios revealed a decrease in relative Mn concentration of the shells followed by a small increase in the shells of the October collection (Figure 5.8), though all changes appeared to be within the deviation of the baseline (January) collection. Statistical analyses (Tables 5.8 and B8) revealed that Mn/Ca ratios were higher in the shells from the baseline collection than those from the shells in the May, July, and October collections. In addition, March Mn/Ca ratios were higher than May and July.

· ••••••• • ••••••••••••••••••••••••••								
One Way Analysis of Variance								
Normality Test:	Failed	ailed (P < 0.050)						
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Missing Median 25% 75%					
January	14	2	1.27E-03	5.35E-04	1.41E-03			

Table 5.7: Summary of analysis of variance on ranks for Mg/Ca.

Table 5.7 (Continued)								
Group	Ν	Missing	Median	25%	75%			
March	16	1	2.89E-03	1.87E-03	3.35E-03			
Мау	11	1	1.66E-03	1.30E-03	1.89E-03			
July	21	1	5.80E-04	5.19E-04	6.51E-04			
October	16	1	5.91E-04	5.13E-04	7.31E-04			
H = 43.8 with 4 degrees of freedom (P = <0.001)								



Figure 5.8: Median Mn/Ca compared to month collected with associated 95% confidence intervals.

One Way Analysis of Variance								
Normality Test:	Failed		(P <	0.050)				
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Median	25%	75%			
January	14	2	5.01E-05	3.18E-05	2.00E-04			
March	16	1	2.32E-05	2.17E-05	3.01E-05			
Мау	11	1	1 1.20E-05 9.44E-06 1.					
July	21	1	1.23E-05	1.02E-05	1.80E-05			
October	16	1	2.02E-05	1.77E-05	2.21E-05			
	H = 47.3	with 4 degrees of f	freedom (P = <0.001)	I				

Table 5.8 [.]	Summarv	of analys	is of var	iance on	ranks for	Mn/Ca
1 4010 0.0.	Ourmany	or analys	3 01 101		1011113 101	win / Oa.

The Si/Ca ratios over time were more variable than the elements previously discussed (Figure 5.9). The statistical analyses (Tables 5.9 and B9) revealed a significant decrease of Si/Ca ratios in the shells collected between January and May and that relative concentrations of silica were higher in shells collected in both July and October as compared to those collected in March and May.



Collection

Figure 5.9: Median Si/Ca compared to month collected with associated 95% confidence intervals.

One Way Analysis of Variance								
Normality Test:	Failed		(P < 0.050)					
Kruskal-Wallis One Way Analysis of Variance on Ranks								
Group	N	Missing	Median	25%	75%			
January	10	1	4.17E-04	2.93E-04	5.26E-04			
March	16	1 1.65E-04 1.45E-04 2.64						
May	11	1	1 1.55E-04 1.26E-04					
July	21	1	5.09E-04	3.70E-04	6.31E-04			
October	16	1 3.82E-04 3.52E-04 5.24E-04						
	H = 35.7	with 4 degrees of f	reedom (P = <0.001)					

Table 5.9: Summary	of anal	ysis of	variance	on ranks	for	Si/Ca.
--------------------	---------	---------	----------	----------	-----	--------

The trend in Zn/Ca by collection month (Figure 5.10) was a small increase in median Zn/Ca ratios of the shells collected between January and March followed by a gradual decrease through July followed by a more dramatic decrease between July and October. The Zn/Ca ratios of the shells from both January and March collections were significantly higher than those from the July and October collections (Tables 5.10 and B10). Thusly, a significantly decreased concentration of zinc was observed from baseline to the final experimental collection.



Figure 5.10: Median Zn/Ca compared to month collected with associated 95% confidence intervals. The October measurements were below the concentration of the standard.

One Way Analysis of Variance										
Normality Test:	Failed	(P < 0.050)								
Kruskal-Wallis One Way Analysis of Variance on Ranks										
Group	N	Missing	Median	25%	75%					
January	14	2	2.00E-05	1.61E-05	2.80E-05					
March	16	2	2.65E-05	1.87E-05	4.07E-05					
Мау	11	1	1.15E-05	8.27E-06	1.45E-05					
July	21	10	4.09E-06	3.06E-06	4.91E-06					
October	13	7	-2.22E-04	-2.75E-04	-1.45E-04					
H = 39.7 with 4 degrees of freedom (P = <0.001)										

Table 5.10: Summary of analysis of variance on ranks for Zn/Ca.

Discussion

The current study demonstrates the changes in 10 element/calcium ratios through time for starved *Mercenaria mercenaria*. These results illustrate both changes due to assumed metabolic stress and alterations occurring post mortem which provides valuable insight into the mechanisms of shell dissolution and utility of specific elements in paleontological analyses aimed at environment reconstruction. The importance of knowledge pertaining to incorporation pathways, organic versus inorganic partitioning, and shell dissolution mechanisms is emphasized.

Boron concentrations as compared to calcium of the collected shells were not found to be significantly lower than the baseline concentrations; instead, there was a dramatic increase in the Ba/Ca of the shells from the July collection. Studies of mollusc shells by Furst et al. (1976) suggested that boron was associated with the inorganic shell fraction and that it appeared to substitute for carbon. The substitution of carbon by boron was later supported during analysis performed by Cook (1977). It was noted, though, that effects of other factors such as temperature and nutrition were not considered (Rosenberg 1980). Additionally, the results of Hemming and Hanson (1992) demonstrated an association between pH and boron incorporation in carbonates, however, the findings were more specific to isotopic ratios. The results of the current study are not ultimately explained by incorporation of boron into the shell void of adequate carbon levels. However, association between the changes in relative concentration of boron in the collected shells and assumed metabolic changes due to starvation stresses cannot be dismissed.

Barium concentrations as compared to calcium were associated with a general decrease over time as compared to the baseline concentration, though a small increase associated with dead shells was observed. The factors associated with incorporation of barium into bivalve shells are still in debate, but recent evidence in Tabouret et al. (2012) with regard to *Pecten maximus* suggests Ba to be directly incorporated into the shell via dissolved phases and in proportion to the concentrations observed in the surrounding waters. This conclusion supported that given in Gilliken (2006) which determined the Ba concentrations observed in the shells of *Mytillus edulis* to be directly associated with [Ba/Ca]_{Water} and not determined by diet or solely/primarily influenced by phytoplankton blooms, thus salinity extrapolations could be made using Ba/Ca from archival shells. Seemingly missing from the discussions of these studies is the potential for change when the target bivalve species is under stress. The present study identified a gradual decrease in Ba/Ca in the shells of starved *M. mercenaria* with a significant decrease from baseline. The gradual loss observed in this study was potentially related to shell dissolution and lost metabolic controls due to starvation/stress, as such, these results suggest that Ba/Ca may not be consistently preserved nor dependable for environmental interpretations. The partitioning of Ba between the mineral and organic shell fractions, however, is an important consideration in determining the effectiveness of Ba as an environmental proxy.

Cobalt concentrations as compared to calcium were associated with a general downward trend to the third experimental collection with a sharp increase observed for the last collection. Cobalt has been a focus of research generally specific to environmental contamination studies (for example, Byron et al., 1978; Szefer et al.,

1990; Hedouin et al., 2006; Rainbow et al., 1993). In a study conducted by Babukutty and Chacko (1992), cobalt was determined to be mostly associated with the shells of *Villorita cyprinoides*; however, Cravo et al. 2004 suggested Co was partitioned in the soft tissues of *Patella aspera*. In the current study, with recognition of potential differences between phyla and mineralogical differences, the observed trend in Co as compared to calcium is seemingly attributable to shell dissolution and organic matter decomposition mechanisms, as well as indicating a possible higher concentration in the organic matter.

Iron concentrations exhibited an initial decrease from baseline levels but were determined to be within the range of baseline values during the last two collections. Iron has been described as an essential component of shell deposition often associated with the organic matrix (Almeida, 1998; Wang et al. 2003; Zhang et al., 2003), and has been suggested to more easily incorporated into soft tissues than in shells (Cravo et al., 2004). Conversely, iron has been associated with the shell and soft tissues of bivalves due to metal contamination (Pitts et al., 1994; Bryan et al., 1978). The trends observed in the current study were probably influenced by ratio alteration due to shell dissolution and preservation of organic fractions.

The relative concentrations of potassium were observed to follow a downward trend with a small, but insignificant, increase between collections four and five. Potassium has been suggested be incorporated through ingestion of particulates (Carriker et al., 1996). The downward trend was potentially associated with shell dissolution and the increase potentially signifying organic content being retained (Lutz and Rhoads, 1977).

An overall decrease in Li/Ca was observed in the present study. Though incorporation dynamics have been associated with plankton and water concentration dynamics (Thebault et al., 2009; Thebault, 2012), the decreased concentrations were determined to be associated with loss due to shell dissolution.

The trend observed regarding Mg/Ca as compared to collection suggested an initial increase in relative concentration followed by a downward trend. Magnesium has been shown to substitute for calcium though primarily into calcite rather than in aragonite (Clarke and Wheeler, 1922; Dodd, 1965). Overall, influences attributable to magnesium incorporation have the potential to be highly variable (Rosenberg, 1980). As such the initial increase in magnesium in the current study could be due to replacement of calcium followed by effects of shell dissolution.

The trend in Mn/Ca over time showed a decreased relative concentration followed by a small increase in the last collection. Manganese has been described as being partitioned in both organic and mineral phases of the bivalve shell (Comfort, 1951; Fox, 1966) as well as capable of substituting for calcium in biogenic carbonates (White et al., 1977; Blanchard and Chasteen, 1976). Overall, the decrease seems to have suggested loss through shell dissolution followed by a potential ratio shift due to retained organic-phase associations.

Silica concentrations relative to calcium decreased initially then rebounded to within baseline ranges. Silica has been shown to be involved with early diagenetic transformations in association with microbial activity and decay of organic matter (Holdaway et al., 1982). These described transformations, however, were aligned with specific sediment environments. The present results were not influenced by diagenetic

effects isolated during burial but were equivalent to changes occurring prior to burial if occurring in the natural environment. The Si/Ca associated trends are most likely due to shell dissolution, with potential influence from microbial activity.

Zinc showed a general decrease over time in the present study, though the concentrations determined for collection four were below the ICP standard concentrations, thus the actual degree of change is not known. As discussed in previous chapters, zinc is often discussed in terms of environmental contamination, though Windisch (2001) did specify zinc as important biologically and associated with metabolic and food web interactions. As shown in Chapter 3, zinc appeared to be associated with metabolic change. As such, the decrease in zinc observed in the current study is likely attributable to both metabolic shifts and shell dissolution alterations.

Conclusion

In summary, Ba, Co, Fe and Si were higher relative to Ca in shells at the start of the experiment then declined as the animals starved, with some increase following the death of the clams. Potassium, Li, Mg, and Mn relative to Ca were high in the baseline shell group then declined at somewhat different rates through the course of the experiment. Zinc to Ca ratios remained high in the live or recently dead shells (through July), but then dropped, with high variability, in the dead shells analyzed from the final collection. Only B, which as described above, tends to substitute for C, appeared to behave very differently relative to Ca, compared to the trends seen in the other trace

elements analyzed. If the major factor in the elemental changes was dissolution, boron could be expected to behave somewhat differently.

6. Conclusions

Recent research specific to the bivalve shell has been predominantly focused on associating changes in elemental concentrations with environmental parameters such as temperature, salinity, seasonal variability in water concentrations, pollution, and effluent river waters. The influences of phytoplankton dynamics (Stecher et al., 1996) or their elemental composition (e.g. Gilliken, 2005) have only recently been suggested as potential factors in the variability of bivalve shell chemistry. Barium and lithium have been the primary elements of interest with regard to phytoplankton associations, though the majority of the studies allude to phytoplankton bloom dynamics as being attributed to elemental signals observed. Barium correlated with phytoplankton blooms (diatoms mostly) in several studies (Stecher et al., 1996; Putten et al., 2000; Thebault et al., 2009; and Lazareth et al., 2007), which was further developed in Tabouret et al. (2012) where Ba enrichment was determined to be most likely due to incorporation of dissolved Ba, which as might be available following an algal bloom (Stecher and Kogut, 1999). Research presented in Gilliken et al. (2006) demonstrated an influence of [Ba/Ca] water on [Ba/Ca] shell of Mytillus edulis and they concluded that there was no direct association between Ba/Ca enrichment and dietary uptake. The influence of phytoplankton blooms on [Ba/Ca]_{water}, however, were later suggested as being a possible explanation for Ba peaks observed and that feeding on algal species with high Ba concentrations is

another plausible influence based on examination of Ba concentration in the gut, though the degree of dietary Ba incorporation was not discussed. With respect to Li/Ca, Thebault and Chauvaud (2011) examined signatures in *Pecten maximus* with the conclusion that growth, potentially temperature and increased lithium due to diatom blooms were responsible for enrichment. Interestingly, though it has been suggested that algal elemental compositions are a plausible influence on the elemental composition of bivalve shells, confirming information remains scarce.

The goal of this dissertation was to assess the influence of diet, specifically consumption of algal taxa, on the elemental chemistries of the shells of two marine bivalve mollusks. Overall, differences were observed between the two bivalve species, as well as among treatments within species. However, experimental design and logistical challenges are suspected to have influenced the results and thereby limited interpretations. Because whole shell samples were examined, including the organic matrix, results were influenced by both the mineral phase and organic phase. Additionally, ontogenetic influences cannot be compared because the ages and sizes of the experimental scallops and clams were different. Algal cultures produced insufficient concentrations for adequate feeding during the last months of the *Mercenaria* experiments. And finally, complications during elemental analysis restricted sample sizes and the elements reported. Nevertheless, the dissertation yielded the following conclusions:

- 1. Chapter 2 experiments with Argopecten irradians concentricus revealed:
 - highest growth rates and survivorship in treatments fed *Pavlova* pinguis or mixed algal diets,

- b. no differences in shell chemistry between the left and right valves,
- c. significant differences in shell chemistries for only two elements (Mg and K) between feeding groups, and
- d. only K in the shells appeared to be directly influenced by diet.
- 2. Chapter 3 experiments with Mercenaria mercenaria revealed that:
 - a. axial growth and shell thickening were variable among treatments, with only the Mixed and Cm treatments exhibiting significant growth, though experimental procedure prevented true growth determinations;
 - b. significant differences in shell chemistry between the left and right valves, with the higher ratios in the right valve;
 - c. several elemental ratios were significantly different between feeding groups, including Fe, Mn, and P;
 - Mg/Ca and Sr/Ca ratios appeared to be primarily influenced by growth rates, though pre-experimental growth and biologic factors not assessed also probably contributed to differences observed;
 - e. Zn/Ca and B/Ca were likely influenced by stress and metabolic changes;
 - f. Ba/Ca, K/Ca, Li/Ca and Si/Ca were likely influenced by factors not addressed in the experimental design; and
 - g. in both scallop and clam shells, Mg/Ca was directly related to growth rate, with diet as a contributing factor as nutritional value of the algal species or algal mix was directly associated with growth rate.

- Normalizing elemental ratios to phosphorus did not enhance resolution of trends in ratios among dietary treatments.
- 4. Temporal assessment of shell chemistry in clams starved for the duration of the experiment indicated changes associated with shell dissolution.

The development of inter-valve differences in the clam experiment was primarily attributed to assumed metabolic changes associated with stress/starvation of the animals, potentially coupled with genetic controls. This association was further supported during diet group comparisons, where relative Zn and B concentrations were determined likely related to stress as exemplified in the starved clams. A starvation control, however, was not established during the scallop trials, thus a comparison between species cannot be made. Interestingly, the differences between clam left and right valves revealed a higher ratio for the right valve, which follows the observations of Carriker et al. (1996) in that right valves of *Crassostrea virginica* were elementally enriched compared to the left valve. This suggests a common genetic control which is emphasized in equivalved bivalves during times of stress.

In both scallop and clam shells, Mg/Ca was most directly attributable to growth rate, but seemingly was confounded by other biologic controls. This association supports previous findings associated with multiple bivalve species (e.g., Carre et al., 2006; Gilliken et al., 2006; and Strasser et al., 2008). The results of this study, however, include Mg in the organic fraction as well as potential influences of shell dissolution. Diet, none-the-less, is a contributing factor as nutritional value of the algal species or algal mix has been demonstrated to be directly associated with growth.

The identification of different elements being attributable to algal diet for the scallops (K/Ca) and clams (Fe/Ca, Mn/Ca, and P/Ca) indicates a potential species specific effect. A difference between the experiments, however, may also have contributed to differences in interpretation. Prior to the experiment, the clams were moved to a brown water system, which increased chances for influence of sediment on K enrichment as proposed by Carriker et al. (1996). This complication was eliminated from the scallop trials. The potential for this or alternative pre-experimental factors influencing the differences revealed in the K/Ca of the clam shell is more plausible than algal diet. Both possibilities, however, suggest K incorporation by ingestion of enriched particles. Partitioning of K in diatoms, the algal class having the highest average K/P (Ho et al., 2003), as well as the digestibility of the cells, may also be inhibiting translation of the algal signal to the shell.

Manganese, copper, zinc, and cadmium have been shown to be primarily associated with the organic matrix in oyster shells (Carriker et al., 1980), as was iron (Almeida et al., 1998) and magnesium in mussel shells (Lorens and Bender, 1980). This association of specific elements with the organic matrix suggests a high level of biologic control. In the present clam study, iron, manganese, and phosphorous were demonstrated to be directly influenced by diet. Because the organic matrix is responsible for shell strength (Addadi et al., 2006), the organic matrix potentially influences elemental content and structure of the shell (Watabe et al., 2001; Takesue and van Green, 2004; and Johnstone, 2008). If diet associated elements are incorporated into the organic matrix and can be manipulated by dietary changes as demonstrated, diet of cultured bivalves could potentially be implicated in differences in

shell strength and structure between wild and aquacultured animals such as those described in Gresfrude and Strand (2006) regarding *Pecten maximus*. At the very least, the demonstrated dietary influence of specific algal classes on shell chemistry demands consideration of diet composition when conducting experiments aimed at describing influences on shell dynamics or determining cause and effect relationships. The elements influenced and subsequent effects of the alteration, however, are most probably species specific, as are the pathways of element incorporation.

The variations in Ba/Ca and Li/Ca in the clam experiments cannot be attributed to the experimental diets. Instead, both ratios indicate pre-experimental factors as the dominant influence of the differences revealed. Though partitioning of Ba in diatoms and digestibility again comes into question, the revealed difference between the right and left valves from the Cm diet seemingly supports direct incorporation as might occur at the sediment-water interface as previously suggested by Tabouret et al. (2012).

Though diet was shown to directly affect the shell chemistry of *Mercenaria mercenaria*, resolution of the algal contribution was limited to taxonomic class with regard to only a few elements, and differences related to the origin of the algal isolate were not apparently translated to the shells as had been hypothesized. The lack of resolution between clams fed different algal diets may, in part, be due to the taxonomic classes of algae used and similarity in element quotas with respect to specific elements. Other explanations include, a. partitioning of the elements in the algal cell or digestibility of the cell inhibiting transfer of elements in concentrations high enough to produce an associated signal, b. incorporation pathway in clams does not support dietary uptake as a major contributor, and c. multiple influences are associated with the elemental

dynamics with any dietary influence being masked. In an attempt to increase the ability to discern the contribution of specific species of algae to shell chemistry and gain insight as to potential cofactors influencing elemental signatures, experimental analyses using Element/P and (Element/P)/Ca were employed.

Several problems were identified with comparing the elemental concentrations to phosphorus, which include variance in P concentrations potentially not being uniform and attributable to the same factor in each group, high P concentrations contributing to unrelated minimal ratios with specific diets, and potential confounding factors outside controlled environments as identified in previous studies. Though there was a greater degree of separation apparent between species in the same taxonomic class, most differences observed were not significant; furthermore, the differences between the neritic and oceanic diets were not translated to the shells, in fact, the results were opposite of those hypothesized. Additionally, because the ratios from the ISO group were consistently revealed as the lowest in the study, factors not assessed and potential contamination of these shells influenced the results, ultimately complicating interpretations. In short, the results of this study are inconclusive. These ratios might be used in succession with currently used methods to provide multiple lines of evidence ultimately helping to further describe element incorporation and associations between controlling influences. However, more research is required and measures must be taken to ensure P concentrations are relatively stable and influenced by a single factor.

The inspection of shell chemistry through time of clams starved for the duration of the experiment showed that, in general, elemental concentration as compared to calcium decreases as the animal approaches death or is under stress. The main

factors are seemingly due to the mechanics of shell dissolution. The results of this study appear to support the retention of original shell elemental chemistry for some elements. However, elements easily absorbed to the shell surface, and those associated with metabolic influences or with the organic fractions of shells, will deviate from initial shell chemistries when compared with calcium as a function of shell dissolution mechanics thus are unreliable as environmental proxies. Continued analysis of incorporation pathways and determination of the degree to which an element is partitioned between the mineral and organic portions of the shell for individual bivalve species is again recommended before environmental factors are associated with elemental composition of archival shell.

The mechanisms of shell deposition and factors controlling elemental incorporation are complex and influenced by many factors. The results of this dissertation demonstrate that specific algal diets can influence elemental shell chemistry both directly and indirectly. Thus, diet should be considered when interpreting shell chemistry during both laboratory and field experiments. Though further research is needed, algal diet may influence shell chemistry in ways that hold promise for developing proxies of ecological shifts associated with changes in phytoplankton composition.

Recommendations for future work

 Laboratory work similar to Ho et al. (2003) is needed to address element quotas on a species-specific basis for algal species commonly used in aquaculture and commonly ingested by bivalves. Associations between shell elemental

composition and diet cannot be definitively made unless the differences between algal species are known.

- Further examination of dietary influence on shell dynamics for multiple species of bivalves exhibiting different shell microstructural properties and life histories is necessary to determine species-specific influences and commonalities among similar species. Experimental design should allow for uniform growth and analysis of shell fractions separately in conjunction with ontogenetic examinations.
- Fracture analysis on subsamples of shells with known diet is necessary to determine the effects diet can have on the strength of the shell.
- Microstructural analyses on fractured shells are needed to determine differences based on elemental chemistry and dietary influences. This will further increase knowledge as to the influence organic matter has on the microstructure and elemental composition of the mineral partition.
- Determine algae combinations for specific study areas to allow differentiation between the cultured animals and wild populations, then test length of retention of the signature. For a marker to be useful, it must be retained long enough to provide the resolution needed as well be distinguishable from environmental variability.
- Perform feeding experiments with controlled environmental changes to further determine influences potentially confounding dietary influences or that are dominant over dietary influence.

- Increased experimentation with local species of bivalve and algae to catalogue potential signatures that can then be examined in the field. Upon these determinations, the use of shell for ecological association will be possible given ability to differentiate between the signatures and natural variability.
- Use information obtained from feeding experiments and microstructural analysis to determine capability of manipulating shell structure via diet. This will potentially allow shell strength engineering or pearl composition engineering by a low cost mean.

References

Addadi, L., Joester, D., Nudelman, F., Weiner, S., 2006, Mollusk shell formation: a source of new concepts for understanding biomineralization processes, *Chemistry* 12(4): 980–987.

Al-Aasm, I. S., and Veizer, J., 1986, Diagenetic stabilization of araqonite and low-Mg calcite, 1. Trace elements in rudists, *Jour. Sed. Petrology* 56: 138–52.

Albentosa, M., Perez-Camacho, A., Labarta, U., Beiras, R., Fernandez-Reiriz, M. J., 1993, Nutritional value of algal diets to clam spat Venerupis pullastra, *Mar. Ecol. Prog. Ser.* 97:261-269.

Almeida, M. J., Moura, G., Machado, J., Coimbra, J., Vilarinho, I., Ribiero, C., Soares-de-Silva, P., 1996, Amino acid and metal content of Crassostrea Gigas shell infested by Polydora sp in the prismatic layer insoluble matrix and blister membrane, *Aquatic living Resour.* 9: 174-186.

Almeida, M.J., Machado, J., Moore, G., Azevedo, M., Coimbra, J., 1998, Temporal and local variations in biochemical composition of Crassostrea gigas shells, *Journal of Sea Research* 40(3-4): 233-249.

Alonso, D. L., Belarbi, E. H., Rodríguez-Ruiz, J., Segura, C. I., Giménez, A., 1998, Acyl lipids of three microalgae, *Phytochemistry* 47(8): 1473-1481.

Amiel, A. J., Friedman, G. M., Miller, D. S., 1973, Distribution and nature of incorporation of trace elements in modern aragonitic corals*, *Sedimentology 20*(1): 47-64.

Anderson, M. A., and Morel, F. M. M., 1982, The influence of aqueous iron chemistry on the uptake of iron by the coastal diatom *Thalassiosira weissflogii*, *Limnol. Oceanogr*. 27: 789–813.

Annett, A. L., Lapi, S., Ruth, T. J., Maldonado, M. T., 2008, The effects of Cu and Fe availability on the growth and Cu:C ratios of marine diatoms, *Limnol. Oceanogr.* 53: 2451–2461.

Ansell, A.D., 1968, The rate of growth of the hard clam *Mercenaria mercenaria* throughout the geographical range, *J. Cons. Perm. Int. Explor. Mer.* 31: 364–409

Babukutty, Y., and Chacko, J., 1992, Trace metals in an esturine bivalve from the southwest coast of India, *AMBIO*. 21(4): 292-296.

Bayne, B. L., (1993), Feeding physiology of bivalves: Time dependence and compensation for changes in food availability, in: Bivalve filter feeders in estuarine and coastal ecosystem process (R. F. Dame, ed.), pp. 1-24, *NATO ASI Ser.V. G33*. Springer.

Becker, B. J., Levin, L., Fodrie, J. F., McMillan, P.A., 2005, Spatial and temporal variation in trace elemental fingerprints of mytilid mussel shells: a precursor to invertebrate larval tracking, *Limnol. Oceanogr.* 50(1): 48-61.

Becker, B. J., Levin, L., Fodrie, J. F., McMillan, P.A., 2007, Complex larval patterns among marine invertebrate populations, *Proc. Natl. Acad. Sci. USA*

Bevelander, G., and Nakahara, H., 1969, An electron microscopic study of the formation of the nacreous layer in the shell of certain bivalve molluscs, *Calcif. Tissue Res.* 3:84-92.

Bevelander, G., and Nakahara, H., 1980, Compartment and envelope formation in the process of biological mineralization, in: The Mechanisms of Biomineralization in Animals and Plants (M. Omari and N. Watabe, eds.), pp 19-27, Tokai Univ. Press.

Blanchard, S.C., and Chasteen, N.D., 1976, Electron paramagnetic resonance spectrum of a seashell, *J. Phys. Chem.* 80: 1362-1367.

Blank, S., Arnoldi, M., Khoshnavaz, S., Treccani, L., Kuntz, M., Mann, K., Grathwohl, G., Fritz, M., 2003, The nacre protein perlucin nucleates growth of calcium carbonate crystals, *J. Microsc.* 212: 280–291

Boggild, O.B., 1930, The shell structure of the mollusk, *K. Dan. Vidensk. Selsk. Skr.* Copenhagn 2: 231-326

Bourgoin B. P., 1990, Mytilus edulis shell as a bioindicator of lead pollution: Considerations on bioavailability and variability, *Mar.Ecol. Prog. Ser.* 61: 253–262.

Brand, L. E., 1991, Minimum iron requirements of marine phytoplankton and the implications for the biogeochemical control of new production, *Limnol. Oceanogr.* 36: 1756–1771.

Brand, L. E., Sunda, W. G., Guillard, R. R. L., 1983, Limitation of marine phytoplankton reproductive rates by zinc, manganese and iron, *Limnol. Oceanogr.* 28: 1182–1198.

Brand, L. E., Sunda, W. G., Guillard, R. R. L., 1986, Reduction of marine phytoplankton reproduction rates by copper and cadmium, *J. Exp. Mar. Biol. Ecol.* 96: 225–250.

Brockington, S., and Clarke, A., 2001, The relative influence of temperature and food on the metabolism of a marine invertebrate, *J. Exp. Mar. Biol. Ecol.* 258:87–99

Brookes, R. R., and Rumsby, M. G., 1965, The biochemistry of trace element uptake by some New Zealand bivalves, *Limnol. Oceanogr.* 10(4): 521-527.

Brown, M. R., 2002, Nutritional value of microalgae for aquculture, in: Avances en Nutrición Acuícola VI. Memorias del VI Simposium Internacional de Nutrición Acuícola (Cruz-Suárez, L. E., Ricque-Marie, D., Tapia-Salazar, M., Gaxiola-Cortés, M. G., Simoes, N., eds.), Cancún, Quintana Roo, México.

Brown, M. R., Dunstan, G. A., Norwood, S. J., Miller, K. A., 1996, Effects of harvest stage and light on the biochemical composition of the diatom *Thalassiosira pseudonana*, *Journal of Phycology* 32(1): 64-73.

Brown, M. R., Jeffrey, S. W., Volkman, J. K., Dunstan, G. A., 1997, Nutritional properties of microalgae for mariculture, *Aquaculture 151*(1): 315-331.

Brown, S. L., 1986, Feces of intertidal benthic invertebrates: influence of particle selection in feeding on trace element concentration, *Mar. Ecol. Prog. Ser.* 28:219-231.

Bryan, G. W., 1973, The occurance and seasonal variation of trace metals in the scallops Pecten maximus and Chlamys opercularis, *J. Mar. Biol. Assoc. U.K.* 53:145-166.

Bryan, G. W., 1976, Some aspects of heavy metal tolerance in aquatic organisms, In: Effect of Pollutants in Aquatic Organisms (A. P. M. Lockwood, ed.), pp. 7-34.

Bryan, G. W., 1980, Recent trends in research on heavy-metal contamination in the sea, Helgoländer Meeresunters 33: 6–25.

Bryan, G. W., and Hummerstone, L. G., 1978, Heavy metals in the burrowing bivalve Scrobicularia plana from contaminated and uncontaminated estuaries, *J. mar. biol. Ass. U. K.* 58: 401–419.

Byrne, R. H., Kump, L. R., Cantrell, K. J., 1988, The influence of temperature and pH on trace metal speciation in seawater, *Mar. Chem.* 25: 163–181.

Carefoot, T.H., and Donovan, D.A., 1995, Functional significance of varices in the muricid gastropod Ceratostoma foliatum, *Biol. Bull.* 189: 59–68.
Carre, M., Bentalab, I., Bruguier, O., Ordinula, E., Barrett, N.T., Fontugne, M., 2006, Calcification rate on trace element concentration in aragonitic bivalve shells: evidences and mechanisms, *Geochimica et Cosmochimica Acta* 70: 4906-4920

Carriker, M.R., 1961, Interrelation of functional morphology, behavior and autoecology in early stages of the bivalve Mercenaria mercenaria, J. Elisha Mitchell Sci. Soc. 77: 168-241.

Carriker, M.R., 1996, The shell and ligament, in: Biology, Culture, and Management of the Eastern Oyster (A. Eble and V Kennedy, eds.), Maryland Office of Sea Grant, College Park, MD.

Carriker, M. R., 2001, Embryogenesis and organogenesis of veligers and early Juveniles, In: Biology of the hard clam (J. N. Kraeuter and M. Castagna, eds.), pp. 103-112, Amsterdam.

Carriker, M.R., and Palmer, R.E., 1979, A new mineralized layer in the hinge of the oyster, *Science* 206: 691-693.

Carriker, M.R., Palmer, R.E., Sick, L.V., Johnson, C.C., 1980, Interaction of mineral elements in seawater and shell of oysters (Crassostrea virginica (Gmelin)) cultured in controlled and natural systems, *J. Exp. Mar. Biol. Ecol.* 46: 279-296.

Carriker, M.R., Swam, C.P., Ewart, J., Counts, C.I. III, 1996, Ontogenetic Trends of elements (Na-Sr) in prismatic shell of living Crassostrea virginica (Gmelin) grown in three ecologically dissimilar habitats for 28 weeks: a proton probe study, *J. Ecol. Mar. Biol. Ecol* 201: 87-135

Carrol, M.L., Johnson, B. J., Hankes, G.A., McMahan, K.W., Voronkov, A., Ambrose, W.G. Jr., Densienko, S.G., 2009, Bivalves as indicators of environmental variation and potential anthropogenic impacts in the Southern Barrents Sea, *Marine Pollution Bulletin* 59: 193-206

Carter, J. G., 1980, Environmental and biological controls of bivalve shell menerology and microstructure, in: Skeletal Growth of Aquatic Organisms, Vol. I, Topics in geobiology (D. C. Rhoades and R. A. Lutz, eds.), pp. 203-254, Plenum Press, New York.

Carter, J.G. (ed), 1990, Skeletal biomineralization: patterns, processes and evolutionary trends, Vol I, Van Nostrand Reinhold, New York.

Castagna, M., Kraeuter, J.N., 1977, *Mercenaria* culture using stone aggregate for predator protection, *Proc. Natl. Shellfish Ass.* 67: 1–6

Cathey, A. M., Miller, N. R., Kimmel D. G., 2012, Microchemistry of juvenile Mercenaria mercenaria shell: implications for modeling larval dispersal, *Mar. Ecol. Prog. Ser.* 465:155-168.

Chateigner, D., Hedegaard, C., Wenk, H. R., 2000, Mollusc shell microstructures and crystallographic textures, *Journal of Structural Geology* 22(11): 1723-1735.

Chauvaud, L., Thouzeau, G., Paulet, Y.-M., 1998, Effects of environmental factors on the daily growth rate of *Pecten maximus* juveniles in the Bay of Brest (France), *J. Exp. Mar. Biol. Ecol.* 227: 83–111.

Chauvaud, L., Thouzeau, G., Paulet, Y.M., 1998, Effects of environmental factors on the daily growth rate of clam *Corbicula fluminea* (Mollusca: Bivalvia), *Limnol. Oceanog*. 35: 756-762.

Chauvaud, L., Lorrain, A., Dunbar, R.B., Paulet, Y.-M., Thouzeau, G., Jean, F., Guarini, J.-M., Mucciarone, D., 2005, Shell of the great scallop *Pecten maximus* as a high-frequency archive of paleoenvironmental changes, *Geochem Geophys Geosyst* 6: Q08001.

Chave, K.E., 1954, Aspects of the biochemistry of magnesium. 1. Calcareous marine organisms, *J. Geol.* 62: 266-283.

Clarke, F. W., and Wheeler, W. C., 1922, The inorganic constituents of marine invertebrates, *U.S. Geol. Surv. Prof. Pap.* 124:1-62.

Clark, G.R. II, 1974, Calcification on an unstable substrate: marginal growth in the mollusk *Pecten diegensis*, *Science* 183:968–970.

Clark, G.R. II, 1975, Periodic growth and biological rhythms in experimentally grown bivalves, in: Growth rhythms and the history of the earth's rotation (G.D. Rosenberg and S.K.Runcorn, eds), p.p. 103–117, Wiley, London.

Clarke, J. H., Clarke, A. N., Wilson, D. J., Freauf, J. J., 1976, Lead levels in freshwater mollusk shells, *J. Environ. Sci. Health Part A* All:65-78.

Coffey, M., Dehairs, F., Collette, O., Luther, G., Church, T., Jickells, T., 1997, The behaviour of dissolved barium in estuaries, *Estuarine Coastal and Shelf Science* 45: 113-121.

Comfort, A., 1951, The pigmentation of molluscan shells, *Biol. Rev.* 26:285-301.

Cook, P. J., 1977, Loss of boron from shells during weathering and possible implications determination of salinity, *Nature* (London) 268:426-427.

Cravo, A., Bebianno, M.J., Foster, P., 2004, Partitioning of trace metals between soft tissues and shells of Patella aspera, *Environment International* 30: 87-98.

Crawford, D. W., Lipsen, M. S., Purdie, D. A., Lohan, M. C., Statham, P. J., Whitney, F. A., Putland, J. N., Johnson, W. K., Sutherland, N., Peterson, T. D., Harrison, P. J., Wong C. S., 2003, Influence of zinc and iron enrichments on phytoplankton growth in the northeastern subarctic Pacific, *Limnol. Oceanogr.* 48: 1583–1600.

Crenshaw, M. A., 1972, The inorganic composition of molluscan extrapallial fluid, *Biol. Bull.* 143: 506–512.

Crenshaw, M. A., 1972, The soluble matrix from Mercenaria mercenaria shell, *Biomineralization* 6: 6-11.

Crenshaw, M. A., 1980, Mechanisms of shell formation and dissolution, in: Skeletal Growth of Aquatic Organisms, Vol. I, Topics in geobiology, (D. C. Rhoades and R. A. Lutz, eds.), pp. 115-132. Plenum Press, New York.

Crenshaw, M. A., and Neff, J.M., 1969, Decalcification at the mantle-shell interface in moluscs, *Am. Zool.* 9:881-885.

Crenshaw, M. A., and Ristedt, H., 1976, The histochemical localization of reactive groups in the septal nacre from Nautilus pompilius L., in: The Mechanisms of Mineralization in the Invertebrates and Plants (N. Watebe and K.M. Wilbur, eds.), pp. 335-367, University of South Carolina Press, Colombia.

Crisp, E. L., 1975, The skeletal trace element chemistry of freshwater bivalves, Ph.D. dissertation, 187pp., University of Indiana, Bloomington.

Crisp, E. L., 1976, Skeletal trace element chemistry of freshwater bivalves, *Fed. Soc. Am. Proc.* 8:15-16.

Cullen, J. T., Sherrell, R. M., 2005, Effects of dissolved carbon dioxide, zinc, and manganese on the cadmium to phosphorus ratio in natural phytoplankton assemblages, *Limnol. Oceanogr.* 50: 1193–1204.

Davis, R.L., and Marshall, N., 1961, The feeding of the bay scallop, Aquipecten irradians, *Proc. Nat. Shellfish Assoc.* 52: 25-29.

Davies, T. T., and Sayre, J. G., 1970, Th effect of environmental stress on pelecypod shell ultrastructure, *Geol. Soc. Am. Abstr., Southeastern Section* 204-205.

DiBacco, C., and Levin, L. A., 2000, Development and application of elemental fingerprinting to track the dispersal of marine invertebrate larvae, *Limnol. and Oceanogr.* 45: 871-880.

Dodd, J. R., 1964, Environmentally controlled variation in the shell structure of a pelecypod species, *J. Paleontol.* 38: 1065-1071.

Dodd, J.R., 1965, Environmental control of strontium and magnesium in Mytilus, *Geochim. Cosmochim. Acta* 29:385-398.

Dodd, J. R., 1967, Magnesium and strontium in calcareous skeletons: A review, *Journal of Paleontology* 41:1313-1329.

Dodd, J. R., and Crisp, E. L., 1982, Non-linear variation with salinity of Sr/Ca and Mg/Ca ratios in water and aragonitic bivalve shells and implications for palaeosalinity studies, *Palaeogeogr. Palaeoclim. Palaeoecol.* 38: 45–56.

Dorval, E., Jones, Hannigan, C. M. R., Montfrans, J., 2007, Relating otolith chemistry to surface water chemistry in a coastal plain estuary, Canadian *Journal of Fisheries Aquatic Sciences* 64:411-424.

D'Souza, F. M., and Kelly, G. J., 2000, Effects of a diet of a nitrogen-limited alga Tetraselmis suecica on growth, survival and biochemical composition of tiger prawn Penaeus semisulcatus larvae, *Aquaculture 181*(3): 311-329.

Dunca, E., Mutvei, H., 2001, Comparison of microgrowth pattern in *Margaritifera margaritifera* shells from south and north Sweden., *Am. Malacol. Bull.* 16: 239–250.

Dunca, E., Schöne, B.R., Mutvei, H., 2005, Freshwater bivalves tell of past climates: but how clearly do shells from polluted rivers speak, *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 228: 43–57.

Enright, C. T., Newkirk, G. F., Craigie, J. S., Castell, J. D., 1986, Evaluation of phytoplankton as diets for juvenile Ostrea edulis L, *J. exp. mar. Biol. Ecol.* 96: 1-13.

Enright, C. T., Newkirk, G. F., Craigie, J. S., Castell, J. D., 1986, Growth of juvenile Ostrea edulis L. fed Chaetoceros gracilis of varied chemical composition. *J. exp. mar. Biol. Ecol.* 96: 15-26.

Epifanio, C.E., 1979, Growth in bivalve molluscs: Nutritional effects of two or more species of algae in diets fed to the American oyster Crassostrea virginica (Gmelin) and the hard clam Mercenaria mercenaria (L.), *Aquaculture* 18: 1–12.

Eversole, A.G., 2001, Reproduction in Mercenaria mercenaria, in: Biology of the hard clam (J. N. Kraeuter and M. Castagna, eds.), pp. 221-256, Amsterdam.

Falini, G., Albeck, S., Weiner, S., Addadi, L., 1996, Control of aragonite or calcite polymorphism by mollusc shell macromolecules, Science 271: 67–69.

Fang, L. –S., and Shen, P., 1984, Foreign elements in a clam shell: a clue to the history of marine pollution events, Mar. Ecol. Prog. Ser. 18: 187-189

Fisher, N.S., Guillard, R.R.L., Bankston, D.C., 1991, The accumulation of barium by marine phytoplankton grown in culture, *Journal of Marine Research* 49: 339-354. Ferrel, R. E., Carville, T. E., Martinez, J. D., 1973, Trace metals in oyster shells, Environ. Lett. 4:311-316.

Foster, L. C., Finch, A. A., Allison, N., Andersson, C., Clarke, L. J., 2008, Mg in aragonitic bivalve shells: Seasonal variations and mode of incorporation in Arctica islandica, *Chemical Geology* 254:113-119.

Foster, P., and Cravo, A., 2003, Minor elements and Trace metals in the shell of marine gastropods from a shore in tropical east Africa, *Water Air and Soil Pollution* 145(1-4): 53-64.

Fox, D. L., 1966, The pigmentation in molluscs, in: Physiology of Mollusca, Vol. 2 (K. M. Wilbur and C. M. Yonge, eds.), pp. 249-274, Academic Press, New York.

Freitas, P.S., Clarke, L.J., Kennedy, H., Richardson, C.A., Abrantes, F., 2006, Environmental and biological controls on elemental (Mg/Ca, Sr/Ca, Mn/Ca) ratios in shells of the King scallop Pectin maximus, *Geochim Cosmochim Acta* 70: 5119-5133.

Fritz, L.W., and Haven, D.S., 1983, Hard clam, *Mercenaria mercenaria*, shell growth patterns in the Chesapeake Bay, *Fish Bull*. 81:697–708.

Fritz, L.W., Ragone, L.M., Lutz, R.A., 1990, Biomineralization of barite in the shell of the freshwater Asiatic clam Corbicula fluminea (Mollusca: Bivalvia), *Limnol. Oceanogr.* 35:756-762.

Fuge, R., Palmer, T. J., Pearce, N. J. G., Perkins, W. T., 1993, Minor and trace element chemistry of modern shells: A laser ablation inductively coupled plasma mass spectrometry study, *Appl. Geochem. Suppl.* 2: 111–116.

Furst, M., Lowenstam, H. A., Burnett, D. S., 1976, Radiographic study of the distribution of boron in recent mollusc shells, *Geochim. Cosmochim. Acta* 40:1381-1386.

Furuhashi, T., Schwarzinger, C., Miksik, I., Smrz, M., Beran, A., 2009, Molluscan shell evolution with review of shell calcification hypothesis, *Comprative Biochemistry and Phsiology*, Part B 154:351-371.

Ganeshram, R.S., Francois, R., Commeau, J.,Brown-Leger, S.L., 2003, An experimental investigation of barite formation in seawater, *Geochimica Cosmochimica Acta* 70: 4617-4634.

Ghiselin, M.T., Degens, E.T., Spencer, D.W., Parker, R.H., 1967, A phylogenetic survey of molluscan shell matrix proteins, *Breviora* 262: 1–35.

Gilliken, D.P., 2005, Geochemistry of marine Bivalve shells: the potential for paleoenvironmental reconstruction, Ph.D. dissertation, University of Brussels.

Gillikin, D.P., Dehairs, F., Baeyens, W., Navez, J., Lorrain, A., André, L., 2005, Interand intra-annual variations of Pb/Ca ratios in clam shells (*Mercenaria mercenaria*): a record of anthropogenic lead pollution, *Mar. Pollut. Bull.* 50:1530–1540.

Gillikin, D.P., Lorrain, A., Navez, J., Taylor, J.W., André, L., Keppens, E., Baeyens, W., Dehairs, F., 2005, Strong biological controls on Sr/Ca ratios in aragonitic marine bivalve shells, *Geochem. Geophys. Geosyst.* 6.

Gillikin, D. P., F. Dehairs, A. Lorrain & D. Steenmans, Baeyens, W., Andre, L., 2006, Barium uptake into the shells of the common mussel Mytilus edulis and the potential for estuarine paleo-chemistry reconstruction, *Geochim. Cosmochim. Acta* 70: 395-407.

Gillikin, D. P., Lorrain, A., Paulet, Y. M., Andre, L., Dehairs, F., 2008, Synchronous barium peaks in high resolution profiles of calcite and aragonite marine bivalve shells, *Geo. Marine* Letters 28: 351-358.

Gomez-Alday, J. J., and Elorza, J., 2001, Diagenesis, regular growth and records of seasonality in inoceramid bivalve shells from mid-Masstrichian hemipelagic beds of the Bay of Biscay, Netherlands, *Journal of Geosciences* 82(3): 289-301.

Gotliv, B.-A., Kessler, N., Sumerel, J. L., Morse, D. E., Tuross, N., Addadi, L., Weiner, S., 2005, Asprich: a novel aspartic acid-rich protein family from the prismatic shell matrix of the bivalve Atrina rigida, *Chembiochem* 6: 304–314.

Gresfrude, E. S., and Strand, O., 2006, Comparison of shell strength in wild and cultured scallops (Pecten maximus), *Aquaculture* 251:306-313.

Gresfrud, E.S., Dauphin, Y., Cuif, J.P., Denis, A., Strand, O., 2008, Modifications in microstructure of cultured and wild scallop shells (Pectin maximus), *Journal of Shellfish Research* 27(4).

Guillard, R.R.L., 1983, Culture of phytoplankton for feeding marine invertebrates, in: Culture of Marine Invertebrates, Selected Readings (C.J. Bert, Jr. and H. Ross), pp. 108-132, Stroudsburg, PA.

Hallam, A., and Price, N. B., 1968, Environmental and biochemical control of strontium in shells of Cardium edule, *Geochim. Cosmochim. Acta* 32:319-328.

Hamer, P.A., and Jenkins, G.P., 2007, Comparison of spatial variation in otolith chemistry of two fish species and relationships with water chemistry and otolith growth, *J. Fish Biol.* 71: 1035–1055.

He, G., and Mai, K., 2001, Ontogenetic trends of mineralogy and elements in the shell of abalone, Halliotis discus Hannai Ino, *Journal of Shellfish Research* 20(2): 685-687.

He´douin, L., 2006, Caracte´risation d'espe`ces bioindicatrices pour la surveillance des activite´s minie`res et la gestion de l'environnement en milieu re´- cifal et lagonaire: application au lagon de la Nouvelle-Cale´donie. Ph.D. thesis, La Rochelle University, France.

He´douin, L., Pringault, O., Metian, M., Bustamante, P., Warnau, M., 2007, Nickel bioaccumulation in bivalves from the New Caledonia lagoon: seawater and food exposure, *Chemosphere* 66: 1449-1457.

Hemming, N.G., and Hanson, G.N., 1992, Boron isotopic composition and concentration in modern marine carbonates, *Geochimica et Cosmochimica Acta* 56: 537-543.

Hintz, C.J., Shaw, T.J., Chandler, G.T., Bernhard, J.M., McCorkle, D.C., blanks, J.K., 2006, Trace/minor element:calcium ratios in cultured benthic foraminifera. Part 1: interspecies and inter-individual variability, *Geochimica Cosmochimica Acta*. 70(8): 1952-1963.

Ho, T. -Y., Quigg, A., Finkel, Z. V., Milligan, A. J., Wyman, K., Falkowski, P. G., Morel, F. M. M., 2003, The elemental composition of some marine phytoplankton, *J. Phycol.* 39: 1145–1159.

Holdaway, H. K., and Clayton, C. J., 1982, Preservation of shell microstructure in silicified brachiopods from the Upper Cretaceous Wilmington Sands of Devon, *Geological Magazine*, 119: 371-382.

Hudson, R. J. M., 1998, Which aqueous species control the rates of trace metal uptake by aquatic biota? Observations and predictions of non-equilibrium effects, *Sci. Total Environ*. 219: 95–115.

Hutchins, D. A., Witter, A. E., Butler, A., Luther, G. W., 1999, Competition among marine phytoplankton for different chelated iron speciation, *Nature* 400: 858–861.

Incze, L.S., Lutz, R.A., Watling, L., 1980, Relationships between effects of environmental temperature and seston on growth and mortality of *Mytilus edulis* in a temperate northern estuary, *Mar. Biol.* 57:147–156.

Jacob, D.E., Soldati, A., Wirth, R., Huth, J., Wehrmeister, U., Hofmeister, W., 2008, Nanostructure, composition and mechanisms of bivalve shell growth, *Geochimica et cosmochimica Acta* 72: 5401-5415.

Jennings, J. R., Rainbow, P. S., Scott, A. G., 1979, Studies on the uptake of cadmium by the crab Carcinus maenas in the laboratory. II. Preliminary investigation of cadmium-binding proteins, *Mar. Biol.* 50: 141–149.

Jodrey, L. H., and Wilbur, K. M., 1955, Studies on shell Formation. IV. The respiratory metabolism of the oyster mantle, *Biol. Bull.* 108:346-358.

Johnstone, M. O., 2008, Characterization of soluble matrix from molluscan shell with an emphasis on two major phosphoproteins from the Eastern Oyster, Crassostrea virginica, Ph.D. dissertation, Clemson University.

Johnstone, M. B., Ellis, S., Mount, A. S., 2008, Visualization of shell matrix proteins in hemocytes and tissues of the Eastern Oyster, Crassostrea virginica, *Journal of Experimental Zoology (Mol Dev Evol*), 310B:227-239.

Jurkiewicz-Karnkowska, E., 2002, Differentiation of phosphorous concentration in selected mollusk pecies from Zegrynski Resevoir (central Poland): implications for P accumulation in mollusk communitis, *Polish journal of Environmental Studies* 11(4): 355-359.

Katz M. E., Fennel K., Falkowski P. G., 2007, Geochemical and biological consequences of phytoplankton evolution, in: Evolution of Primary Production in the Sea, (P. G. Falkowski and A. H.Knoll, eds.), pp 405–430, Amsterdam: Elsevier.

Kennedy, W. J., Taylor, J. D., Hall, A., 1969, Environmental and Biological Controls on Bivalve Shell Mineralogy, *Biological Reviews* 44(4): 499-530.

Kennish, M.J., Olsson, R.K., 1975, Effects of thermal discharges on the microstructural growth of *Mercenaria mercenaria*, *Environ Geol* 1:41–64.

Klein, R.T., Lohmann, K.C., Thayer, C.W., 1996, Bivalve skeletons record seasurface temperature and delta O-18 via Mg/Ca and O-18/O-16 ratios, *Geology* 24: 415-418.

Klein, R.T., Lohmann, K.C., Thayer, C.W., 1996, Sr/Ca and 13C/12C ratios in skeletal calcite of Mytilus trossulus: covariation with metabolic rate, salinity, and carbon isotopic composition of seawater, *Geochimica Cosmochimica Acta* 60: 4207-4221.

Kobayashi, I., 1975, Preliminary study on the distribution of some elements in the shell of some bivalvian molluscs by the electron microprobe analysis, *Sci. Rep. Niigata Univ. Ser. E.* 3:41-50.

Kobayashi, I., and Samata, T., 2006, Bivalve shell structure and organic matrix, *Materials Science and Engineering*: C 26(4): 692-698.

Krause-Nehring, J., Klugel, A., Nehrke, G., Brellochs, B., and Brey, T., 2011, Impact of sample pretreatment on the measured element concentrations in the bivalve Arctica islandica, *Geochemistry Geophysics and Geosystems* 12: Q07015.

Lazareth, C.E., Vander Putten, E., Andre, L., Dehairs, F., 2003, High- resolution trace element profiles in shells of the mangrove bivalve Isognomon ephippium: a record of environmental spatio-temporal variation, *Estuarine coastal and shelf science* 57: 1103-1114.

Lazareth, C. E., Guzman, N., Poitrasson, F., Candaudap, L., Ortlieb, L., 2007, Nyctemeral variations of magnesium intake in the calcitic layer of a Chilean mollusk shell (Concholepas concholepas, Gastropoda), *Geochim. Cosmochim. Acta* 71: 5369–5383.

Lee, J. G., and Morel, F.M.M. 1995. Replacement of zinc by cadmium in marine phytoplankton, *Mar. Ecol. Prog. Ser.* 127:305–309.

Lee, J. G., Roberts, S. B., Morel, F. M. M., 1995, Cadmium: a nutrient for the marine diatom Thalassiosira weissflogii, *Limnol. Oceanogr.* 40:1056–63.

Leng, M. and Pearce, N., 1999, Seasonal variation of trace element and isotopic composition in the shell of a coastal mollusk, Mactra isabelleana, *J. Shellfish Res.* 18: 569–574.

Levi-Kalisman, Y., Falini, G., Addadi, L., Weiner, S., 2001, Structure of the nacreous organic matrix of a bivalve mollusk shell examined in the hydrated state using cryo-TEM, *J. Struct. Biol.* 135: 8–17.

Lingard S. M., Evans R. D., Bourgoin B. P., 1992, Method for the Estimation of Organic-Bound and Crystal-Bound Metal Concentrations in Bivalve Shells, *Bull. Environ. Contam. Toxicol.* 48: 179–184.

Loosanoff, V.L., and Davis, H.C., 1963, Rearing of bivalve molluscs, *Adv. Mar. Biol.* 1:1–136.

Lorens, R. B., 1981, Sr, Cd, Mn and Co distribution coefficients in calcite as a function of calcite precipitation rate, *Geochim. Cosmochim. Acta* 45: 553–561.

Lorens, R. B, and Bender M. L., 1980. The impact of solution chemistry on Mytilus edulis calcite and aragonite, *Geochim. Cosmochim. Acta*. 44:1265-1278.

Lorrain, A., Gillikin, D.P., Paulet, Y.-M., Chauvaud, L., Le Mercier, A., Navez, J., André, L., 2005, Strong kinetic effects on Sr/Ca ratios in the calcitic bivalve *Pecten maximus*, *Geology* 33:965–968.

Lowenstam, H.A., and Weiner, S., 1989, On Biomineralization. Oxford University Press, pp. 324.

Lu, Y., and Blake, N. J., 1997, The culture of the southern bay scallop in Tampa Bay, an urban Florida estuary, *Aquaculture International* 5: 439–450.

Lutz, R. A., and Rhoades, D. C, 1977, Anaerobiosis and a theory of growth line formation, *Science* 198:1222-1227.

Lutz, R. A., and Rhoades, D. C, 1980, Growth Patterns within the Molluscan shell, in: Skeletal Growth of Aquatic Organisms, Vol. I, Topics in geobiology, (D. C. Rhoades and R. A. Lutz, eds.), pp. 203-254. Plenum Press, New York.

Mann, S., Webb, J., and Williams, R. J. P., 1989, Biomineralization: Chemical and Biochemical Perspectives, VCH, Weinheim.

Markich, S.J., Jeffree, R.A., Burke, P.T., 2002, Freshwater bivalve shells as archival indicators of metal pollution from a copper-uranium mine in tropical northern Australia, *Environ. Sci. Technol.* 36:821–832.

Martin, J. H., and Knauer, G. A., 1973, The elemental composition of plankton, *Geochim. Cosmochim. Acta* 37:1639–53.

Martin, G. B., and Thorrold, S. R., 2005, Temperature and salinity effects on magnesium, manganese, and barium incorporation in otoliths of larval and early juvenile spot Leiostomus xanthurus, *Mar. Ecol. Prog. Ser.* 293:223-232.

Martincié, D., Nürnberg, H.W., Stoeppler, M., Branica, M., 1984, Bioaccumulation of heavy metals by bivalves from Lim Fjord (North Adriatic Sea), *J. Marine Biology* 81(2): 177-188.

Martínez-Fernández, E., Acosta-Salmón, H., Southgate, P. C., 2006, The nutritional value of seven species of tropical microalgae for black-lip pearl oyster Pinctada margaritifera larvae, *Aquaculture 257*(1): 491-503.

Meyers, J. M., Johnstone, M. B., Mount, A. S., Silverman, H., Wheeler, A. P., 2007, TEM immunochemistry of a 48kDa MW organic matrix phosphoprotein produced in the mantle epithelial cells of the Eastern oyster (Crassostrea virginica), *Tissue and Cell* 39:247-256.

Milke, L.M., Bricelj, V.M., Parrish, C.P., 2006, Comparison of early life history stages of the bay scallop Argopecten irradians: effects of microalgae diets on growth and biochemical composition, *Aquaculture* 254(1-4): 526-533.

Milliman, J. D., 1974, Recent sedimentary carbonates, in: Marine carbonates, Part 1, 375 pp., Berlin, New York: Springer Verlag.

Milner, H. W., 1953, The chemical composition of algae, *Algal culture from laboratory to pilot plant. Publ. Cameg. Instn 600*: 285-302.

Mount, A.S., 1999, Nucleation of calcite in the Eastern oyster, Crassostrea virginica: biochemical and functional studies of the organic matrix from foliated shell, Ph.D. Dissertation, Clemson University.

Mount, A.S., Wheeler, A.P., Paradkar, R.P., Snider, D., 2004, Hemocyte mediated shell mineralization in the Eastern oyster, *Science* 304 (5668): 297–300.

Nakahara, H., and Bevelander, G., 1970, An electron microscope study of the muscle attachment in the mollusc Pinctada radiate, *Texas Rep. Biol. Med.* 28:279-286.

Neff, J. M., 1972, Ultrastructure of the outer epithelium of the mantle in the clam, Mercenaria mercenaria, in relation to calcification of the shell, *Tissue Cell* 4:591-600.

Nudelman, F., Chen, H.H., Goldberg, H.A., Weiner, S., Lia Addadi, L., 2007, Lessons from biomineralization: comparing the growth strategies of mollusc shell prismatic and nacreous layers in Atrina rigida, *Faraday Discuss*. 136: 9–25.

Nudelman, F., Gotliv, B.A., Addadi, L., Weiner, S., 2006, Mollusk shell formation: mapping the distribution of organic matrix components underlying a single aragonitic tablet in nacre, *J. Struct. Biol.* 153: 176–187.

Palmer, R.E., and Carriker, M.R., 1979, Effects of cultural conditions on morphology of the shell of the oyster Crussostrea virginica, *Proc. Natl. Shellfish. Assoc.* 69:57-72.

Panella, G. and Mac Clintock, C., 1968, Biological and environmental rhythms reflected in molluscan shell growth, *J. Paleontol.* 42: 64-80.

Peers G., Quesnel S. A., Price N. M., 2005, Copper requirements for iron acquisition and growth of coastal and oceanic diatoms, *Limnol. Oceanogr*. 50: 1149–1158.

Peirson, W.M., 1983, Utilization of eight algal species by the bay scallop Argopecten irradians concentricus (Say), *J. Exp. Biol. Ecol.* 68: 1-11.

Peterson, C. H. & S. R. Fegley. 1986. Seasonal allocation of resources to growth of shell, soma, and gonads in Mercenaria mercenaria. *Biol. Bull*.171:597-610.

Peterson, R.P., 1998, A method for trace element determination of marine periphyton communities on discs of float glass (without sample preparation) using total-reflection X-ray fluorescence spectrometry, *Spectrochimica Acta Part B: Atomic Spectroscopy* 53(1): 101-115.

Phillips, D.J.H., 1977, The use of biological indicator organisms to monitor trace metal pollution in marine and estuarine environments: a review, *Environ. Pollut.* 13: 281–317.

Phillips, D.J.H., 1990. Use of macroalgae and invertebrates as monitors of metal levels in estuaries and coastal waters, in: Heavy Metals in the Marine Environment (F.W. Fumess and P.S. Rainbow, eds.), pp. 8 I-99, CRC Press, Boca Raton, FL,.

Pitts, L.C.,and Wallace, G.T., 1994, Lead deposition in the shell of the bivalve, Mya arenaria: an indicator of dissolved lead in seawater, *Estuar. Coast. Shelf Sci.* 39: 93–104.

Poigner, H., Monien, D., Monien, P., Kriews, M., Brumsack, H., J., Wilhelms-Dick, D., Abele, D., 2012, Element ratios between digestive gland and gill tissues of the Antarctic bivalve Laternula elliptica as a proxy for element uptake from different environmental sources, *Geophysical Research Abstracts* 14: EGU2012-6576.

Price, N. M., and Morel, F. M. M., 1990, Cadmium and cobalt substitution for zinc in a marine diatom, *Nature* 344: 658–660.

Puente, X., Villares, R., Carral, E., Carballeira , A., 1996, Nacreous shell of Mytilus galloprovincialis as a biomonitor of heavy metal pollution in Galiza (NW Spain), *Sci. Total Environ.* 183: 205–211.

Quigg, A., Finkel, Z. V., Irwin, A. J., Rosenthal, Y., Ho, T.-Y., Reinfelder, J. R., Schofield, O., Morel, F. M. M., Falkowski, P. G., 2003, The evolutionary inheritance of elemental stoichiometry in marine phytoplankton, *Nature* 425:291–294.

Rainbow, P.S., 1993, The significance of trace metal concentrations in marine invertebrates, in: Ecotoxicology of metals in invertebrates (Dallinger R, Rainbow PS, eds), Lewis Publishers, Boca Raton, FL, pp 4–23

Redfield, A. C., 1934, On the proportions of organic derivatives in sea water and their relation to the composition of plankton, in: James Johnstone Memorial Volume (R. J. Daniel, ed.), pp. 176–92, Liverpool University Press.

Redfield, A. C., 1958, The biological control of the chemical factors in the environment, *Am. Sci.* 46:205–21.

Redfield, A. C., Ketchum B. H., Richards F. A., 1963, The influence of organisms on the composition of sea-water, in: The Sea (M. N Hill., ed.) pp. 26–77, New York: Interscience Publication.

Reinfelder, J. R., and Fisher, N. S., 1991, The assimilation of elements ingested by marine copepods, *Science* 251: 794-796.

Reinfelder, J. R., and Fisher, N. S., 1994, The Assimilation of Elements Ingested by Marine Planktonic Bivalve Larvae, *Limnol. Oceanog.* 39(1): 12-20.

Reinfelder, J.R., Fisher, N.S., Luoma, S.N., Nichols, J.W., Wang, W.-X., 1998, Trace element trophic transfer in aquatic organisms: A critique of the kinetic model approach, *Science of The Total Environment* 219(2–3): 117-135.

Reitan, K. I., Rainuzzo, J. R., Øie, G., Olsen, Y., 1997, A review of the nutritional effects of algae in marine fish larvae, *Aquaculture* 155(1–4): 207-221.

Reitan, K. I., Rainuzzo, J. R., Olsen, Y., 2004, Effect of nutrient limitation on fatty acid and lipid content of marine microalgae, *Journal of Phycology 30*(6): 972-979.

Renaud, S. M., Zhou, H. C., Parry, D. L., Thinh, L. V., Woo, K. C., 1995, Effect of temperature on the growth, total lipid content and fatty acid composition of recently isolated tropical microalgae Isochrysis sp., Nitzschia closterium, Nitzschia paleacea, and commercial species Isochrysis sp.(clone T. ISO), *Journal of Applied phycology 7*(6): 595-602.

Renaud, S. M., Thinh, L. V., Lambrinidis, G., and Parry, D. L., 2002, Effect of temperature on growth, chemical composition and fatty acid composition of tropical Australian microalgae grown in batch cultures, *Aquaculture 211*(1): 195-214.

Rhoads, D.C., and Lutz, R.A., 1980, Skeletal records of environmental change, in: Skeletal Growth of Aquatic Organisms (D.C. Rhoads and R. A. Lutz, eds.), pp. 1-19, Plenum Press, New York.

Ridgway, I.D., Richardson, C.A., Enos, E., Ungvari, Z., Austad, S.N., Philipp, E.E.R., Csiszar, A., 2011, New species longevity record for the northern quahog (=hard clam), Mercenaria mercenaria, *Journal of Shellfish Research* 30(1): 35-38.

Riley, J. P., Roth, I., 1971, The Distribution of Trace Elements in Some Species of Phytoplankton Grown in Culture, *Journal of the Marine Biological Association of the united Kingdom* 51(01): 63-72.

Risk, M., Burchell, M., de Roo, K., Nairn, R., Tubrett, M., Forsterra, G., 2010, Trace elements in bivalve shells from the Rio Cruces, Chile, *Aquat. Biol.* 10:85-97.

Roditi, H.A., and Fisher, N.S., 1999, Rate and Routes of trace element uptake in zebra mussels, *Limnol. Oceanograph*. 44(7): 1730-1749.

Rollion-Bard, C., Chaussidon, M., France-Lanord, C., 2003, pH control on oxygen isotopic composition of symbiotic corals, *Earth and Planetary Science Letters* 215: 275-288.

Romberger, H. P., and Epifanio, C. E., 1981, Comparative effects of diets consisting of one or two algal species upon assimilation efficiencies and growth of juvenile oysters, Crassostrea virginica (Gmelin), *Aquaculture* 25: 77-87.

Rosenberg, G. D., 1980, An ontogenetic approach to the environmental significance of bivalve shell chemistry, in: Skeletal Growth of Aquatic Organisms, Vol. I, Topics in geobiology (D. C. Rhoads and R. A. Lutz, eds.), pp. 133-168, Plenum, New York.

Rosenberg, G.D., 1990, The "vital effect" on skeletal trace element content as exemplified by magnesium, in: Skeletal Biomineralization: Patterns, Processes and Evolutionary Trends, Vol. 1 (J.G. Carter and Van Nostrand Reinhold, eds.), pp. 567-577, New York, NY.

Rosenberg, G. D., and Hughes, W. W., 1991, A metabolic model for the determination of shell composition in the bivalve mollusc, Mytilus edulis, *Lethaia* 24: 83–96.

Rucker, J.B., and Valentine, J. W., 1961, Salinity response of trace element concentration in Crassostrea virginica, *Nature (London)* 190:1099-1100.

Sánchez, S., Martínez, M., & Espinola, F. (2000). Biomass production and biochemical variability of the marine microalga< i> Isochrysis galbana</i> in relation to culture medium. *Biochemical engineering journal*, *6*(1), 13-18.

Schöne, B. R. 2008. The curse of physiology-challenges and opportunities in the interpretation of geochemical data from mollusk shells. *Geo. Mar. Lett.* 28: 269-285

Schone, B. R., Zhang, Z., Jacob, D., Gillikin, D. P., Tutken, T., Garbe-Schonberg, D., McConnaughey, T., Soldati, A., 2010, Effect of organic matrices on the determination of he trace element chemistry (Mg, Sr, Mg/Ca, Sr/Ca) of aragonite bivalve shells (Artica islandica) – Comparison of ICP-OES and LA-ICP-MS data, *Geochemical Journal* 44:23-37.

Schopf, T.J.M., 1980, Paleoceanography, 341p, Harvard University Press, Cambridge.

Segar, D. A., Collins, J. D., and Riley, J. P., 1971, The distribution of the major and some minor elements in marine animals. Part II: Moluscs, *J. Mar. Biol. Assoc. U.K.* 51:131-136.

Shumway, S. E., 1977, Effect of salinity fluctuation on osmoticpressure and Na+, Ca 2+ and Mg 2+ ion concentrations in hemolymph of bivalve mollusks, *Mar. Biol.* 41: 153–177.

Shumway, S. E., Selvin, R., Schick, D. F., 1987, Food resources related to habitat in the scallop Placopecten magellanicus (Gmelin, 1791): a qualitative study, *J. Shellfish Res*, 6(2): 89-95.

Simkiss, K., and Wilbur, K. M., 1989, Biomineralization, in: Cell Biology and Mineral Deposition, 337 pp, Academic Press, San Diego.

Simpson, R. D., 1979, Uptake and loss of zinc and lead by mussels (Mytilus edulis) and relationships with body weight and reproductive cycle, *Mar. Pollut. Bull.*10: 74–78.

Spencer, C. P., 1957, Utilization of trace elements by marine unicellular algae, *Journal of General Microbiology 16*(1): 282-285.

Stanley, S.M., 1970, Relation of shell form to life habitats in the Bivalvia, *Geol. Soc. Am. Mem.* 125:1-296.

Stanton, R. J., and Dodd, J. T., 1970, Paleoecologic techniques – comparison of faunal and geochemical analyses of Pliocene paleoenvironments, Kettlemen Hills, California, *J. Paleontal.* 44:1092-1121.

Stecher, H.A. III, Krantz, D.E., Lord, C.J. III, Luther, G.W. III, Bock, K.W., 1996, Profiles of strontium and barium in *Mercenaria mercenaria* and *Spisula solidissima* shells, *Geochim Cosmochim Acta* 60:3445–3456.

Stecher, H.A., and Kogut, M.B., 1999, Rapid barium removal in the Delaware estuary, *Geochimica Cosmochimica Acta* 63: 1003-1012.

Strasser, C.A., Mullineaux, L.S., Walther, B.D., 2008, Growth rate and age effects on Mya arenia shell chemistry: implications for biogeochemical studies, *J. Exp. Mar. Biol. Ecol.* 355: 153-163.

Strasser, C.A., Mullineaux L.S., Thorrold S.R., 2008, Temperature and salinity effects on elemental uptake in the shells of larval and juvenile softshell clams Mya arenaria, *Mar. Ecol. Prog. Ser.* 370: 155-169.

Sudo, S., Fujikawa, T., Nagakura, T., Ohkubo, T., Sakaguchi, K., Tanaka, M., Nakashima, K., Takahashi, T., 1997, Structure of mollusc shell framework proteins, *Nature* 387:563–564.

Sunda, W. G., 1988/1989, Trace metal interactions with marine phytoplankton, *Biol. Oceanogr*. 6: 411–442.

Sunda, W. G., 2012, Feedback interactions between trace metal nutrients and phytoplankton in the ocean, *Frontiers in Microbiology* 3: 204.

Sunda, W. G., and Huntsman S. A., 1986, Relationships among growth rate, cellular manganese concentrations and manganese transport kinetics in estuarine and oceanic species of the diatom *Thalassiosira*, *J. Phycol.* 22: 259–270.

Sunda, W. G., and Huntsman, S. A., 1988, Effect of sunlight on redox cycles of manganese in the southwestern Sargasso Sea, *Deep Sea Res*. 35: 1297–1317.

Sunda, W. G., and Huntsman, S. A., 1992, Feedback interactions between zinc and phytoplankton in seawater, *Limnol. Oceanogr.* 37: 25–40.

Sunda, W. G., and Huntsman, S. A., 1995, Cobalt and zinc interreplacement in marine phytoplankton: biological and geochemical implications, *Limnol. Oceanogr.* 40: 1404–1417.

Sunda, W. G., and Huntsman, S. A., 1995, Iron uptake and growth limitation in oceanic and coastal phytoplankton, *Mar. Chem.* 50: 189–206.

Sunda, W. G., and Huntsman, S. A., 1997, Interrelated influence of iron, light and cell size on marine phytoplankton growth, *Nature* 390: 389–392.

Sunda, W. G., and Huntsman, S. A., 1998, Interactions among Cu²⁺, Zn²⁺, and Mn²⁺ in controlling cellular Mn, Zn, and growth rate in the coastal alga *Chlamydomonas*, *Limnol. Oceanogr.* 43:1055–1064.

Sunda, W. G., and Huntsman, S. A., 2000, Effect of Zn, Mn, and Fe on Cd accumulation in phytoplankton: implications for oceanic Cd cycling, *Limnol. Oceanogr.* 45: 1501–1516.

Surge, D.M., Lohmann, K.C., Goodfriend, G.A., 2003, Reconstructing estuarine conditions: oyster shells as recorders of environmental change, Southwest Florida, *Estuar. Coast Shelf Sci.* 57: 737–756.

Szefer, P., Szefer, K., Skwarzec, B., 1990, Distribution of trace metals in some representative fauna of the southern Baltic, *Marine Pollution Bulletin* 21(2): 60-62.

Tabouret, H., Pomerleau, S., Aurélie Jolivet, A., Pécheyran, C., Riso, R., Thébault, J., Chauvaud, L., Amouroux, D., 2012, Specific pathways for the incorporation of dissolved barium and molybdenum into the bivalve shell: An isotopic tracer approach in the juvenile great scallop (Pecten maximus), *Marine Environmental Research* (2012), doi:0.1016/j.marenvres.2012.03.006.

Takesue, R. K., and van Geen, A., 2004, Mg/Ca, Sr/Ca, and stable isotopes in modern and Holocene Protothaca staminea shells from a northern California coastal upwelling region, *Geochim. Cosmochim. Acta* 68: 3845–3861.

Takesue, R. K., Bacon, C. R., Thompson, J. K., 2008, Influences of organic matter and calcification rate on trace elements in aragonitic estuarine bivalve shells, *Geochimica et Cosmochimica Acta* 72(22): 5431-5445.

Tanaka, N., Monaghan, M. C., Rye, D. M., 1986, Contribution of metabolic carbon to mollusc and barnacle shell carbonate, *Nature* 320: 520–523.

Tang, B., Liu, B., Wang, G., Zhang, T., Xiang, J., 2006, Effects of Various algal diets and starvation on larval growth and survival of Metrix metrix, *Aquaculture* 254(1-4): 526-533

Taylor, J. D., 1973, The structural evolution of the bivalve shell, *Palaeontology* 16: 519-534.

Thébault, J., Chauvaud, L., L'Helguen, S., Clavier, J., Barats, A., Jacquet, S., Pécheyran, C., Amouroux, D., 2009, Barium and molybdenum records in bivalve shells: Geochemical proxies for phytoplankton dynamics in in coastal environments? *Limnology and Oceanography* 54:1002-1014.

Thebault, J., and Chauvaud, L., 2012, Li/Ca enrichments in gret scallop shells (Pecten maximus) and their relationship with phytoplankton blooms, *Palaeogeography Palaeoclimatology Palaeoecology* 373: 108-122.

Thompson, T. G., and Chow, J., 1955, The strontium---calcium atom ratio in the carbonate secreting marine organisms, *Deep Sea Res. Suppl.* 3: 20--39.

Thompson, P. A., Guo, M.-X., Harrison, P. J., 1993, The influence of irradiance on the biochemical composition of three phytoplankton species and their nutritional value for larvae of the Pacific oyster (Crassostrea gigas), *Marine Biology* 117: 259-268.

Thompson, P. A., Guo, M. X., Harrison, P. J., Whyte, J. N., 2004, Effects of variation in temperature on the fatty acid composition of eight species of marine phytoplankton, *Journal of phycology* 28(4): 488-497.

Thorrold, S.R., Jones, G. P., Hellberg, M. E., Burton, R. S., Swearer, S., Neigel, J. E., Morgan, S. E., Warner, R. R., 2002, Quantifying larval retention and connectivity in marine populations with artificial and natural markers, *Bull. Mar. Sci.* 70:291-308.

Thorrold, S. R., Zacherl, D. C., Levin, L. A., 2007, Population connectivity and larval dispersal: using geochemical signatures in calcified structures. *Oceanogr.* 20: 81-88.

Tsujii, T., 1968, Studies on the mechanism of shell and pearl formation. X. The submicroscopic structure of the epithelial cells on the mantle of the pearl oyster, Petria martensii, *Rep. Fish. Mie Univ.* 6:41-58.

Tung-Yuan Ho, T. –Y., Wen, L. –S., You, C. –F., Lee, D.-C., 2007, The Trace-Metal Composition of Size-Fractionated Plankton in the South China Sea: Biotic versus Abiotic Sources, *Limnology and Oceanography* 52(5): 1776-1788.

Vander Putten, E., Dehairs F., Andre´ L., and Baeyens W., 1999, Quantitative in situ microanalysis of minor and trace elements in biogenic calcite using infrared laser ablation-inductively coupled plasma mass spectrometry: A critical evaluation, Anal. Chim. Acta 378: 261–272.

Vander Putten, E., Dehairs, F., Keppens, E., Baeyens, W., 2000, High resolution distribution of trace elements in the calcite shell layer of modern Mytilus edulis: environmental and biological controls, Geochimica et Cosmochimica Acta 64: 997-1011.

Wanamaker, A.D. Jr., Kruetz, K.J., Borns, H.W. Jr., Introne, D.S., Feindal, S., Barber, B.J., 2006, An Aquaculture Based method for calibrated bivalve isotope paleothermometry, Geochemistry Geophysics Geosystems 7(9).

Wanamaker, A.D. Jr., Kruetz, K.J., Borns H.W. Jr., Introne, D.S., Feindal, S., Funder, S., Barber, B.J., 2007, Experimental determination of salinity, temperature, growth and metabolic effects on shell isotope chemistry of Mytilis edulis collected from Maine and Greenland, Peleoceanography 22: PA2217.

Wanamaker, A.D. Jr., Kreutz, K.J., Schöne, B.R., Pettigrew, N., Borns, H.W., Introne, D.S., Belknap, D., Maasch, K.A., Feindel, S., 2008, Coupled North Atlantic slope water forcing on Gulf of Maine temperatures over the past millennium, Clim. Dyn. 31:183–194.

Wang, W. –X., 2002, Interactions of trace metals and different marine food chains, Mar. Ecol. Prog. Ser. 243:295-309.

Wang, W. -X., and Fisher, N.S., 1996, Assimilation of trace elements and carbon by the mussel Mytilus edulis: effects of food composition, Limnology and Oceanography 41(2): 197-207.

Wang, X., Liu, B., Xiang, J., 2003, Potential role of iron and ferritin subunit (MmeFer) for larval shell formation in clam Metrix metrix, J Exp Biol Ecol.

Ward, J.E., and Shumway, S.E., 2004, Separating the grain from the chaff: particle selection in suspension- and deposit-feeding bivalves, Journal of Experimental Marine Biology and Ecology 300: 83-130.

Watabe, N. (1965). Studies on shell formation: XI. Crystal—matrix relationships in the inner layers of mollusk shells, *Journal of ultrastructure research*, *12*(3): 351-370.

Watabe, N., and Wilbur, K. M., 1960, Influence of the organic matrix on crystal type in molluscs, Nature (London) 188:334.

Watabe, N., Kingsley, R. J., and Kawaguchi, T., 1993, Functions of organic matrices in some invertebrate calcifying systems, in: Structure, Formation and Evolution of Fossil Hard Tissues (Kobayashi, I., Mutvei, H. and Sahni, A., eds.), pp. 3–11, Tokai Univ. Press, Japan.

Watanabe, T., Minagawa, M., Oba, T. and Winter, A., 2001, Pretreatment of coral aragonite for Mg and Sr analysis: Implications for coral thermometers, Geochem. J. 35: 265–269.

Weiner, S., and Traub, W., 1980, X-ray diffraction study of the insoluble organic matrix of mollusk shell, FEBS 111: 311–316.

Weiner, S., and Addadi, L., 1991, Acidic macromolecules of mineralized tissues: the controllers of crystal formation. Trends in biochemical science 16: 252-256.

Weiner, S., and Addadi, L., 1997, Design strategies in mineralized biological materials, J. Mater. Chem. 7: 689-702.

Weiner, S., and Dove, P.M., 2003, An overview of biomineralization processes and the problem of the vital effect, in: Reviews in Mineralogy and Geochemistry: Biomineralization (Dove, P.M., De Yoreo, J.J. and Weiner, S., eds.), pp. 1–29, Mineralogical Society of America and Geochemical Society, Washington.

Weiss, I.M., Renner, C., Strigl, M.G., Fritz, M., 2002, A simple and reliable method for the determination and localization of chitin in abalone nacre, Chem. Mater. 14: 3252–3259.

Wen-Xiong, W., and Fisher, N. S., 1996, Assimilation of trace elements by the mussel Mytilus edulis: effects of diatom chemical composition, Marine Biology 125(4): 715-724.

Wheeler, A., 1992, Mechanisms of molluscan shell formation, in: Calcification in biological systems (Bonucci E., ed.). CRC, Boca Raton, pp. 179-216.

Wheeler, A. P., and Wilbur, K. M., 1977, Shell growth in the scallop, Argopecten irradians. II. Processes of shell growth, J. Mollusc. Stud. 43:155-161.

Wheeler A.P., Rusenko K.W., George J.W., Sikes C.S., 1987, Evaluation of calcium binding by molluscan shell organic matrix and its relevance to biomineralization, Comp. Biochem. Physiol. 87B:953–960.

White, L. K., Szabo, A., Carkner, P., Chasteen, N. D., 1977, An electron paramagnetic study of Mn II in the aragonite lattice of a clam shell, Mya arenaria, J. Phys. Chem. 81:1420-1424.

Whyte, J.N.C., 1987, Biochemical composition and energy content of six species of phytoplankton used in mariculture of bivalves, *Aquaculture* 60(3–4): 231-241.

Whyte, J.N.C., Boume, N., Hodgson, C.A., 1989, Influence of algal diets on biochemical composition and energy reserves in Patinopecten yessoensis (Jay) larvae, Aquaculture 78: 333-347.

Wikfors, G.H., Twarog, J.W. and Ukeles, R., 1984. Influence of chemical composition of algal food sources on growth of juvenile oysters, Crassostrea virginica. Biol. Bull. 167: 251-263.

Wilbur, K. M., 1964, Shell formation and regeneration, in: Physiology of Mollusca, Vol. 1 (K. M. Wilbur and C. M. Yonge, eds.), pp. 243-282, Academic Press, New York.

Wilbur, K. M., 1972, Shell formation in molluscs, in: Chemical Zoology, Vol. VII, Mollusca (M. Florkin and B. Scheer, eds.), pp. 103-154, Academic Press, New York.

Wilbur, K. M., and Simkiss, K., 1968, Calcified shells, in: Comprehensive Biochemistry 26A (M. Florkin and E. H. Stotz, eds.), pp. 229-295, Elsevier, Amsterdam.

Wilbur, K. M., and Saleuddin, A. S. M., 1983, Shell Formation, in: The Mollusca (ed. A. S. M. Saleuddin and K. M. Wilbur), pp. 236–279. Acad. Press, New York.

Windisch, W., 2001/2002, Interaction of chemical species with biological regulation of the metabolism of essential trace elements, Anal. Bioanal. chem. 372:421-425.

Witbaard, R., Duineveld, C.A., Bergman, M., 2001, The effect of tidal resuspension on benthic food quality on the southern North Sea, Senckenbergiana maritima 31:225–234.

Wolfe, K. H., Chilingar, G. V., and Beals, F.W., 1967, Elemental composition of carbonate skeletons, minerals, and sediments, in: Carbonate Rocks, Developments in Sedimentology, Vol. 9B (G. V. Chilingar, H. J. Bissel, and R. W. Fairbridge, eds.), pp. 23-150, Elsevier, Amsterdam.

Xiaolin Hou, Xiaojun Yan, 1998, Study on the concentration and seasonal variation of inorganic elements in 35 species of marine algae, Science of The Total Environment 222(3): 141-156.

Zhang, Y., Xie, L., Meng, Q., Jiang, T., Pu, R., Chen, L., Zhang, R., 2003, A novel matrix protein participating in the nacre framework formation of pearl oyster, Pinctada fucata, Comp Biochem Physiol 135B:565–573.

Zhu, C. J., Lee, Y. K., Chao, T. M., 1997, Effects of temperature and growth phase on lipid and biochemical composition of Isochrysis galbana TK1, *Journal of applied phycology 9*(5): 451-457.

Zischke, J. A., Watabe, N., and Wilbur, K. M., 1970, Studies on Shell Formation: Measurement of growth in the gastropod Ampullarius glaucus, Malacologia 10:423-439.

Appendix A:

Chapter 4 Post Hoc analysis results

diets					
All Pairwise Multiple Comparison Procedures (Dunn's Method) :					
Comparison	Diff of Ranks	Q	P<0.05		
Cg vs. ISO	114.869	5.411	Yes		
Cm vs. ISO	107.152	4.967	Yes		
CTRL vs. ISO	93.427	4.562	Yes		
Mixed vs. Cm	73.134	3.305	Yes		
Mixed vs. CTRL	86.859	4.124	Yes		
Mixed vs. ISO	180.286	8.104	Yes		
Mixed vs. Pav609	112.777	4.981	Yes		
Pav1209 vs. ISO	117.822	5.331	Yes		
TI vs. ISO	116.359	5.265	Yes		
Cg vs. Cm	7.716	0.366	No		
Cg vs. CTRL	21.441	1.073	No		
Cg vs. Pav609	47.36	2.188	No		
Cm vs. CTRL	13.725	0.675	No		
Cm vs. Pav609	39.643	1.804	No		
CTRL vs. Pav609	25.918	1.24	No		
Mixed vs. Cg	65.417	3.002	No		
Mixed vs. Pav1209	62.464	2.759	No		
Mixed vs. TI	63.927	2.824	No		
Pav1209 vs. Cg	2.953	0.136	No		
Pav1209 vs. Cm	10.669	0.485	No		
Pav1209 vs. CTRL	24.394	1.167	No		
Pav1209 vs. Pav609	50.313	2.237	No		
Pav1209 vs. TI	1.463	0.065	No		
Pav609 vs. ISO	67.509	3.055	No		

Table A1: Results summary of the post hoc analyses (Dunn's Method) for (B/P)/Ca differences between diets

Table A1 (Continued)

Comparison	Diff of Ranks	Q	P<0.05
TI vs. Cg	1.49	0.0689	No
TI vs. Cm	9.207	0.419	No
TI vs. CTRL	22.932	1.097	No
TI vs. Pav609	48.85	2.172	No

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Cg vs. ISO	141.186	6.421	Yes	
Cm vs. ISO	126.495	5.606	Yes	
CTRL vs. ISO	129.922	6.066	Yes	
Mixed vs. ISO	181.038	8.194	Yes	
Mixed vs. Pav1209	77.854	3.502	Yes	
Mixed vs. Pav609	78.621	3.49	Yes	
Pav1209 vs. ISO	103.184	4.52	Yes	
Pav609 vs. ISO	102.417	4.431	Yes	
TI vs. ISO	129.57	5.71	Yes	
Cg vs. Cm	14.691	0.672	No	
Cg vs. CTRL	11.264	0.545	No	
Cg vs. Pav1209	38.002	1.718	No	
Cg vs. Pav609	38.768	1.729	No	
Cg vs. TI	11.616	0.528	No	
Cm vs. Pav1209	23.311	1.027	No	
Cm vs. Pav609	24.077	1.047	No	
CTRL vs. Cm	3.427	0.161	No	
CTRL vs. Pav1209	26.738	1.24	No	
CTRL vs. Pav609	27.505	1.258	No	
CTRL vs. TI	0.352	0.0164	No	
Mixed vs. Cg	39.852	1.865	No	
Mixed vs. Cm	54.544	2.484	No	
Mixed vs. CTRL	51.116	2.459	No	
Mixed vs. TI	51.469	2.33	No	
Pav1209 vs. Pav609	0.767	0.033	No	
TI vs. Cm	3.075	0.136	No	
TI vs. Pav1209	26.386	1.156	No	
TI vs. Pav609	27.152	1.175	No	

Table A2: Results summar	v of the	post hoc analys	ses (Dunn's	Method) for ((B/P)) differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Cg vs. ISO	122.247	5.725	Yes	
Cm vs. ISO	138.969	6.405	Yes	
CTRL vs. ISO	95.778	4.65	Yes	
Mixed vs. CTRL	91.824	4.398	Yes	
Mixed vs. ISO	187.602	8.494	Yes	
Mixed vs. Pav1209	77.426	3.443	Yes	
Mixed vs. Pav609	81.901	3.642	Yes	
Mixed vs. TI	77.651	3.453	Yes	
Pav1209 vs. ISO	110.176	4.957	Yes	
Pav609 vs. ISO	105.701	4.756	Yes	
TI vs. ISO	109.951	4.947	Yes	
Cg vs. CTRL	26.468	1.317	No	
Cg vs. Pav1209	12.071	0.555	No	
Cg vs. Pav609	16.546	0.76	No	
Cg vs. TI	12.296	0.565	No	
Cm vs. Cg	16.722	0.788	No	
Cm vs. CTRL	43.191	2.111	No	
Cm vs. Pav1209	28.793	1.303	No	
Cm vs. Pav609	33.268	1.505	No	
Cm vs. TI	29.018	1.313	No	
Mixed vs. Cg	65.355	3.023	No	
Mixed vs. Cm	48.633	2.214	No	
Pav1209 vs. CTRL	14.398	0.685	No	
Pav1209 vs. Pav609	4.475	0.198	No	
Pav1209 vs. TI	0.225	0.00994	No	
Pav609 vs. CTRL	9.923	0.472	No	
TI vs. CTRL	14.173	0.674	No	
TI vs. Pav609	4.25	0.188	No	

Table A3: Results summary of the post hoc analyses (Dunn's Method) for (Ba/P)/Ca differences between diets

Table A4: Results summary of the post hoc analyses (Dunn's Method) for (Ba/P) differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :					
Comparison	Diff of Ranks	Q	P<0.05		
Cg vs. ISO	128.058	5.873	Yes		
Cm vs. ISO	144.482	6.457	Yes		
CTRL vs. ISO	122.712	5.777	Yes		

Table A4 (Continued)			
Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. ISO	88.741	4.089	Yes
Pav1209 vs. ISO	91.709	4.051	Yes
Pav609 vs. ISO	121.183	5.286	Yes
TI vs. ISO	77.379	3.33	Yes
Cg vs. CTRL	5.346	0.261	No
Cg vs. Mixed	39.317	1.874	No
Cg vs. Pav1209	36.349	1.656	No
Cg vs. Pav609	6.874	0.309	No
Cg vs. TI	50.679	2.247	No
Cm vs. Cg	16.425	0.758	No
Cm vs. CTRL	21.77	1.031	No
Cm vs. Mixed	55.741	2.584	No
Cm vs. Pav1209	52.773	2.344	No
Cm vs. Pav609	23.299	1.022	No
Cm vs. Tl	67.103	2.904	No
CTRL vs. Mixed	33.971	1.666	No
CTRL vs. Pav1209	31.003	1.45	No
CTRL vs. Pav609	1.528	0.0705	No
CTRL vs. TI	45.333	2.059	No
Mixed vs. TI	11.362	0.506	No
Pav1209 vs. Mixed	2.968	0.136	No
Pav1209 vs. TI	14.33	0.613	No
Pav609 vs. Mixed	32.442	1.466	No
Pav609 vs. Pav1209	29.474	1.278	No
Pav609 vs. TI	43.805	1.853	No

Table A5: Results summary of the post hoc analyses (Dunn's Method) for (Co/P)/Ca differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Pav1209 vs. CTRL	44.029	3.162	Yes	
Mixed vs. CTRL	34.595	2.887	No	
Pav1209 vs. ISO	34.267	2.38	No	
Cg vs. Cm	2.458	0.207	No	
Cg vs. CTRL	29.512	2.231	No	
Cg vs. ISO	19.75	1.439	No	
Cg vs. Pav609	13.32	1.074	No	
Cm vs. CTRL	27.054	2.392	No	

Comparison	Diff of Ranks	Q	P<0.05
Cm vs. ISO	17.292	1.454	No
Cm vs. Pav609	10.862	1.052	No
ISO vs. CTRL	9.762	0.738	No
Mixed vs. Cg	5.083	0.406	No
Mixed vs. Cm	7.542	0.719	No
Mixed vs. ISO	24.833	1.981	No
Mixed vs. Pav609	18.404	1.664	No
Mixed vs. TI	2.238	0.149	No
Pav1209 vs. Cg	14.517	1.008	No
Pav1209 vs. Cm	16.975	1.341	No
Pav1209 vs. Mixed	9.433	0.711	No
Pav1209 vs. Pav609	27.837	2.119	No
Pav1209 vs. TI	11.671	0.704	No
Pav609 vs. CTRL	16.192	1.367	No
Pav609 vs. ISO	6.43	0.519	No
TI vs. Cg	2.845	0.178	No
TI vs. Cm	5.304	0.367	No
TI vs. CTRL	32.357	2.078	No
TI vs. ISO	22.595	1.413	No
TI vs. Pav609	16.165	1.087	No

Table A6: Results summary of the post hoc analyses (Dunn's Method) for (Fe/P)/Ca differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Cm vs. Cg	65.925	3.141	Yes	
Cm vs. ISO	68.015	3.153	Yes	
Mixed vs. Cg	102.604	4.763	Yes	
Mixed vs. CTRL	97.942	4.705	Yes	
Mixed vs. ISO	104.694	4.734	Yes	
Pav1209 vs. Cg	93.503	4.371	Yes	
Pav1209 vs. CTRL	88.841	4.299	Yes	
Pav1209 vs. ISO	95.593	4.351	Yes	
Cm vs. CTRL	61.263	3.026	No	
Mixed vs. TI	61.183	2.699	No	
Pav609 vs. ISO	64.368	2.93	No	
Cg vs. ISO	2.09	0.099	No	
Cm vs. Pav609	3.647	0.167	No	

Table A6 (Contiunued)			
Comparison	Diff of Ranks	Q	P<0.05
Cm vs. Tl	24.504	1.107	No
CTRL vs. Cg	4.662	0.236	No
CTRL vs. ISO	6.752	0.331	No
Mixed vs. Cm	36.679	1.668	No
Mixed vs. Pav1209	9.101	0.407	No
Mixed vs. Pav609	40.326	1.802	No
Pav1209 vs. Cm	27.578	1.262	No
Pav1209 vs. Pav609	31.225	1.404	No
Pav1209 vs. TI	52.082	2.312	No
Pav609 vs. Cg	62.278	2.911	No
Pav609 vs. CTRL	57.616	2.788	No
Pav609 vs. TI	20.857	0.926	No
TI vs. Cg	41.422	1.909	No
TI vs. CTRL	36.759	1.752	No
TI vs. ISO	43.511	1.954	No

Table A7: Results summary of the post hoc analyses (Dunn's Method) for (Fe/P) differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Pav609 vs. Cg	81.171	3.664	Yes	
Pav609 vs. ISO	80.629	3.526	Yes	
Cm vs. Cg	58.412	2.688	No	
Cm vs. CTRL	33.763	1.603	No	
Cm vs. ISO	57.87	2.578	No	
Cm vs. Mixed	9.256	0.426	No	
Cm vs. Tl	41.728	1.836	No	
CTRL vs. Cg	24.649	1.206	No	
CTRL vs. ISO	24.108	1.137	No	
CTRL vs. TI	7.966	0.37	No	
ISO vs. Cg	0.542	0.0248	No	
Mixed vs. Cg	49.156	2.327	No	
Mixed vs. CTRL	24.507	1.199	No	
Mixed vs. ISO	48.615	2.223	No	
Mixed vs. TI	32.473	1.466	No	
Pav1209 vs. Cg	64.506	2.95	No	
Pav1209 vs. Cm	6.094	0.271	No	
Pav1209 vs. CTRL	39.857	1.879	No	

Comparison	Diff of Ranks Q		P<0.05
Pav1209 vs. ISO	63.964	2.832	No
Pav1209 vs. Mixed	15.35	0.702	No
Pav1209 vs. TI	47.823	2.092	No
Pav609 vs. Cm	22.759	1.001	No
Pav609 vs. CTRL	56.522	2.628	No
Pav609 vs. Mixed	32.015	1.445	No
Pav609 vs. Pav1209	16.665	0.729	No
Pav609 vs. TI	64.488	2.787	No
TI vs. Cg	16.683	0.753	No
TI vs. ISO	16.142	0.706	No

Table A8: Results summary of the post hoc analyses (Dunn's Method) for (K/P)/Ca differences between
diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. CTRL	73.426	3.558	Yes
Mixed vs. ISO	75.382	3.466	Yes
Cg vs. CTRL	42.074	2.187	No
Cg vs. ISO	44.03	2.156	No
Cg vs. TI	25.694	1.149	No
Cm vs. Cg	11.571	0.57	No
Cm vs. CTRL	53.645	2.755	No
Cm vs. ISO	55.601	2.693	No
Cm vs. Pav1209	7.136	0.335	No
Cm vs. Pav609	2.936	0.14	No
Cm vs. TI	37.265	1.651	No
CTRL vs. ISO	1.956	0.0998	No
Mixed vs. Cg	31.351	1.463	No
Mixed vs. Cm	19.78	0.914	No
Mixed vs. Pav1209	26.917	1.202	No
Mixed vs. Pav609	22.717	1.027	No
Mixed vs. TI	57.046	2.418	No
Pav1209 vs. Cg	4.435	0.21	No
Pav1209 vs. CTRL	46.509	2.29	No
Pav1209 vs. ISO	48.465	2.261	No
Pav1209 vs. TI	30.129	1.293	No
Pav609 vs. Cg	8.635	0.415	No
Pav609 vs. CTRL	50.709	2.535	No

Table A8 (Continued)

Comparison	Diff of Ranks	Q	P<0.05
Pav609 vs. ISO	52.665	2.49	No
Pav609 vs. Pav1209	4.2	0.193	No
Pav609 vs. TI	34.329	1.49	No
TI vs. CTRL	16.38	0.758	No
TI vs. ISO	18.336	0.808	No

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
Cm vs. ISO	67.17	3.15	Yes
Mixed vs. ISO	77.864	3.651	Yes
Cg vs. CTRL	32.346	1.647	No
Cg vs. ISO	50.219	2.405	No
Cg vs. TI	34.625	1.526	No
Cm vs. Cg	16.952	0.817	No
Cm vs. CTRL	49.298	2.451	No
Cm vs. Pav1209	13.644	0.62	No
Cm vs. Pav609	0.77	0.0355	No
Cm vs. TI	51.577	2.232	No
CTRL vs. ISO	17.873	0.883	No
CTRL vs. TI	2.279	0.103	No
Mixed vs. Cg	27.645	1.332	No
Mixed vs. Cm	10.693	0.504	No
Mixed vs. CTRL	59.991	2.982	No
Mixed vs. Pav1209	24.337	1.105	No
Mixed vs. Pav609	11.464	0.528	No
Mixed vs. TI	62.27	2.695	No
Pav1209 vs. Cg	3.308	0.153	No
Pav1209 vs. CTRL	35.654	1.7	No
Pav1209 vs. ISO	53.526	2.417	No
Pav1209 vs. TI	37.933	1.59	No
Pav609 vs. Cg	16.181	0.76	No
Pav609 vs. CTRL	48.527	2.348	No
Pav609 vs. ISO	66.4	3.039	No
Pav609 vs. Pav1209	12.874	0.571	No
Pav609 vs. TI	50.806	2.154	No
TI vs. ISO	15.594	0.672	No

Table A9: Results summary of the post hoc analyses (Dunn's Method) for (K/P) differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
Cg vs. ISO	86.382	4.17	Yes
Cm vs. ISO	93.122	4.471	Yes
CTRL vs. ISO	86.858	4.393	Yes
Mixed vs. ISO	142.2	6.528	Yes
Mixed vs. Pav609	87.826	3.964	Yes
Pav1209 vs. ISO	82.399	3.861	Yes
TI vs. ISO	93.724	4.133	Yes
Cg vs. Pav1209	3.983	0.189	No
Cg vs. Pav609	32.008	1.516	No
Cm vs. Cg	6.739	0.327	No
Cm vs. CTRL	6.264	0.319	No
Cm vs. Pav1209	10.723	0.505	No
Cm vs. Pav609	38.748	1.826	No
CTRL vs. Cg	0.476	0.0244	No
CTRL vs. Pav1209	4.459	0.221	No
CTRL vs. Pav609	32.484	1.609	No
Mixed vs. Cg	55.818	2.589	No
Mixed vs. Cm	49.079	2.265	No
Mixed vs. CTRL	55.342	2.679	No
Mixed vs. Pav1209	59.801	2.699	No
Mixed vs. TI	48.476	2.067	No
Pav1209 vs. Pav609	28.025	1.29	No
Pav609 vs. ISO	54.374	2.548	No
TI vs. Cg	7.342	0.327	No
TI vs. Cm	0.602	0.0267	No
TI vs. CTRL	6.866	0.318	No
TI vs. Pav1209	11.325	0.492	No
TI vs. Pav609	39.35	1.708	No

Table A10: Results summary of the post hoc analyses (Dunn's Method) for (Li/P)/Ca differences between diets

Table A11: Results summary of the post hoc analyses (Dunn's Method) for (Li/P) differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :					
Comparison Diff of Ranks Q P<0.05					
Cg vs. ISO	115.098 5.48 Yes				
Cm vs. ISO 110.215 5.138 Ye					
CTRL vs. ISO 125.724 6.175 Yes					

Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. ISO	130.555	6.087	Yes
Pav1209 vs. ISO	75.455	3.411	Yes
Pav609 vs. ISO	90.054	4.098	Yes
TI vs. ISO	99.139	4.283	Yes
Cg vs. Cm	4.884	0.234	No
Cg vs. Pav1209	39.643	1.838	No
Cg vs. Pav609	25.044	1.169	No
Cg vs. TI	15.959	0.706	No
Cm vs. Pav1209	34.76	1.58	No
Cm vs. Pav609	20.16	0.923	No
Cm vs. TI	11.076	0.481	No
CTRL vs. Cg	10.626 0.538		No
CTRL vs. Cm	15.509	0.767	No
CTRL vs. Pav1209	50.269	2.401	No
CTRL vs. Pav609	35.669	1.716	No
CTRL vs. TI	26.585	1.207	No
Mixed vs. Cg	15.457	0.74	No
Mixed vs. Cm	20.341	0.954	No
Mixed vs. CTRL	4.832	0.239	No
Mixed vs. Pav1209	55.101	2.505	No
Mixed vs. Pav609	40.501	1.853	No
Mixed vs. TI	31.417	1.364	No
Pav609 vs. Pav1209	14.6	0.649	No
TI vs. Pav1209	23.684	1.001	No
TI vs. Pav609	9.084	0.386	No

Table A11 (Continued)

Table A12: Results summary of the post hoc analyses (Dunn's Method) for (Mg/P)/Ca differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks Q F			
Cg vs. ISO	78.114	3.701	Yes	
Cm vs. ISO	121.729	5.675	Yes	
CTRL vs. ISO	84.475 4.149		Yes	
Mixed vs. Cg	74.517 3.439		Yes	
Mixed vs. CTRL	68.157	68.157 3.255		
Mixed vs. ISO 152.631 6.9		6.901	Yes	
Mixed vs. Pav1209	82.413	3.661	Yes	
Mixed vs. Pav609	95.513	4.243	Yes	

Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. TI	78.433	3.44	Yes
Pav1209 vs. ISO	70.218	3.196	Yes
TI vs. ISO	74.198	3.332	Yes
Cg vs. Pav1209	7.896	0.367	No
Cg vs. Pav609	20.996	0.976	No
Cg vs. TI	3.916	0.179	No
Cm vs. Cg	43.615	2.079	No
Cm vs. CTRL	37.255	1.841	No
Cm vs. Pav1209	51.511	2.357	No
Cm vs. Pav609	64.611	2.957	No
Cm vs. Tl	47.531	2.146	No
CTRL vs. Cg	6.361	0.32	No
CTRL vs. Pav1209	14.257	0.686	No
CTRL vs. Pav609	27.357	1.316	No
CTRL vs. TI	10.277	0.487	No
Mixed vs. Cm	30.902	1.405	No
Pav1209 vs. Pav609	13.1	0.586	No
Pav609 vs. ISO	57.118	2.599	No
TI vs. Pav1209	3.98	0.176	No
TI vs. Pav609	17.08	0.754	No

Table A12 (Continued)

Table A13: Results summary of the post he	oc analyses (Dunn's	s Method) for (Mg/P)	differences between
diets			

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
Cg vs. ISO	101.91	4.699	Yes
Cm vs. ISO	137.851	6.194	Yes
Cm vs. TI	103.77	4.577	Yes
CTRL vs. ISO	151.067	7.151	Yes
CTRL vs. Pav1209	83.319	3.891	Yes
CTRL vs. TI	116.986	5.425	Yes
Mixed vs. ISO	127.771	5.834	Yes
Mixed vs. TI	93.69	4.197	Yes
Pav609 vs. ISO	87.23	3.826	Yes
Cg vs. Pav1209	34.163	1.555	No
Cg vs. Pav609	14.68	0.664	No
Cg vs. Tl	67.83	3.067	No
Cm vs. Cg	35.941	1.667	No

Table A13 (Continued)			
Comparison	Diff of Ranks	Q	P<0.05
Cm vs. Mixed	10.08	0.463	No
Cm vs. Pav1209	70.103	3.112	No
Cm vs. Pav609	50.62	2.233	No
CTRL vs. Cg	49.157	2.411	No
CTRL vs. Cm	13.216	0.63	No
CTRL vs. Mixed	23.296	1.13	No
CTRL vs. Pav609	63.836	2.96	No
Mixed vs. Cg	25.861	1.221	No
Mixed vs. Pav1209	60.023	2.707	No
Mixed vs. Pav609	40.54	1.816	No
Pav1209 vs. ISO	67.747	2.991	No
Pav1209 vs. TI	33.667	1.46	No
Pav609 vs. Pav1209	19.483	0.845	No
Pav609 vs. TI	53.15	2.29	No
TI vs. ISO	34.08	1.495	No

Table A14: Results summary of the post hoc analyses (Dunn's Method) for (Mn/P)/Ca differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
Cg vs. CTRL	81.388	4.084	Yes
Cm vs. Cg	85.662	4.071	Yes
Cm vs. CTRL	167.05	8.233	Yes
Cm vs. ISO	75.309	3.501	Yes
ISO vs. CTRL	91.741	4.493	Yes
Mixed vs. Cg	135.393	6.231	Yes
Mixed vs. CTRL	216.781	10.323	Yes
Mixed vs. ISO	125.041	5.637	Yes
Mixed vs. Pav609	76.036	3.368	Yes
Pav1209 vs. Cg	76.557	3.548	Yes
Pav1209 vs. CTRL	157.945	7.577	Yes
Pav609 vs. CTRL	140.745	6.752	Yes
TI vs. Cg	87.445	4.024	Yes
TI vs. CTRL	168.833	8.04	Yes
TI vs. ISO	77.092	3.475	Yes
Cm vs. Pav1209	9.105	0.415	No
Cm vs. Pav609	26.305	1.2	No
ISO vs. Cg	10.353	0.489	No

Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. Cm	49.731	2.254	No
Mixed vs. Pav1209	58.836	2.606	No
Mixed vs. TI	47.949	2.111	No
Pav1209 vs. ISO	66.205	3.004	No
Pav1209 vs. Pav609	17.2	0.767	No
Pav609 vs. Cg	59.357	2.751	No
Pav609 vs. ISO	49.005	2.224	No
TI vs. Cm	1.783	0.0808	No
TI vs. Pav1209	10.887	0.482	No
TI vs. Pav609	28.087	1.244	No

Table A15: Results summary of the post hoc analyses	s (Dunn's Method) for (Mn/P) differences between
diets	

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
Cg vs. CTRL	81.535	3.966	Yes
Cm vs. Cg	85.192	3.92	Yes
Cm vs. CTRL	166.727	7.877	Yes
Cm vs. ISO	74.519	3.321	Yes
ISO vs. CTRL	92.208	4.329	Yes
Mixed vs. Cg	104.541	4.919	Yes
Mixed vs. CTRL	186.077	9.002	Yes
Mixed vs. ISO	93.868	4.272	Yes
Pav1209 vs. CTRL	135.415	6.315	Yes
Pav609 vs. Cg	83.873	3.761	Yes
Pav609 vs. CTRL	165.408	7.607	Yes
Pav609 vs. ISO	73.2	3.184	Yes
TI vs. Cg	69.705	3.147	Yes
TI vs. CTRL	151.241	7.005	Yes
Cm vs. Pav1209	31.313	1.387	No
Cm vs. Pav609	1.319	0.0577	No
Cm vs. TI	15.487	0.682	No
ISO vs. Cg	10.673	0.488	No
Mixed vs. Cm	19.349	0.886	No
Mixed vs. Pav1209	50.662	2.291	No
Mixed vs. Pav609	20.669	0.923	No
Mixed vs. TI	34.836	1.565	No
Pav1209 vs. Cg	53.879	2.449	No

Table A15 (Continued)

Comparison	Diff of Ranks	Q	P<0.05
Pav1209 vs. ISO	43.206	1.903	No
Pav609 vs. Pav1209	29.993	1.297	No
Pav609 vs. TI	14.167	0.609	No
TI vs. ISO	59.032	2.584	No
TI vs. Pav1209	15.826	0.689	No

Table A16: Results summary of the post hoc analyses (Dunn's Method) for (Si/P)/Ca differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
Mixed vs. ISO	69.106	3.147	Yes
Pav1209 vs. Cg	110.926	5.126	Yes
Pav1209 vs. Cm	77.984	3.548	Yes
Pav1209 vs. CTRL	75.02	3.589	Yes
Pav1209 vs. ISO	132.912	6.014	Yes
Pav609 vs. Cg	69.801	3.225	Yes
Pav609 vs. ISO	91.787	4.153	Yes
TI vs. ISO	85.206	3.804	Yes
Cg vs. ISO	21.986	1.036	No
Cm vs. Cg	32.942	1.561	No
Cm vs. ISO	54.928	2.546	No
CTRL vs. Cg	35.906	1.797	No
CTRL vs. Cm	2.964	0.146	No
CTRL vs. ISO	57.892	2.827	No
Mixed vs. Cg	47.119	2.192	No
Mixed vs. Cm	14.177	0.649	No
Mixed vs. CTRL	11.214	0.54	No
Pav1209 vs. Mixed	63.807	2.854	No
Pav1209 vs. Pav609	41.125	1.828	No
Pav1209 vs. TI	47.707	2.093	No
Pav609 vs. Cm	36.859	1.677	No
Pav609 vs. CTRL	33.895	1.621	No
Pav609 vs. Mixed	22.682	1.014	No
Pav609 vs. TI	6.582	0.289	No
TI vs. Cg	63.219	2.881	No
TI vs. Cm	30.278	1.359	No
TI vs. CTRL	27.314	1.287	No
TI vs. Mixed	16.1	0.711	No

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
CTRL vs. ISO	80.815	3.794	Yes
Pav1209 vs. Cg	75.645	3.438	Yes
Pav1209 vs. ISO	103.239	4.548	Yes
Pav1209 vs. Mixed	85.777	3.898	Yes
Pav609 vs. Cg	84.059	3.77	Yes
Pav609 vs. Cm	76.338	3.339	Yes
Pav609 vs. ISO	111.654	4.857	Yes
Pav609 vs. Mixed	94.192	4.224	Yes
TI vs. ISO	76.529	3.329	Yes
Cg vs. ISO	27.595	1.262	No
Cg vs. Mixed	10.133	0.479	No
Cm vs. Cg	7.722	0.355	No
Cm vs. ISO	35.317	1.574	No
Cm vs. Mixed	17.855	0.822	No
CTRL vs. Cg	53.22	2.589	No
CTRL vs. Cm	45.498	2.15	No
CTRL vs. Mixed	63.352	3.082	No
CTRL vs. TI	4.285	0.197	No
Mixed vs. ISO	17.462	0.799	No
Pav1209 vs. Cm	67.923	3.009	No
Pav1209 vs. CTRL	22.425	1.046	No
Pav1209 vs. TI	26.71	1.155	No
Pav609 vs. CTRL	30.84	1.418	No
Pav609 vs. Pav1209	8.415	0.364	No
Pav609 vs. TI	35.125	1.501	No
TI vs. Cg	48.934	2.194	No
TI vs. Cm	41.213	1.803	No
TI vs. Mixed	59.067	2.649	No

Table A17: Results summary of the post hoc analyses (Dunn's Method) for (Si/P) differences between diets

Table A18: Results summary of the post hoc analyses (Dunn's Method) for (Sr/P)/Ca differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Cg vs. ISO	107.723	5.35	Yes	
Cm vs. ISO	110.844	5.289	Yes	
CTRL vs. ISO	82.581	4.252	Yes	
Table A18 (Continued)				
-----------------------	---------------	-------	--------	--
Comparison	Diff of Ranks	Q	P<0.05	
Mixed vs. CTRL	80.596	3.973	Yes	
Mixed vs. ISO	163.177	7.627	Yes	
Mixed vs. Pav1209	89.303	3.844	Yes	
Mixed vs. Pav609	83.332	3.829	Yes	
Pav1209 vs. ISO	73.873	3.286	Yes	
Pav609 vs. ISO	79.844	3.81	Yes	
TI vs. ISO	96.231	4.498	Yes	
Cg vs. CTRL	25.142	1.327	No	
Cg vs. Pav1209	33.85	1.533	No	
Cg vs. Pav609	27.879	1.358	No	
Cg vs. TI	11.492	0.548	No	
Cm vs. Cg	3.121	0.152	No	
Cm vs. CTRL	28.264	1.426	No	
Cm vs. Pav1209	36.971	1.619	No	
Cm vs. Pav609	31	1.453	No	
Cm vs. TI	14.614	0.672	No	
CTRL vs. Pav1209	8.707	0.406	No	
CTRL vs. Pav609	2.736	0.138	No	
Mixed vs. Cg	55.454	2.645	No	
Mixed vs. Cm	52.332	2.405	No	
Mixed vs. TI	66.946	3.018	No	
Pav609 vs. Pav1209	5.971	0.262	No	
TI vs. CTRL	13.65	0.673	No	
TI vs. Pav1209	22.357	0.962	No	
TI vs. Pav609	16.386	0.753	No	

Table A19: Results summary of the post hoc analyses (Dunn's Method) for (Sr/P) differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Cg vs. ISO	137.971	6.62	Yes	
Cm vs. ISO	149.743	6.835	Yes	
CTRL vs. ISO	125.375	6.175	Yes	
Mixed vs. ISO	162.158	7.665	Yes	
Mixed vs. Pav1209	78.338	3.443	Yes	
Mixed vs. TI	71.975	3.315	Yes	
Pav1209 vs. ISO	83.82	3.631	Yes	
Pav609 vs. ISO	114.843	5.242	Yes	

Comparison	Diff of Ranks	Q	P<0.05
TI vs. ISO	90.183	4.089	Yes
Cg vs. CTRL	12.596	0.643	No
Cg vs. Pav1209	54.15	2.411	No
Cg vs. Pav609	23.128	1.088	No
Cg vs. TI	47.788	2.233	No
Cm vs. Cg	11.772	0.554	No
Cm vs. CTRL	24.368	1.176	No
Cm vs. Pav1209	65.923	2.811	No
Cm vs. Pav609	34.9	1.565	No
Cm vs. TI	59.56	2.654	No
CTRL vs. Pav1209	41.555	1.892	No
CTRL vs. Pav609	10.532	0.508	No
CTRL vs. TI	35.192	1.686	No
Mixed vs. Cg	24.188	1.181	No
Mixed vs. Cm	12.415	0.576	No
Mixed vs. CTRL	36.783	1.846	No
Mixed vs. Pav609	47.315	2.194	No
Pav609 vs. Pav1209	31.023	1.323	No
Pav609 vs. TI	24.66	1.099	No
TI vs. Pav1209	6.362	0.27	No

Table A19 (Continued)

Table A20: Results summary of the post hoc analyses (Dunn's Method) for (Zn/P)/Ca differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
Cg vs. ISO	58.313	3.719	Yes	
Cm vs. ISO	56	3.212	Yes	
Mixed vs. CTRL	45.979	3.329	Yes	
Mixed vs. ISO	84.039	4.876	Yes	
Mixed vs. Pav609	48.971	3.196	Yes	
Pav1209 vs. ISO	72.25	3.973	Yes	
Cg vs. Cm	2.313	0.16	No	
Cg vs. CTRL	20.252	1.714	No	
Cg vs. Pav609	23.244	1.715	No	
Cg vs. Tl	9.229	0.557	No	
Cm vs. CTRL	17.939	1.276	No	
Cm vs. Pav609	20.932	1.346	No	
Cm vs. TI	6.917	0.379	No	

Table A20 (Continued)					
Comparison	Diff of Ranks	Q	P<0.05		
CTRL vs. ISO	38.061	2.479	No		
CTRL vs. Pav609	2.993	0.227	No		
Mixed vs. Cg	25.727	1.815	No		
Mixed vs. Cm	28.039	1.742	No		
Mixed vs. Pav1209	11.789	0.698	No		
Mixed vs. TI	34.956	1.937	No		
Pav1209 vs. Cg	13.938	0.91	No		
Pav1209 vs. Cm	16.25	0.95	No		
Pav1209 vs. CTRL	34.189	2.283	No		
Pav1209 vs. Pav609	37.182	2.269	No		
Pav1209 vs. TI	23.167	1.222	No		
Pav609 vs. ISO	35.068	2.096	No		
TI vs. CTRL	11.023	0.678	No		
TI vs. ISO	49.083	2.55	No		
TI vs. Pav609	14.015	0.798	No		

Table A21: Results summary of the post hoc analyses (Dunn's Method) for (Zn/P) differences between diets

All Pairwise Multiple Comparison Procedures (Dunn's Method) :						
Comparison	Diff of Ranks	Diff of Ranks Q				
Cg vs. ISO	62.538	3.989	Yes			
Mixed vs. ISO	68.692	3.986	Yes			
Cg vs. Cm	19.387	1.345	No			
Cg vs. CTRL	17.988	1.523	No			
Cg vs. Pav1209	22.743	1.485	No			
Cg vs. Pav609	26.178	1.932	No			
Cg vs. TI	16.776	1.013	No			
Cm vs. ISO	43.151	2.475	No			
Cm vs. Pav1209	3.356	0.196	No			
Cm vs. Pav609	6.79	0.437	No			
CTRL vs. Cm	1.399	0.0995	No			
CTRL vs. ISO	44.55	2.902	No			
CTRL vs. Pav1209	4.755	0.317	No			
CTRL vs. Pav609	8.19	0.622	No			
Mixed vs. Cg	6.154	0.434	No			
Mixed vs. Cm	25.541	1.587	No			
Mixed vs. CTRL	24.142	1.748	No			
Mixed vs. Pav1209	28.896	1.71	No			

Table A21 (Continued)

Comparison	Diff of Ranks	Diff of Ranks Q			
Mixed vs. Pav609	32.331	2.11	No		
Mixed vs. TI	22.93	1.271	No		
Pav1209 vs. ISO	39.795	2.189	No		
Pav1209 vs. Pav609	3.435	0.21	No		
Pav609 vs. ISO	36.36	2.174	No		
TI vs. Cm	2.611	0.143	No		
TI vs. CTRL	1.212	0.0745	No		
TI vs. ISO	45.762	2.377	No		
TI vs. Pav1209	5.967	0.315	No		
TI vs. Pav609	9.402	0.535	No		

Appendix B:

Chapter 5 post hoc analysis summaries

Table B1: Statistical analy	ysis results summar	y – Dunn's pairwise co	mparison procedure – for B/Ca
-----------------------------	---------------------	------------------------	-------------------------------

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
July vs. January	28.077	3.714	Yes	
July vs. March	21.133	2.916	Yes	
July vs. May	42.4	5.16	Yes	
July vs. October	43.067	5.943	Yes	
March vs. October	21.933	2.831	Yes	
January vs. May	14.323	1.605	No	
January vs. October	14.99	1.864	No	
March vs. January	6.944	0.864	No	
March vs. May	21.267	2.455	No	
May vs. October	0.667	0.077	No	

Table B2: Statistical anal	ysis results summar	y – Dunn's pairwise	comparison	procedure – for Ba/Ca
	5	1		

All Pairwise Multiple Comparison Procedures (Dunn's Method) :				
Comparison	Diff of Ranks	Q	P<0.05	
January vs. July	40.848	5.404	Yes	
January vs. March	23.356	2.905	Yes	
January vs. May	38.223	4.283	Yes	
January vs. October	30.156	3.751	Yes	
March vs. July	17.492	2.414	No	
March vs. May	14.867	1.716	No	
March vs. October	6.8	0.878	No	
May vs. July	2.625	0.319	No	
October vs. July	10.692	1.475	No	
October vs. May	8.067	0.931	No	

All Pairwise Multiple Comparison Procedures (Holm-Sidak method):					
	Overa	all significand	ce level = 0.05		
Comparison	Diff of Means	t	Unadjusted P	Critical Level	Significant?
January vs. July	7.39E-07	4.723	0.000272	0.006	Yes
March vs. July	5.60E-07	3.575	0.00276	0.009	Yes
October vs. January	5.59E-07	2.96	0.00974	0.01	Yes
October vs. July	1.30E-06	7.101	0.00000362	0.005	Yes
October vs. March	7.38E-07	3.911	0.00139	0.007	Yes
October vs. May	1.24E-06	4.145	0.000864	0.006	Yes
January vs. March	1.80E-07	1.099	0.289	0.025	No
January vs. May	6.78E-07	2.396	0.0301	0.013	No
March vs. May	4.99E-07	1.761	0.0985	0.017	No
May vs. July	6.08E-08	0.218	0.831	0.05	No

Table B3: Statistical analysis results summary -	pairwise comparison	procedure – for Co/Ca
--	---------------------	-----------------------

Table B4: Statistical analysis results summary – Dunn's pairwise comparison procedure – for Fe/Ca

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
January vs. March	39.056	4.858	Yes
January vs. May	40.323	4.518	Yes
July vs. March	29.283	4.041	Yes
July vs. May	30.55	3.718	Yes
October vs. March	30.8	3.976	Yes
October vs. May	32.067	3.702	Yes
January vs. July	9.773	1.293	No
January vs. October	8.256	1.027	No
March vs. May	1.267	0.146	No
October vs. July	1.517	0.209	No

Table B5: Statistical anal	ysis results summar	y – Dunn's	pairwise co	mparison	procedure -	– for K/Ca

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
January vs. July	32.808	4.34	Yes
March vs. July	42.3	5.837	Yes
March vs. October	24.533	3.167	Yes
May vs. July	31.5	3.833	Yes
January vs. May	1.308	0.147	No
January vs. October	15.041	1.871	No
March vs. January	9.492	1.181	No

Table B5 (Continued)

Comparison	Diff of Ranks	Q	P<0.05
March vs. May	10.8	1.247	No
May vs. October	13.733	1.585	No
October vs. July	17.767	2.452	No

Table B6: Statistical analysis results summary – Dunn's pairwise comparison procedure – for Li/Ca

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
January vs. July	30.815	4.077	Yes
January vs. October	45.749	5.69	Yes
March vs. October	30.6	3.95	Yes
May vs. October	26.333	3.04	Yes
January vs. March	15.149	1.884	No
January vs. May	19.415	2.176	No
July vs. October	14.933	2.061	No
March vs. July	15.667	2.162	No
March vs. May	4.267	0.493	No
May vs. July	11.4	1.387	No

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
March vs. January	26.833	3.31	Yes
March vs. July	41.667	5.829	Yes
March vs. October	39.333	5.147	Yes
May vs. July	27.8	3.43	Yes
May vs. October	25.467	2.981	Yes
January vs. July	14.833	1.941	No
January vs. October	12.5	1.542	No
March vs. May	13.867	1.623	No
May vs. January	12.967	1.447	No
October vs. July	2.333	0.326	No

Table B7: Statistical analysis results summary – Dunn's pairwise comparison procedure – for Mg/Ca

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
January vs. July	42.233	5.526	Yes
January vs. May	49.283	5.5	Yes
January vs. October	23.55	2.905	Yes
March vs. July	27.35	3.826	Yes
March vs. May	34.4	4.026	Yes
October vs. May	25.733	3.012	Yes
January vs. March	14.883	1.836	No
July vs. May	7.05	0.87	No
March vs. October	8.667	1.134	No
October vs. July	18.683	2.614	No

Table B8: Statistical analysis results summary – Dunn's pairwise comparison procedure – for Mn/Ca

Table B9: Statistical analysis results summary – Dunn's pairwise comparison procedure – for Si/Ca

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
January vs. May	28.9	3.135	Yes
July vs. March	29.233	4.266	Yes
July vs. May	36.2	4.659	Yes
October vs. March	25.8	3.522	Yes
October vs. May	32.767	4.001	Yes
January vs. March	21.933	2.593	No
July vs. January	7.3	0.907	No
July vs. October	3.433	0.501	No
March vs. May	6.967	0.851	No
October vs. January	3.867	0.457	No

Table B10: Statistical analysis results summary – Dunn's pairwis	vise comparison procedure – f	or Zn/Ca
--	-------------------------------	----------

All Pairwise Multiple Comparison Procedures (Dunn's Method) :			
Comparison	Diff of Ranks	Q	P<0.05
January vs. July	23.917	3.71	Yes
January vs. October	33.333	4.317	Yes
March vs. July	28.143	4.523	Yes
March vs. October	37.56	4.984	Yes
January vs. May	12.167	1.84	No
July vs. October	9.417	1.201	No
March vs. January	4.226	0.696	No
March vs. May	16.393	2.564	No

Table B10 (Continued)

Comparison	Diff of Ranks	Q	P<0.05
May vs. July	11.75	1.741	No
May vs. October	21.167	2.654	No