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Cryptic biodiversity and phylogeographic patterns of Seychellois *Ligia* isopods

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Ligia isopods are conspicuous inhabitants of rocky intertidal habitats exhibiting several biological traits that severely limit their dispersal potential. Their presence in patchy habitats and low vagility may lead to long term isolation, allopatric isolation and possible cryptic speciation. Indeed, various species of *Ligia* have been suggested to represent instead cryptic species complexes. Past studies; however, have largely focused in Eastern Pacific and Atlantic species of *Ligia*, leaving in doubt whether cryptic diversity occurs in other highly biodiverse areas. The Seychelles consists of 115 islands of different ages and geological origins spread across the western Indian Ocean. They are well known for their rich biodiversity with recent reports of cryptic species in terrestrial Seychellois organisms. Despite these studies, it is unclear whether coastal invertebrates from the Seychelles harbor any cryptic diversity. In this study, we examined patterns of genetic diversity and isolation within *Ligia* isopods across the Seychelles archipelago by characterizing individuals from locations across both inner and outer islands of the Seychelles using mitochondrial and nuclear markers. We report the presence of highly divergent lineages of independent origin. At Aldabra Atoll, we uncovered a lineage closely related to the *Ligia vitiensis* cryptic species complex. Within the inner islands of Cousine, Silhouette, and Mahé we detected the presence of two moderately divergent and geographically disjunct lineages most closely related to *Ligia dentipes*. Our findings suggest that the Seychelles may harbor at least three novel species of *Ligia* in need of description and that these species may have originated independently.

1 **Cryptic biodiversity and phylogeographic patterns of Seychellois *Ligia* isopods**

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18 **ABSTRACT**

19 *Ligia* isopods are conspicuous inhabitants of rocky intertidal habitats exhibiting several
20 biological traits that severely limit their dispersal potential. Their presence in patchy habitats
21 and low vagility may lead to long term isolation, allopatric isolation and possible cryptic
22 speciation. Indeed, various species of *Ligia* have been suggested to represent instead cryptic
23 species complexes. Past studies; however, have largely focused in Eastern Pacific and Atlantic
24 species of *Ligia*, leaving in doubt whether cryptic diversity occurs in other highly biodiverse
25 areas. The Seychelles consists of 115 islands of different ages and geological origins spread
26 across the western Indian Ocean. They are well known for their rich biodiversity with recent
27 reports of cryptic species in terrestrial Seychellois organisms. Despite these studies, it is unclear
28 whether coastal invertebrates from the Seychelles harbor any cryptic diversity. In this study, we
29 examined patterns of genetic diversity and isolation within *Ligia* isopods across the Seychelles
30 archipelago by characterizing individuals from locations across both inner and outer islands of
31 the Seychelles using mitochondrial and nuclear markers. We report the presence of highly
32 divergent lineages of independent origin. At Aldabra Atoll, we uncovered a lineage closely
33 related to the *Ligia vitiensis* cryptic species complex. Within the inner islands of Cousine,
34 Silhouette, and Mahé we detected the presence of two moderately divergent and geographically
35 disjunct lineages most closely related to *Ligia dentipes*. Our findings suggest that the Seychelles
36 may harbor at least three novel species of *Ligia* in need of description and that these species may
37 have originated independently.

38

39

40 INTRODUCTION

41 The Seychelles encompass some 115 islands (Stoddart, 1984) spread across a large swath of the
42 western Indian Ocean. These islands can be broadly divided into outer and inner islands based
43 on their geological origin, geographic locality, and unique geological histories (Braithwaite
44 1984). The “outer” islands are low-lying coralline islands that vary in size, age and geographic
45 distribution, with some closer to Africa than to other Seychelles islands (e.g. Aldabra Atoll), and
46 others closer to the “inner” islands. The inner islands, on the other hand, consist of granitic
47 islands thought to have formed as the Indian and Madagascar Plates separated during the breakup
48 of Gondwana some 65 million years ago (Mya) (Plummer & Belle 1995). The inner islands
49 comprise of some 40 islands, including major ones such as Mahé, Praslin and Silhouette as well
50 as smaller islands encircling them (Fig 1). The complex geological history of the Seychelles
51 islands coupled with their remoteness may help explain a richly biodiverse fauna characterized
52 by high rates of endemism and for the presence of highly divergent cryptic lineages recently
53 reported across a variety of Seychellois terrestrial (e.g. Rocha et al. 2011; Rocha et al. 2013;
54 Silva et al. 2010) and freshwater (e.g. Daniels 2011) organisms. These recent reports indicate
55 that our understanding of the Seychelles fauna may be incomplete and that cryptic species may
56 exist in poorly studied Seychellois organisms. This is particularly true for organisms in which
57 cryptic lineages have been reported from other highly isolated archipelagos.

58 *Ligia* isopods have been reported to exhibit high levels of cryptic diversity in several
59 regions of the world, including highly isolated habitats such as the Hawaiian islands (Santamaria
60 et al. 2013; Taiti et al. 2003). In this remote and isolated archipelago, *Ligia hawaiiensis* was
61 thought to be the only intertidal species of this genus to occupy this habitat (Schmalzfuss 2003).
62 This species, however, appears to represent a paraphyletic taxon composed of several highly

63 divergent (>10% COI K2P) lineages (Santamaria et al. 2013). These results indicate that this
64 species may be actually several cryptic species. The high levels of divergence observed in *L.*
65 *hawaiensis* match those reported in other *Ligia* species (Eberl et al. 2013; Hurtado et al. 2010;
66 Jung 2008; Santamaria et al. 2014; Santamaria et al. 2013; Taiti et al. 2003) and are thought to be
67 a product of their biological traits that severely restrict dispersal abilities: poor desiccation
68 resistance, poor ability to swim, direct development, and inhabitation of patchy rocky habitats.
69 Such traits, when combined with the isolation of oceanic archipelagos can easily disrupt gene
70 flow leading to local cryptic speciation in populations. This leads to the question of whether
71 *Ligia* populations in isolated and remote islands such as those of the Seychelles may also harbor
72 cryptic lineages.

73 In the Seychelles, *Ligia* isopods have been reported from both inner (Taiti 2014) and
74 outer islands (Ferrara & Taiti 1985). The earliest published record from the Seychelles
75 archipelago indicates the presence of *Ligia exotica* at Aldabra (Ferrara & Taiti 1985). This
76 record is based on of a single 8-mm long juvenile specimen collected at Aldabra Atoll in 1983.
77 More recently, the presence of *Ligia dentipes*, a species redescribed and illustrated by Ferrara
78 and Taiti (1982), from the island of Silhouette was reported (Taiti 2014). These reports are
79 based on morphological identification and in some cases of partial or incomplete organisms,
80 casting doubt on some of the identifications (e.g. *L. exotica* from Aldabra) and leaving
81 unanswered whether Seychelles *Ligia* harbor highly divergent lineages. In this study, we apply
82 molecular and morphological approaches to *Ligia* individuals collected in both the inner and
83 outer islands to determine which *Ligia* species inhabit the Seychelles and whether they harbor
84 any highly divergent cryptic lineages that may present putative cryptic species.

85

86 MATERIALS AND METHODS

87 We collected *Ligia* individuals by hand in locations in the inner and outer islands of the
88 Seychelles Archipelago, Zanzibar and mainland Tanzania, and Sri Lanka. All specimens were
89 collected during 2014, field-preserved in 70% Ethanol, and frozen upon arrival at the laboratory.
90 Collections in the Seychelles were carried out under Seychelles Bureau of Standards permit
91 A0157. The data produced from these samples was complemented using publicly available
92 sequences for four *Ligia* species: *L. occidentalis* (Mexico), *L. exotica* (Mexico), *L. dentipes*
93 (Thailand), and *L. vitiensis* (Indonesia, Timor-Leste, Madagascar). Detailed information for all
94 localities is provided in Table 1.

95 We extracted total genomic DNA from pereopods/pleopods for 10 individuals per
96 locality following the solid tissue protocol of the ZR Quick-gDNA Miniprep Kit (Zymo
97 Research). During dissections, male specimens were identified to species by visually inspecting
98 the appendix masculina of the second pleopod and comparing it to illustrations from species
99 descriptions and re-illustrations (Ferrara & Taiti 1982; Khalaji-Pirbalouty & Wägele 2010; Taiti
100 et al. 2003; Taiti et al. 1992). For *Ligia* collected in the Seychelles, we PCR amplified a 710-bp
101 segment of the Cytochrome Oxidase I mitochondrial gene (hereafter COI) and a 710-bp fragment
102 of the α -subunit of the Sodium-Potassium ATPase gene (hereafter NaK) for 1–10 individuals per
103 locality using previously published primers and conditions (COI: Folmer et al. 1994; NaK: Tsang
104 et al. 2008). We PCR-amplified an additional three mitochondrial genes for a subset of
105 individuals from each locality in the Seychelles (see Fig 1, Table 1): (a) ~490-bp of the 16S
106 rDNA gene (primers 16Sar/16Sbr; Palumbi 1996); ~495-bp of 12S rDNA (primers crust-
107 12Sf/crust-12Sr; Podsiadlowski & Bartolomaeus 2005); and a 361-bp fragment of the
108 Cytochrome-b (Cytb) gene (primers 144F/151F and 270R/272R; Merritt et al. 1998). For *Ligia*

109 from localities outside the Seychelles, we amplified all four mitochondrial genes mentioned
110 above for 1–2 individuals per locality (see Table 1). Positive PCR amplicons were identified
111 using gel electrophoresis prior to cleaning and sequencing at the University of Arizona Genetics
112 Core (UAGC). Sequences were assembled and edited (i.e. primers removed) using Geneious
113 R8.0.2. No evidence indicative of pseudo-genes (i.e. no gaps, indels, early stop codons) was
114 observed in any of the protein coding genes used in this study (i.e., NaK, cyt-b, and COI).

115 We used TCS v1.21 (Clement et al. 2000) to visualize the relationships between COI
116 haplotypes recovered from *Ligia* from all Seychelles localities to both determine the geographic
117 distribution of COI haplotypes recovered within the archipelago and the levels of fine scale
118 divergence within the archipelago. We calculated the 95% most parsimonious branch
119 connections between haplotypes under the cladogram estimation algorithm of Templeton et al.
120 (1992) with all other settings as default. We also calculated Kimura 2-Parameter distances
121 (K2P) within and between localities for COI in MEGA v7.0.7 (Kumar et al. 2016). We estimated
122 relationships between NaK alleles using a similar approach.

123 We combined the four mitochondrial gene fragments produced in this study with publicly
124 available sequences for other *L. vitiensis* from the Indo-Pacific and for two other *Ligia* species to
125 be used as outgroups (i.e. *Ligia occidentalis* and *Ligia exotica*). Locality and sequence
126 information for these sequences is presented in Table 1. Since ribosomal genes used in this
127 study exhibit secondary structure that may lead to ambiguous alignments, we aligned each gene
128 dataset separately using MAFFT v.7.0 (Kato & Standley 2013) as implemented under the
129 GUIDANCE2 algorithm (Sela et al. 2015) in the GUIDANCE server
130 (<http://mafft.cbrc.jp/alignment/server/>) using 100 bootstrap replicates and all other settings as
131 default. This approach produces confidence scores for each nucleotide position and sequence in

132 the alignment that aid in the non-biased removal of misaligned nucleotide positions. We
133 excluded nucleotide positions from our final alignment if they: (a) produced a confidence score
134 below 1.00 in the Guidance alignments, and/or (b) if they exhibited obvious misalignments. We
135 used MEGA v7.0.7 (Kumar et al. 2016) to estimate pairwise K2P genetic distances for COI and
136 for the 16S rDNA genes, after excluding ambiguously aligned sites.

137 We used jModeltest v2.1 (Darriba et al. 2012) to determine the most appropriate model of
138 nucleotide evolution for the final concatenated mitochondrial alignment as well as for each
139 mitochondrial gene. We selected from 1,624 models by evaluating their likelihoods on a fixed
140 BioNJ-JC tree under the Bayesian Information Criterion (BIC). We used the most likely model
141 of nucleotide evolution chosen by jModeltest in our phylogenetic reconstructions with two
142 general exceptions. When the chosen model was not implemented by the software, we applied
143 the next more complex model implemented. When the chosen model implemented the joint
144 estimation of Γ and I, we used a simpler $+\Gamma$ model as the joint estimation of Γ and I can be
145 problematic (see RAxML manual; and pages 113-114 of Yang 2006).

146 We carried out Maximum Likelihood phylogenetic reconstructions in RAxML v8.2.6
147 (Stamatakis 2014; Stamatakis et al. 2008) and GARLI v2.0 (Zwickl 2006). In RAxML, we used
148 the Rapid Bootstrap Algorithm to carry out 1,000 bootstrap replicate searches followed by a
149 thorough ML search under the GTR $+\Gamma$ model. All other settings were as default. Searches in
150 GARLI consisted of 1,000 bootstrap replicates using the appropriate model of evolution
151 identified by jModeltest, with all other settings as default. For each search, we produced a
152 majority-rule consensus tree of all bootstrap replicates using the SumTrees command of
153 DendroPy v3.10.1 (Sukumaran & Holder 2010).

154 We also carried out phylogenetic reconstructions under Bayesian Inference. Parameters
155 used in MrBayes v3.1.2 (Huelsenbeck & Ronquist 2001; Ronquist & Huelsenbeck 2003)
156 searches are presented in Table 2. All other parameters used were as default. We also carried out
157 Bayesian searches implementing polytomy priors (Lewis et al. 2005) under Phycas v2.2.0 (Lewis
158 et al. 2015) as to ensure that support values produced by MrBayes were not overestimated (i.e.,
159 “star-tree paradox”) (Suzuki et al. 2002). We determined if Bayesian analyses had reached
160 stationarity prior to estimating the posterior probability for each node by building majority-rule
161 consensus trees of the stationary stage of each run using the SumTrees command (Sukumaran &
162 Holder 2010). Samples prior to stationarity were discarded as “burnin.”

163 We repeated all phylogenetic searches under three partitioning schemes: (a) all positions
164 within a single partition; (b) positions partitioned by gene; and (c) the best partitioning scheme
165 according to the BIC implemented in PartitionFinder v1.0.0 (Lanfear et al. 2012).
166 PartitionFinder searches were run using the following settings: branch lengths = linked; models =
167 all; model selection = BIC; search = greedy; and an *a priori* partitioning scheme accounting for
168 codon positions and genes.

169

170 RESULTS

171 We sequenced COI and NaK for a total of 50 individuals from across five locations in the
172 Seychelles archipelago. All individuals from the inner islands of the Seychelles were putatively
173 identified as *L. dentipes* based on the gonopod morphology, while those from Aldabra were
174 putatively assigned to *L. vitiensis*. All sequences produced in this study have been deposited in
175 GenBank under accession numbers: XXXXXX-XXXXXXX.

176

177 *COI and NaK haplotype networks*

178 For COI, we observed a total of 16 haplotypes divided into three networks (Fig 3). Two
179 haplotypes separated by two mutational steps were found solely in individuals collected at
180 Aldabra. A second haplotype network contained all COI individuals collected from Cousine
181 island (E1) and consisted of seven haplotypes separated by 2–9 steps. A third and final network
182 consisted of the seven COI haplotypes recovered from Silhouette (E2) and Mahé (E3, E4) which
183 were separated by 2–15 steps. We recovered two alleles for the NaK gene, with all *Ligia*
184 individuals from the inner islands (E2–E4) sharing a single allele that diverged by 27 steps from
185 an allele found in all the individuals from Aldabra (E1).

186

187 *Phylogenetic reconstructions*

188 Our concatenated mitochondrial dataset included a total of 16 *Ligia* individuals: five from
189 localities within the Seychelles, nine from other Indo-Pacific localities, and the two outgroups.
190 The final concatenated alignment included 2019 nucleotides, of which 292 positions could not be
191 confidently aligned and were excluded for the phylogenetic analyses (16S rDNA: 138; 12S
192 rDNA: 154). Of the resulting 1733 nucleotides positions, 579 were parsimony informative.
193 jModeltest identified a model consisting of three substitution rates (rate matrix: 012010; see
194 jModeltest manual) as well as +F, +I, and + Γ parameters for our final concatenated
195 mitochondrial dataset under the BIC, AIC, and AICc. We applied this model in GARLI
196 analyses; however, the more complex GTR + Γ model was applied in all other searches as the
197 chosen model is not available in the other software packages (e.g. RAxML, Phycas). The use of
198 GTR was justified as it was included in the 99% cumulative weight interval under all three
199 selection criteria.

200 Mitochondrial phylogenetic reconstructions (Fig 2) recovered a well-supported split [100
201 Bootstrap Support (BS) and Posterior Probability (PP)] between the *Ligia* specimens from the
202 Indo-Pacific and the two outgroup taxa. Within the Indo-Pacific *Ligia*, we observed a split
203 between two highly divergent lineages: an ‘East African’ clade (*Clade ABC*; BS: 71–96; PP: 91–
204 99) and an ‘Asian’ clade (*Clade DEF*; BS: 100; PP: 100). COI K2P divergences between these
205 two clades ranged from 21.8–27.8% with a mean divergence of 25.2%.

206

207 The ‘East African’ clade (*Clade ABC*) contained all specimens identified as *L. vitiensis*
208 and included the localities of Aldabra Atoll in the Seychelles, Nosy Bay in Madagascar, three
209 localities in Tanzania (i.e., Dar-Es-Salaam, Kilwa Masoko, and Stone Town), and two localities
210 from the Indonesian Archipelago (Komodo, Indonesia and Dili, Timor-Leste). These localities
211 formed three distinct lineages: (a) *Ligia* from Madagascar, mainland Tanzania, and Aldabra
212 (Seychelles) were placed in the well-supported *Clade A* (light orange in all figures; BS: 100; PP:
213 100); (b) Indonesian archipelago *Ligia* were placed in *Clade B* (cyan in all figures; BS: 100; PP:
214 100); and (c) *Clade C* was composed of *Ligia* from Stone Town, Zanzibar, Tanzania (light grey
215 in all figures). Although some analyses suggested a sister relationship between clades *A* and *B*,
216 supports for this relationship was low in ML analyses (BS: < 60) and highly variable in BI (PP:
217 71–100). COI K2P divergences between the three main lineages in the East African clade
218 ranged from 7.12–9.09% (Table 3).

219 The ‘Asian’ clade (*DEF*) included all individuals identified as *L. dentipes* which were
220 separated into three main lineages: (a) *Clade D* (red in all figures; BS: 91–100, PP:100) included
221 *Ligia* from Galle (Sri Lanka) and Phuket (Thailand); (b) *Clade E* (blues in all figures; BS: 88–
222 100; PP: 100) consisting of *Ligia* from the inner islands of Seychelles; and (c) *Clade F* (green in

223 all figures) represented solely from *Ligia* collected in Trincomalee, Sri Lanka. As in *Clade ABC*,
224 relationships within the ‘Asian’ clade were not well resolved. We observed a sister relationship
225 between clades *E* and *F*; however, support values for this relationship were often low (BS: <60–
226 74; PP: <60–65). COI K2P divergences between the three main lineages in the Asian clade
227 ranged from 12.07–14.83% (Table 3). Divergences within the inner Seychelles locations (i.e.
228 *Clade E*) ranged from 0.5–5.2% with all comparisons to the island of Cousine being above 4.5%
229 (Table 4).

230

231 **DISCUSSION**

232 To date, two species of *Ligia* have been reported to inhabit intertidal habitats in the Seychelles
233 archipelago: *L. dentipes* (Taiti 2014) and *L. exotica* (Ferrara & Taiti 1985). The former is
234 considered to be endemic to coastlines in the north-eastern Indian Ocean, with confirmed records
235 from the Nicobar and Andaman islands, the Maldives, Sri Lanka, and the Seychelles (Taiti 2014
236 and references therein). *Ligia exotica*, on the other hand, is a species of possible East-Asian
237 origin (Hurtado in preparation) thought to have a cosmopolitan distribution due to human-aided
238 introductions. These records; however, are based on morphological identifications, leaving in
239 doubt their validity and whether they represent highly divergent genetic lineages as reported for
240 *Ligia* species in other regions (Eberl et al. 2013; Hurtado et al. 2010; Santamaria et al. 2014;
241 Santamaria et al. 2013; Taiti et al. 2003). By applying molecular approaches to *Ligia* individuals
242 from both inner and outer islands in the Seychelles, we have determined that *Ligia* diversity in
243 the Seychelles archipelago is underreported. Our phylogenetic reconstructions and
244 morphological identifications suggest the presence in the Seychelles of highly divergent lineages
245 belonging to at least two species complexes: *L. vitiensis* and *L. dilatata*. *Ligia* from the outer

246 island of Aldabra represent a highly divergent lineage within a clade composed of *L. vitiensis*
247 individuals from around the Indo-Pacific, while those from the inner islands of Cousin, Mahé,
248 and Silhouette represent two highly divergent lineages within a clade composed of *L. dentipes*
249 individuals from Asia. These findings are supported by male gonopod morphology and suggest
250 that *Ligia* have colonized the Seychelles archipelago on at least independent two occasions.

251 Phylogenetic reconstructions place *Ligia* individuals collected at Aldabra Atoll, in *Clade*
252 *ABC* with *L. vitiensis* individuals collected from East Africa and Madagascar. Despite the poorly
253 resolved relationships within this clade, the geological history of Aldabra, oceanographic
254 patterns in the region, and previous phylogeographic findings suggest a Malagasy or East
255 African origin to be the most likely source of the *Ligia* populations in Aldabra. This outer island
256 is a low lying atoll that originated some 20 Mya from coral reefs growing on volcanic seamounts
257 (Plummer 1995). The atoll does not appear to have experienced any connection to continental
258 landmasses or other nearby-islands, but has undergone several periods of submersion and
259 emergence, including what is thought to have been a complete submersion some 125,000 years
260 ago (Braithwaite et al. 1973). Such geological history would seem to preclude Aldabra as the
261 place of origin for *Ligia* in the *ABC clade*. Colonizers instead appear more likely to have
262 originated in either Madagascar or East Africa given the proximity of Aldabra to these
263 continental landmasses (~420km from the coastline of Madagascar and ~650km from the East
264 African coastline).

265 *Ligia* individuals collected from the inner Seychelles islands of Cousin, Mahé, and
266 Silhouette were placed in *Clade DEF* with *L. dentipes* individuals from Asia indicating either an
267 Asian origin for these populations, or a Seychellois origin for *L. dentipes*. Although the
268 relationships within *Clade DEF* are not well resolved, our findings suggest the Seychelles

269 lineage to have evolved from an Asian ancestor: phylogenetic reconstructions place *Ligia* from
270 the inner islands in *Clade E*, which is suggested to have split from an Asian ancestor (though
271 poorly supported). Furthermore, the COI K2P divergences observed between *Ligia* from the
272 inner islands and other *Ligia* from the region range from 12.07 to 27.62% (*L. dentipes*: 12.07–
273 14.83% COI K2P; *L. vitiensis*: 21.85–27.62% COI K2P). These divergences would require
274 mutation rates that are about six times lower than the mutation rate for COI reported for other
275 marine isopods (2.5%/My; Ketmaier et al. 2003) in order to match the age of the inner
276 Seychelles islands (~65 My; Plummer & Belle 1995). These findings, though not conclusive,
277 would suggest an Asian origin for *Ligia* from the inner islands.

278

279 The finding that *Ligia* from the inner islands are most closely related to those from Asia
280 represent a rare, yet not exceptional, occurrence for Seychellois flora and fauna. Overall, fauna
281 endemic to the Seychelles appears to largely be of an East African or Malagasy origin
282 (Agnarsson & Kuntner 2012 and references therein). Such findings are consistent with the closer
283 proximity of these islands to East African and Madagascar than to Asian coastlines, and the
284 predominant oceanic currents in the area. Nonetheless, an Asian affinity has been reported for
285 several Seychellois organisms such as the Sooglossidae and Nasikabatrachidae anuran families
286 (Biju & Bossuyt 2003), coleopterans (Gerlach 2009), and *Nepenthes* pitcher plants (Meimberg et
287 al. 2001). Such affinity may be explained by Gondwanan organisms rafting on the Indian
288 continent until its collision with Asia (Karanth 2006) or by the over-water dispersal from Asia to
289 the Seychelles or vice-versa. This latter mechanism has been suggested as the origin for a few
290 terrestrial organisms from the Seychelles (Agnarsson & Kuntner 2012 and references therein)
291 and may explain the origin of *Ligia* in the inner islands. Our phylogeographic findings and

292 morphological identifications coupled with the occurrence of *L. dentipes* across Indian Ocean
293 archipelagos such as the Maldives (Taiti 2014) indicate the possibility of a colonization of the
294 Seychelles archipelago after rafting or “island-hopping” throughout the Indian Ocean; however,
295 the poor support for relationships within *Clade DEF* indicates additional research remains
296 needed to determine the origin and *Ligia* from the inner islands of the Seychelles.

297 Additional work is also required to determine whether *Ligia* from the Seychelles
298 represent yet to be described cryptic species. We report levels of genetic divergence between
299 sampled individuals from the Seychelles and other members of their clades that exceed those
300 proposed for species-level divergences (COI K2P > 3%; Hebert et al. 2003) and largely match or
301 exceed those reported from comparisons between valid *Ligia* species (Hurtado et al. 2010;
302 Santamaria et al. 2014; Santamaria et al. 2013; Taiti et al. 2003). *Ligia* from Aldabra exhibit
303 COI K2P divergences of ~27% when compared to other *L. vitiensis* individuals from *Clade ABC*,
304 while individuals from the inner islands show COI K2P divergences of ~15% from *L. dentipes*
305 individuals in *Clade DEF*. Lastly, individuals from Cousine Island exhibit COI K2P divergences
306 of 4.5–5.3% when compared to other localities (Mahé and Silhouette) within the inner islands.
307 Interestingly, this pattern is similar to those previously reported for Seychellois herpetofauna
308 from the inner islands (Rocha et al. 2011; Rocha et al. 2013; Silva et al. 2010). The sum of our
309 findings thus suggest the presence of 2–3 putative cryptic species of *Ligia* in the Seychelles:
310 Aldabra *Ligia* being one, and those of Cousine and Silhouette+Mahé representing one or two
311 putative species.

312 Morphological inspections are needed to determine whether diagnostic characters exist
313 that differentiate between these highly divergent lineages. Uncovering such differences may
314 prove difficult, as previous morphological comparisons of highly divergent *Ligia* lineages within

315 the same species complex suggest lineages to be nearly morphologically identical (Santamaria et
316 al. 2016; Santamaria et al. 2013; Taiti et al. 2003). Our findings also suggest that additional
317 lineages may occur in other *Ligia* populations across the Seychelles, particularly in yet to be
318 sampled outer islands.

319

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331

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Figure 1(on next page)

Map of sampled localities across the Seychelles

Localities are A1-Basin Cabri, Aldabra Atoll, Seychelles; E1-Cousine Island, Seychelles; E2-Silhouette Island, Seychelles; E3-L'Islette, Western Mahé, Seychelles; E4-Anse Parnel, South-eastern Mahé, Seychelles. Colors and labels for each locality correspond with other figures and tables. Detailed information for each locality is presented in Table 1.

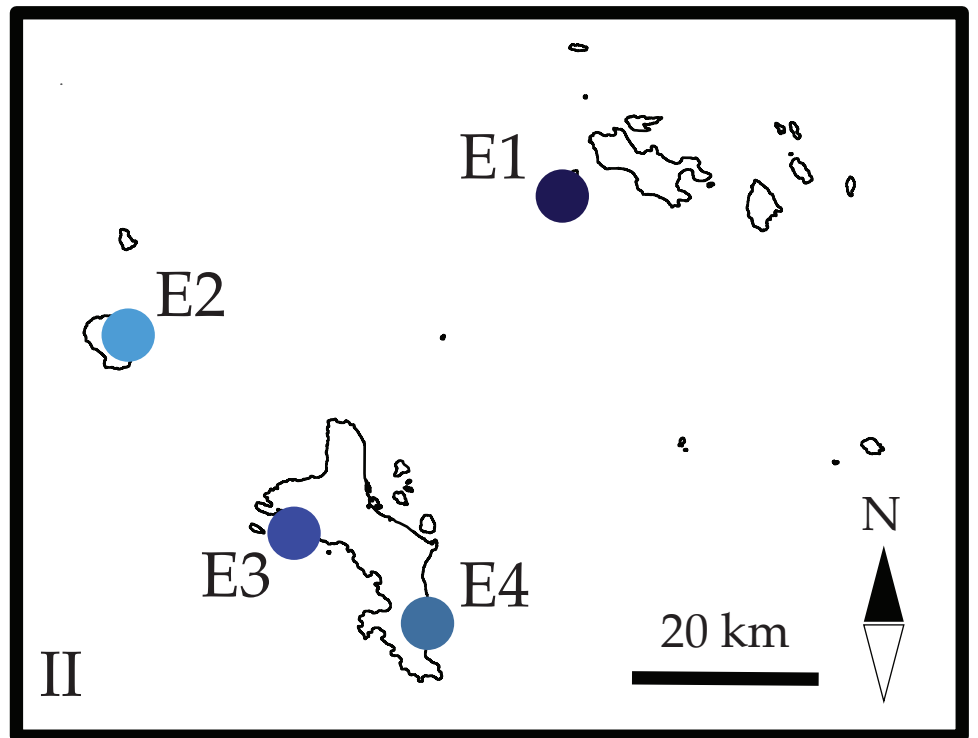
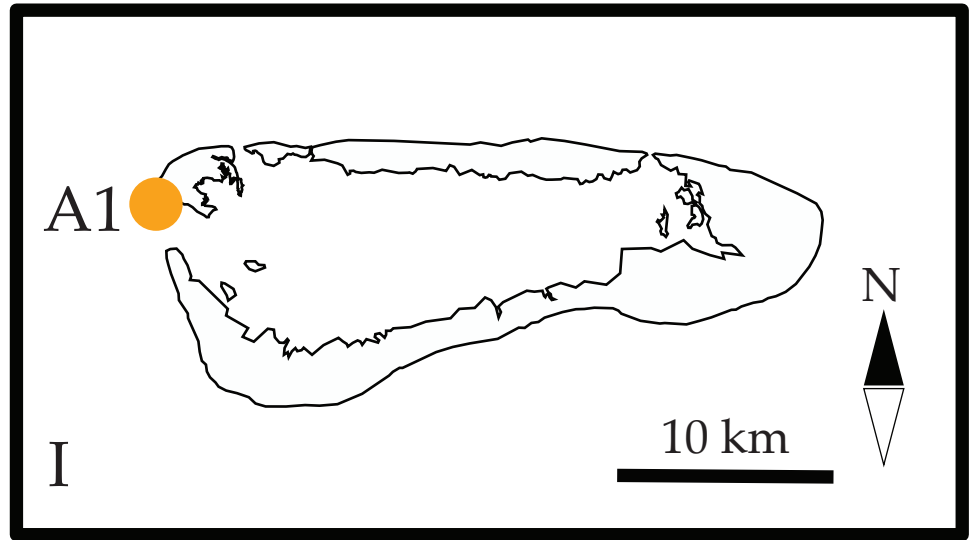
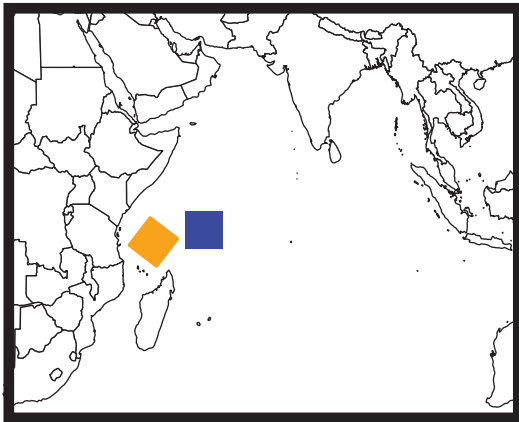


Figure 2(on next page)

Majority rule consensus tree produced by Bayesian Analysis (GTR + Γ , unpartitioned, Phycas) of the concatenated mitochondrial dataset of *Ligia* samples and outgroups included in this study

Numbers by nodes indicate the corresponding range of percent Bootstrap Support (BS; top) for Maximum likelihood; and Posterior Probabilities (PP; bottom) for Bayesian inference methods. Nodes receiving 100% for all methods are denoted with an * while an NS indicates less than 50% node support. Colors represent major lineages as discussed in text.

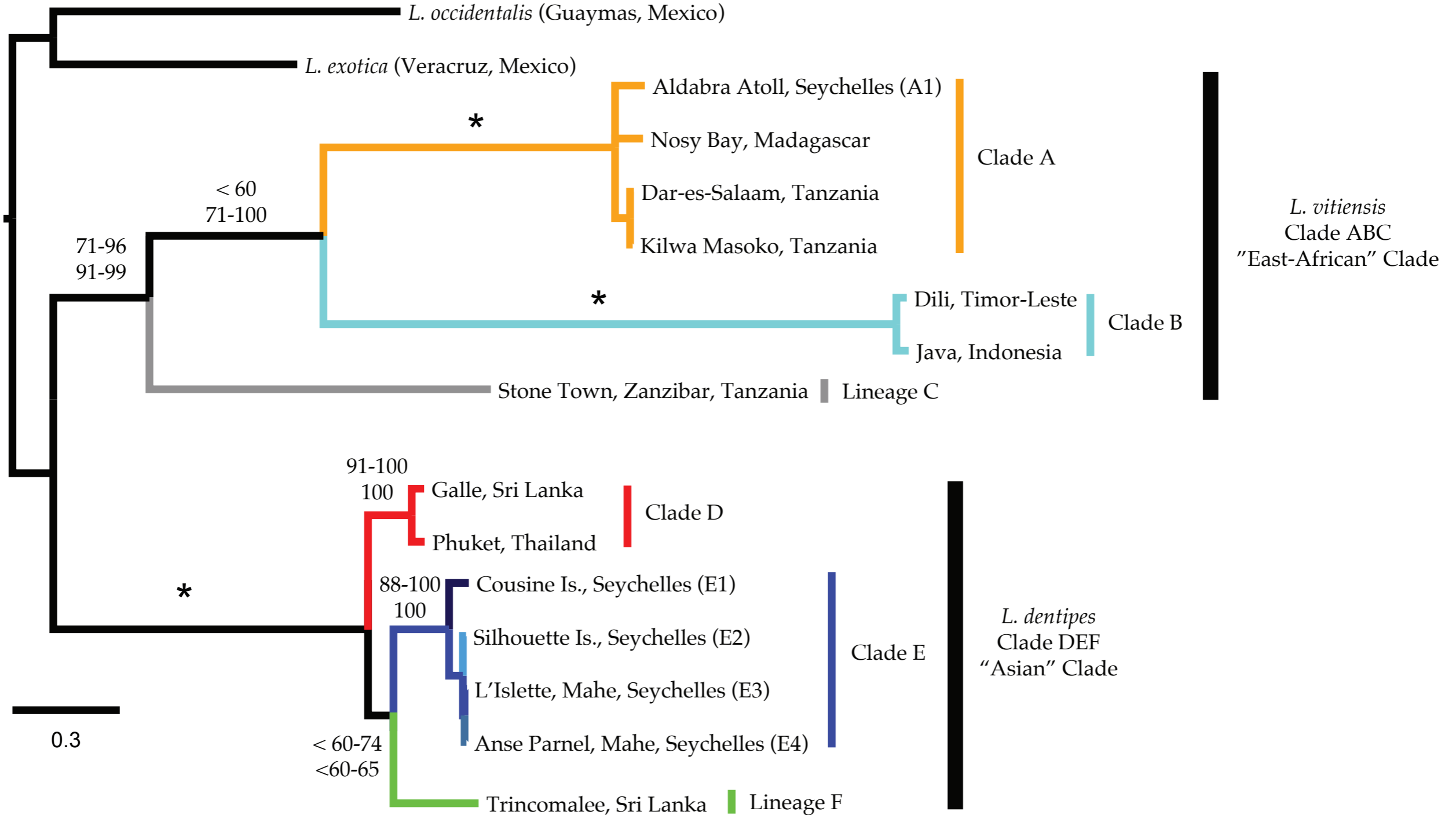


Figure 3(on next page)

COI haplotype networks for Seychellois *Ligia*

Colors and Locality IDs correspond with those use in all other Figures in Figure 1. Empty circles and hash marks represent unsampled (i.e., missing) haplotypes, while the size of circles is proportional to the frequency at which each haplotype was recovered.

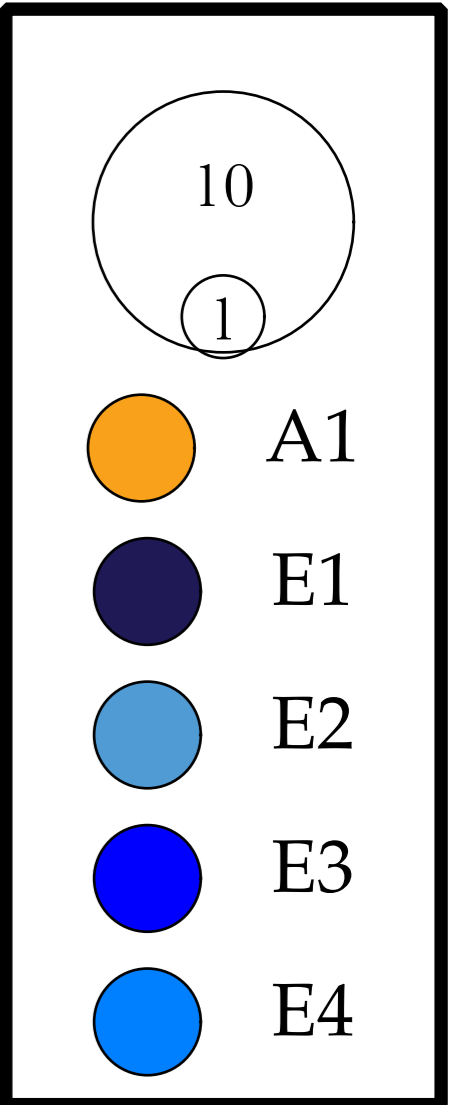
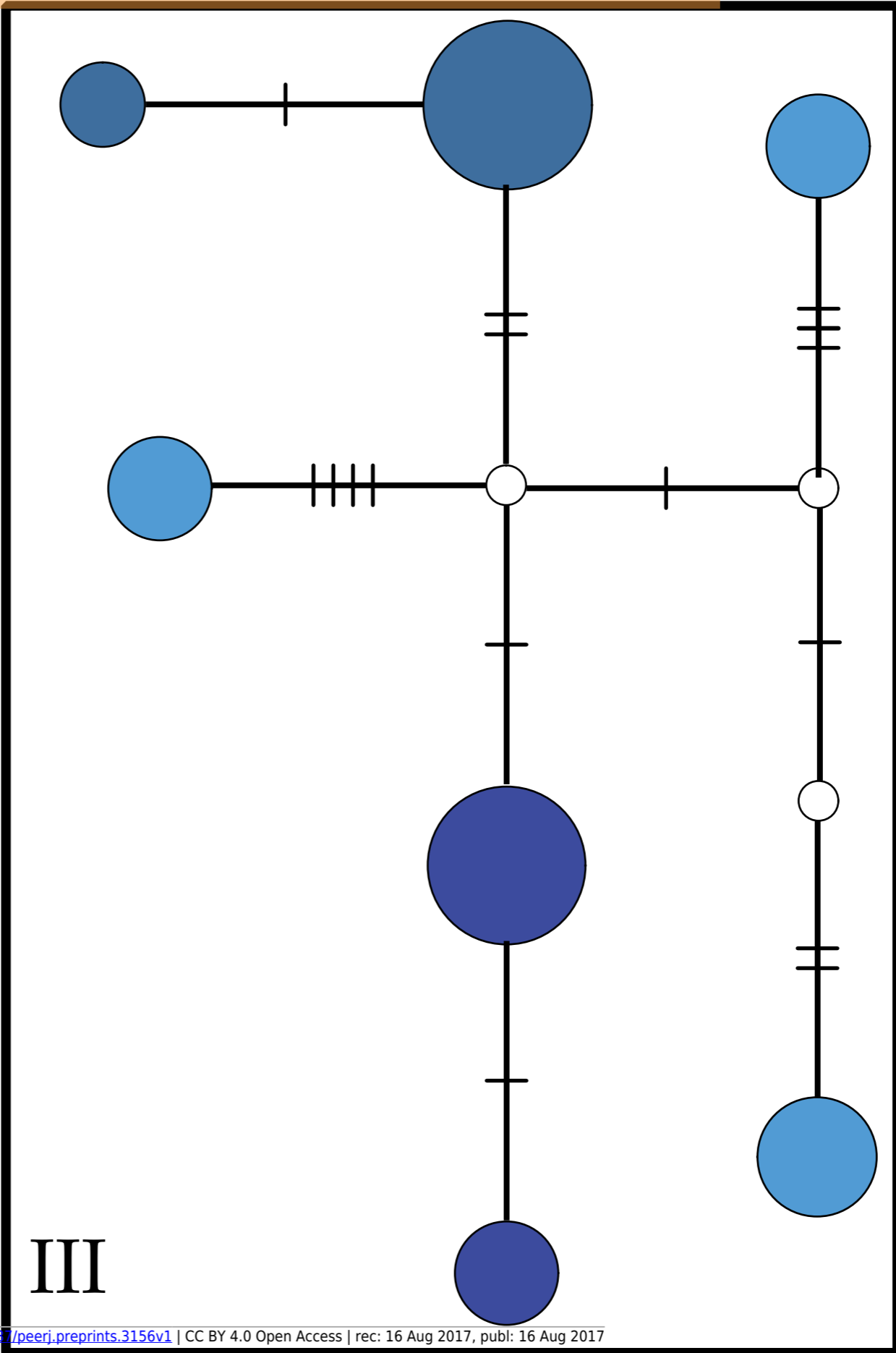
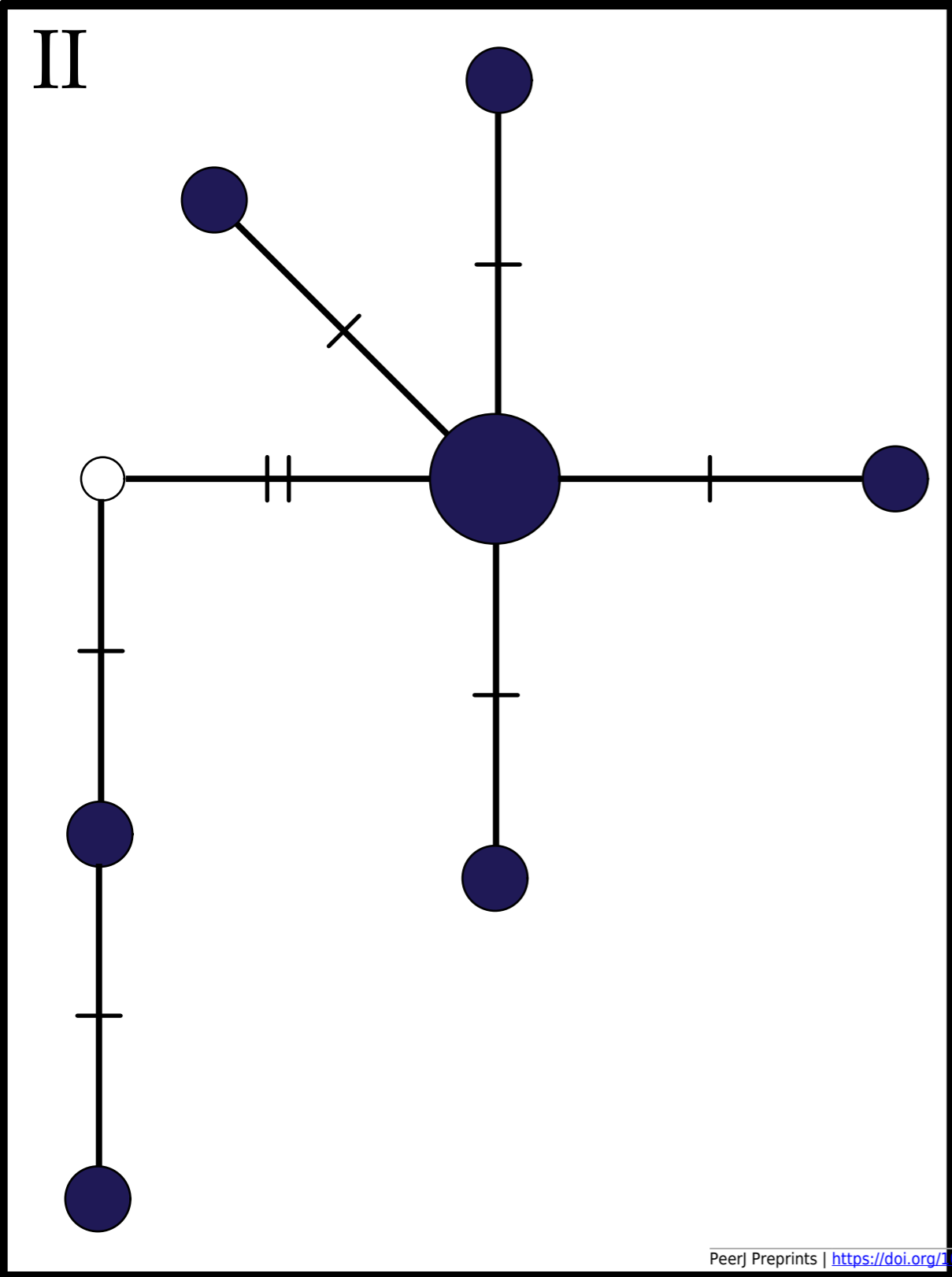
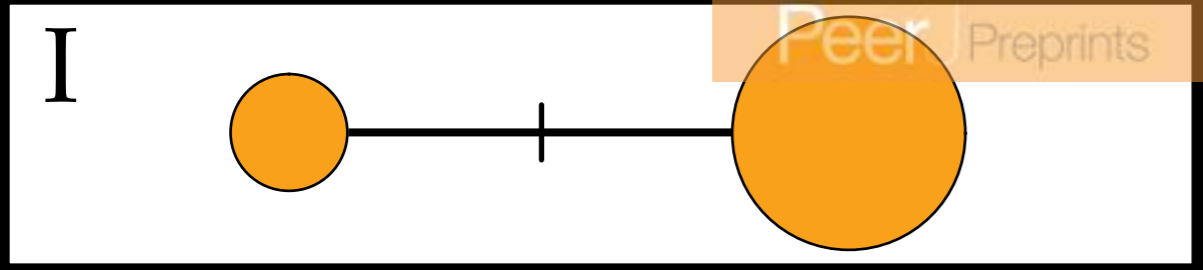


Table 1 (on next page)

Localities included and corresponding GenBank Accession Numbers for all genetic markers used, latitude, and longitude

Map labels correspond with other figures and tables

Species	Locality	Map Label	16S Acc. No.	12S Acc. No.	COI Acc. No.	Cytb Acc. No.	NaK acc no	Latitude	Longitude
<i>L. vitiensis</i>	Basin Cabri, Aldabra Atoll, Seychelles	A1	XXXXX	XXXXX	XXXXX	XXXXX	XXXXX XX	N/A	N/A
<i>L. vitiensis</i>	Nosy Be, Madagascar		AY051342	N/A	AY051323	N/A	N/A	N/A	N/A
<i>L. vitiensis</i>	The Slipway, Dar-Es-Salaam, Tanzania		XXXXX	XXXXX	XXXXX	XXXXX	N/A	6°45'06.9"S	39°16'18.6"E
<i>L. vitiensis</i>	Kilwa Masoko, Tanzania		XXXXX	XXXXX	XXXXX	XXXXX	N/A	8°55'42.7"S	39°31'17.9"E
<i>L. vitiensis</i>	Dili, Timor-Leste		KF546556	KF546581	KF546662	KF546725	N/A	N/A	N/A
<i>L. vitiensis</i>	Parangtritis, Java, Indonesia		KF546554	KF546582	KF546665	KF546727	N/A	8°01'46.4"S	110°20'29.8"E
<i>L. vitiensis</i>	Stone Town, Zanzibar, Tanzania		XXXXX	XXXXX	XXXXX	XXXXX	N/A	6°09'33.8"S	39°11'26.4"E
<i>L. dentipes</i>	Galle, Sri Lanka		XXXXX	XXXXX	XXXXX	XXXXX	N/A	6°01'49.9"N	80°13'03.3"E
<i>L. dentipes</i>	Patong Bay, Phuket, Thailand		KF555801	KF555838	KF555841	KF555754	N/A	7°53'11.0"N	98°17'10.3"E
<i>L. dentipes</i>	Cousine Island, Seychelles	E1	XXXXX	XXXXX	XXXXX	XXXXX	XXXXX	4°20'55.3"S	55°38'41.9"E
<i>L. dentipes</i>	Silhouette Island, Seychelles	E2	XXXXX	XXXXX	XXXXX	XXXXX	XXXXX	4°29'08.4"S	55°15'12.4"E
<i>L. dentipes</i>	L'Islette, W. Mahé, Seychelles	E3	XXXXX	XXXXX	XXXXX	XXXXX	XXXXX	4°39'46.7"S	55°24'35.0"E
<i>L. dentipes</i>	Anse Parnel, S.E. Mahé, Seychelles	E4	XXXXX	XXXXX	XXXXX	XXXXX	XXXXX	4°46'01.1"S	55°31'19.3"E
<i>L. dentipes</i>	Dutch Bay, Trincomalee, Sri Lanka		XXXXX	XXXXX	XXXXX	XXXXX	XXXXX	8°33'52.6"N	81°14'27.8"E
<i>L. occidentalis</i>	Guaymas, Mexico		KF546553	KF546583	KF546666	KF546728	N/A	27°54'44.3"N	110°56'49.6"W
<i>L. exotica</i>	Veracruz, Mexico		KF546552	KF546584	KF546664	KF546726	N/A	19°11'40.2"N	96°07'24.4"W

Table 2 (on next page)

Settings for Maximum Likelihood and Bayesian analyses for the concatenated mitochondrial dataset

Software	Model & Priors ^A	Part Scheme ^B	iterations gen./bootstrap replicates	Sample Freq	Runs/Chain	Burnin	ASDSF ^C	Bayes Factor/ ML Scores (-lnL) ^D	ESS >200 ^E	PSRF ^F
RAXML	GTR + Γ	Unpart	1,000	n/a	n/a	n/a	n/a	-9006.5300	n/a	n/a
RAXML	GTR + Γ	Gene	1,000	n/a	n/a	n/a	n/a	-8865.7562	n/a	n/a
RAXML	GTR + Γ	BP	1,000	n/a	n/a	n/a	n/a	-8334.7701	n/a	n/a
Garli	012010 + Γ + F	Unpart	1,000	n/a	n/a	n/a	n/a	-8986.6239	n/a	n/a
Garli	Mixed Model	Gene	1,000	n/a	n/a	n/a	n/a	-8972.0488	n/a	n/a
Garli	Mixed Model	BP	1,000	n/a	n/a	n/a	n/a	-8212.4919	n/a	n/a
MrBayes	GTR + Γ	Unpart	2 x 10 ⁸	5,000	4	25%	0.000724	-8959.8260	Yes	1
MrBayes	GTR + Γ	Gene	2 x 10 ⁸	5,000	4	25%	0.001157	-8928.4492	Yes	1
MrBayes	GTR + Γ	BP	2 x 10 ⁸	5,000	4	25%	0.001170	-8608.5042	Yes	1
Phycas	GTR + Γ	Unpart	1*10 ⁶	50	n/a	25%	n/a	-8959.6281	Yes	n/a
Phycas	GTR + Γ	Gene	1*10 ⁶	50	n/a	25%	n/a	-8879.1040	Yes	n/a
Phycas	GTR + Γ	BP	1*10 ⁶	50	n/a	25%	n/a	-8324.1730	Yes	n/a

1 ^A: All others default; ^B: BP as indicated by PartitionFinder (1: 16S+12S+Cytb 2nd codons; 2: COI 1st codons+Cytb 1st codons; 3: COI
2 2nd codons; 4: Cytb 3rd codons + COI 3rd codons); ^C: Average Standard Deviation of Split Frequencies; ^D: Estimated in Tracer v.1.5.; ^E:
3 Effective Sample Size; ^F: Potential Scale Reduction Factor for all parameters.

4

Table 3 (on next page)

Estimates of evolutionary divergence, as measured by Kimura 2-parameter distances, for main *Ligia* lineages from the study area and outgroups

When applicable minimum and maximum (top values) as well as average divergences (values in parentheses) are provided

	<i>Clade A</i>	<i>Clade B</i>	<i>Clade C</i>	<i>Clade D</i>	<i>Clade E</i>	<i>Clade F</i>	<i>L. exotica</i>	<i>L. occidentalis</i>
<i>Clade A</i>	7.1–9.1% (4.3%)							
<i>Clade B</i>	22.9–24.7% (24.0%)	2.50%						
<i>Clade C</i>	25.8–28.1% (26.8%)	25.9–26.6% (26.2%)	N/A					
<i>Clade D</i>	25.3–27.1% (26.3%)	22.0–23.8% (22.8%)	25.9–27.2% (26.6%)	4.70%				
<i>Clade E</i>	24.3–27.6% (26.3%)	21.9–23.7% (22.1%)	24.7–25.4% (25.2%)	12.4–14.8% (13.7%)	0.5–4.7% (2.9%)			
<i>Clade F</i>	26.3–27.8% (27.3%)	23.3–23.75% (23.5%)	27.7%	13.3–14.2% (13.8%)	12.1–12.7% (12.3%)	N/A		
<i>L. exotica</i>	22.6–25.4% (23.4%)	25.7–25.7% (25.7%)	25.7%	25.9–28.1% (27.0%)	24.9–26.9% (25.9%)	26.3%	N/A	
<i>L. occidentalis</i>	27.5–28.3% (28.0%)	23.8–24.9% (24.3%)	24.6%	24.9%	19.0–20.9% (19.8%)	20.8%	25.6%	N/A

1
2

Table 4(on next page)

Estimates of evolutionary divergence, as measured by Kimura 2-parameter distances, for *Ligia* localities from Seychelles inner island localities

	E1	E2	E3	E4
E1	-----			
E2	4.9%	-----		
E3	5.1%	0.8%	-----	
E4	5.2%	0.9%	0.5%	-----

1