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Clinical significance of CD44s, CD44v3 and CD44v6 in breast cancer

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Abstract

Objectives: To evaluate levels of CD44 standard variant (CD44s), CD44 variant exon 3 (CD44v3) and CD44 variant exon 6 (CD44v6) protein in breast cancer tissue, and investigate their relationships with clinicopathological characteristics of the disease.

Methods: Immunohistochemistry for CD44s, CD44v3 and CD44v6 was retrospectively performed on formalin-fixed paraffin wax-embedded breast cancer tissue samples.

Results: Tumour tissue samples from 60 patients with breast cancer were included. There was a significant relationship between CD44s positivity and tumour diameter and lymph node involvement. CD44v6 positivity was significantly associated with tumour–node–metastasis (TNM) stage and lymph node involvement. There were significant negative correlations between CD44s immunopositivity, tumour diameter and TNM stage, and significant positive correlations between CD44v6 immunopositivity, tumour diameter and TNM stage.

Conclusions: CD44s and CD44v6 appear to play opposing roles in the development of breast cancer, but their precise functions and mechanisms of action remain unclear.

Keywords

Breast cancer, adhesion molecule, CD44, immunohistochemistry, clinical significance

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Introduction

Breast cancer is the most prevalent malignancy and the second most common cause of cancer-related mortality in women worldwide.¹ Improvement in the clinical management of breast cancer is dependent on having a greater understanding of known prognostic factors and the identification of indicators that may help to assess tumour behaviour.

The multifunctional and multistructural transmembrane glycoprotein CD44 was originally characterized as a hyaluronan (HA) receptor and lymphocyte homing receptor.² CD44 plays a pivotal role in the prognosis of malignancies including breast cancer.^{3–5} CD44 exists as both the standard form (CD44s) and a number of isoforms generated by alternative splicing of variant exons (CD44v).⁶

The standard form, CD44s, is related to the proliferation, infiltration, angiogenesis, metastasis and prognosis of breast cancer.^{6,7} CD44s expression is significantly higher in gastric cancer than in normal gastric tissue, and this increased expression is associated with adenocarcinoma tumourigenesis, metastasis and clinically aggressive behaviour.⁸ In contrast, loss of CD44s has been found to correlate with lymph node metastasis and unfavourable outcome in patients with breast carcinoma.⁹ The role of CD44s in cancer remains to be fully elucidated.

The CD44 exon 3 variant (CD44v3) plays a role in breast cancer development.¹⁰ CD44v3 promotes tumourigenesis in noma of the head and neck, and might be an effective tumour marker for targeted therapy.^{10,11} In addition, CD44v3 is associated with relapse and reduced overall survival in people with vulvar cancer.¹² CD44 exon 6 variant (CD44v6) is responsible for the regulation of tumour invasion, progression and metastasis in rat carcinoma cells.¹³ CD44v6 is also correlated with prognosis in breast,¹⁴ gastric,¹⁵ colorectal,¹⁶ ovarian,¹⁷ bladder¹⁸ and liver cancer.¹⁹

The mechanisms by which CD44 isoforms exert their effects in cancer are unclear. The present study used immunohistochemistry to evaluate CD44s, CD44v3 and CD44v6 levels in breast cancer tissue, and investigate their relationships with clinicopathological characteristics.

Patients and methods

Study population

The study included samples of tumour tissue from female patients with breast cancer who underwent surgical resection at Department of Radiation Oncology, The First People's Hospital of Xuzhou, Xuzhou, China, between January 2011 and June 2012. Patient diagnoses were confirmed by two independent pathologists (G. L. and Y. X.), and tumours were classified according to the tumour–node–metastasis (TNM) system of the International Union against Cancer (1988).²⁰ No patient received any radiotherapy or chemotherapy prior to enrolment. The Medical Ethics Committees of Soochow University and Nanjing Medical University approved the study and all patients provided written informed consent prior to enrolment.

Immunohistochemistry

Formalin-fixed, paraffin wax-embedded tumour tissue and adjacent normal mucosa samples were sliced into 4 µm-thick sections. Immunohistochemistry for CD44s, CD44v3 and CD44v6 was performed using ultrasensitive immunohistochemical kits (Maxim Co., Ltd, Fuzhou, China), according to the manufacturer's instructions.

Immunoreactivity for CD44 proteins was observed as brown, granular staining on the cytoplasmic membrane of both cancerous and stromal cells. Protein staining was evaluated in arbitrarily selected visual fields. Staining was classified as: negative (–), 0–<10% positive cells; weak positive

(+), 10–25% positive cells; positive (++) , 26–50% positive cells; or strong positive (+++), $\geq 51\%$ positive cells. For the purposes of this study, all positive staining levels (+, ++ and +++) were defined as positive.

Statistical analyses

Data were presented as *n*. Analysis of variance, Wilcoxon's rank sum test and Spearman's rank correlation analysis were used to analyse the relationships between clinicopathological features and CD44s, CD44v3 and CD44v6 immunopositivity. Statistical analyses were performed using SPSS[®] version 11.0 (SPSS Inc., Chicago, IL, USA) for Windows[®]. *P*-values < 0.05 were considered statistically significant.

Results

The study included tissue samples from 60 female patients with breast cancer (42 infiltrating ductal carcinomas, seven intraductal carcinomas, six mucinous adenocarcinomas, one medullary carcinoma, three papillary adenocarcinomas and one lobular infiltrating carcinoma). Patients were aged 32–81 years (median age, 51 years).

Positive staining rates of CD44s, CD44v3 and CD44v6 in breast cancer tissue samples were 78.3%, 75.0% and 78.3%, respectively. The cellular location of CD44 proteins varied, with CD44s found both on the cytoplasmic membrane and in the cytoplasm, and CD44v3 and CD44v6 mainly found in the cytoplasm and only occasionally on the cytoplasmic membrane (Figure 1).

Relationships between CD44s, CD44v6 and CD44v3 immunopositivity in breast cancer tissue samples and clinicopathological characteristics are shown in Table 1. Using analysis of variance, there was a significant relationship between CD44s positivity and tumour diameter ($P = 0.020$) and

lymph node involvement ($P = 0.001$). CD44v6 was significantly associated with tumour stage ($P = 0.010$). There were no other statistically significant relationships between CD44s, CD44v6 or CD44v3 and any other clinicopathological characteristic (Table 1). Analyses using Wilcoxon's rank sum test indicated significant relationships between lymph node involvement status and immunopositivity for CD44s ($P = 0.001$) and CD44v6 ($P = 0.009$), but not CD44v3.

There were significant negative correlations between CD44s immunopositivity in breast cancer tissue and tumour diameter ($r = -0.338$, $P < 0.01$) and stage ($r = -0.298$; $P < 0.05$), and significant positive correlations between CD44v6 immunopositivity and tumour diameter ($r = 0.257$; $P < 0.05$) and stage ($r = 0.383$; $P < 0.01$). There were no significant correlations between CD44v3 and tumour diameter or stage.

Discussion

The molecular and cellular processes underlying breast cancer development are poorly understood, and there are few definitive biological markers. The present study was designed to determine the value of CD44 molecules as tumour markers for breast cancer. We found that CD44s was located to the cytoplasmic membrane, whereas CD44v3 and v6 were found in the cytoplasm. This suggests that CD44s may be the more useful of these molecules as a therapeutic target.

The present study observed that CD44s and CD44v6 had identical rates of immunopositivity, but their cellular location and biological function were different. This identical rate of positive staining is likely to be a coincidence, but larger numbers of samples are required to confirm this.

Both CD44s and CD44v6 positivity were associated with tumour stage and diameter in the present study, indicating that these molecules may be useful tumour markers.

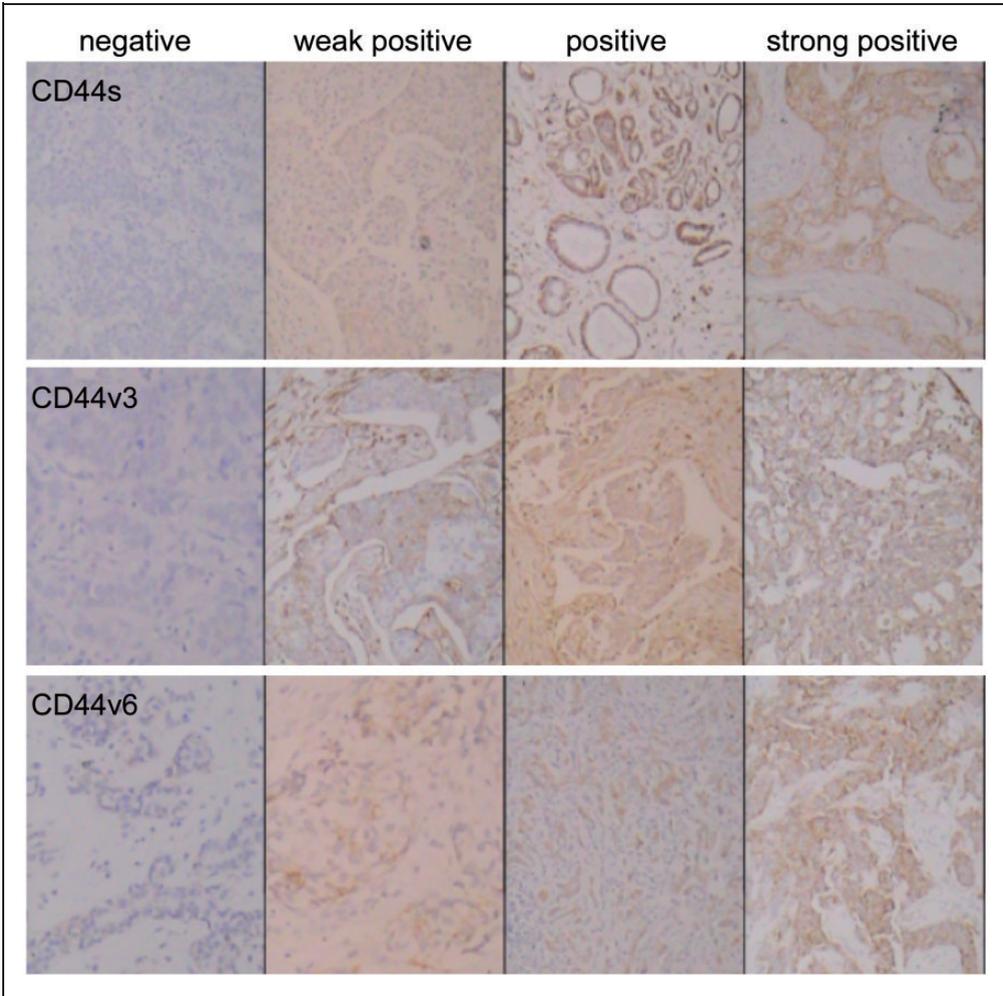


Figure 1. Representative light photomicrographs showing immunohistochemical staining of breast cancer tissue samples for CD44s (standard variant), CD44 variant exon 3 (CD44v3) and CD44 variant exon 6 (CD44v6). Original magnification $\times 200$. The colour version of this figure is available online at www.sagepub.com.

In addition, CD44s and CD44v6 were both correlated with lymph node involvement. In contrast to our findings, others have found significant associations between CD44v3 and tumour development or prognosis.^{10–12} This inconsistency may be due to heterogeneity within the tumour itself, different standards for evaluating staining, or variations in antibody specificity or sensitivity.

Interestingly, tumour diameter and stage were negatively correlated with CD44s but positively correlated with CD44v6 in the present study, suggesting that these molecules have opposing roles in breast cancer development. Patients with CD44v6-positive/CD44s-negative tumours may have an increased risk of tumour invasion and metastasis. Others have found that normal ductal epithelial cells of mammary glands

Table 1. Relationship between immunopositivity for standard and variant forms of CD44 in tumour tissue samples and clinicopathological characteristics of breast cancer ($n = 60$).

Characteristic	CD44s				CD44v3				CD44v6			
	-	+	++	+++	-	+	++	+++	-	+	++	+++
Tumour diameter, cm	$P = 0.020$				NS				NS			
<2	0	3	6	7	3	10	2	1	2	8	6	0
2–5	10	8	8	13	11	7	13	8	10	6	11	12
>5	3	1	1	0	1	0	2	2	1	2	2	0
Lymph node involvement	$P = 0.001$				NS				NS			
No	2	4	11	14	8	11	10	2	8	12	3	3
Yes	11	8	4	6	7	6	7	9	5	14	1	4
TNM stage	NS				NS				$P = 0.010$			
I	0	2	5	5	3	6	2	1	2	8	2	0
II	6	8	8	11	8	10	9	6	9	6	13	5
III	4	1	2	3	4	0	4	2	2	2	3	3
IV	3	1	0	1	0	1	2	2	0	0	1	4
Tumour type	NS				NS				NS			
Infiltrating ductal carcinoma	9	9	10	14	9	10	11	12	10	18	9	5
Intraductal carcinoma	2	1	2	2	1	2	2	1	2	2	1	2
Mucinous adenocarcinoma	2	0	2	2	2	2	0	2	2	1	1	2
Medullary carcinoma	0	1	0	0	1	0	0	0	0	0	1	0
Papillary adenocarcinoma	0	2	1	0	2	1	0	0	1	0	1	1
Lobular infiltrating carcinoma	0	0	1	0	0	1	0	0	0	0	0	1

Data presented as n .

Staining: -, 0–<10% positive cells; +, 10–25% positive cells; ++, 26–50% positive cells; +++, ≥51% positive cells.

CD44s, CD44 standard variant; CD44v3, CD44 variant exon 3; CD44v6, CD44 variant exon 6; NS, not statistically significant ($P \geq 0.05$, analysis of variance); TNM, tumour–node–metastasis.²⁰

and proliferative mammary cells do not express CD44 variants.²¹ In addition, CD44v6-positive breast cancers were more likely to have lymph node involvement than CD44v6-negative tumours.²¹ It is possible that transcriptional alterations in CD44 result in functional and structural abnormalities, impacting on recognition, adhesion and information transfer among cells, ultimately altering biological behaviour. Moreover, these adhesion factors could be important molecular biological markers that predict the invasion and metastatic prognosis of breast cancer. Members of the CD44 family were found to be biological markers in colorectal cancer.²²

It is well known that CD44 is the major HA receptor, and that HA-bound CD44 participates in tumour progression, metastasis and proliferation.⁶ CD44 has been demonstrated to play a role in promoting cell-to-extracellular matrix interaction,²³ binding to HA,^{24,25} binding to matrix metalloproteinase,²⁴ and erbB receptor tyrosine kinases.²⁴

The present study has several limitations, notably the small sample size. In particular, only five patients had stage IV tumours. Future studies should include at least 100 participants in order to meet statistical requirements as well as nontumour control tissues. A further limitation was that reverse transcription–polymerase chain reaction

was not performed. This would allow direct assessment of gene expression and enable the investigation of its correlation with protein levels. Finally, we included a limited number of clinicopathological variables. Data regarding histological grade, proliferation index, and oestrogen receptor/HER2 status should be included in future studies.

In conclusion, CD44s and CD44v6 appear to play opposing roles in the development of breast cancer, but their precise function and mechanisms of action remain unclear.

Declaration of conflicting interest

The authors declare that there are no conflicts of interest.

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References

1. Ferlay J, Shin HR, Bray F, et al. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer* 2010; 127: 2893–2917.
2. Naor D, Sionov RV and Ish-Shalom D. CD44: structure, function, and association with the malignant process. *Adv Cancer Res* 1997; 71: 241–319.
3. Perez A, Neskey DM, Wen J, et al. CD44 interacts with EGFR and promotes head and neck squamous cell carcinoma initiation and progression. *Oral Oncol* 2013; 49: 306–313.
4. Franzmann EJ, Reategui EP, Pedrosa F, et al. Soluble CD44 is a potential marker for the early detection of head and neck cancer. *Cancer Epidemiol Biomarkers Prev* 2007; 16: 1348–1355.
5. Watanabe O, Kinoshita J, Shimizu T, et al. Expression of a CD44 variant and VEGF-C and the implications for lymphatic metastasis and long-term prognosis of human breast cancer. *J Exp Clin Cancer Res* 2005; 24: 75–82.
6. Anand MT and Kumar S. CD44: A key player in breast cancer. *Indian J Cancer* 2014; 51: 247–250.
7. Afify A, Purnell P and Nguyen L. Role of CD44s and CD44v6 on human breast cancer cell adhesion, migration, and invasion. *Exp Mol Pathol* 2009; 86: 95–100.
8. Liu YJ, Yan PS, Li J, et al. Expression and significance of CD44s, CD44v6, and nm23 mRNA in human cancer. *World J Gastroenterol* 2005; 11: 6601–6606.
9. Gong Y, Sun X, Huo L, et al. Expression of cell adhesion molecules, CD44s and E-cadherin, and microvessel density in invasive micropapillary carcinoma of the breast. *Histopathology* 2005; 46: 24–30.
10. Ryś J, Kruzak A, Lackowska B, et al. The role of CD44v3 expression in female breast carcinomas. *Pol J Pathol* 2003; 54: 243–247.
11. Franzmann EJ, Weed DT, Civantos FJ, et al. A novel CD44 v3 isoform is involved in head and neck squamous cell carcinoma progression. *Otolaryngol Head Neck Surg* 2001; 124: 426–432.
12. Tempfer C, Sliutz G, Haeusler G, et al. CD44v3 and v6 variant isoform expression correlates with poor prognosis in early-stage vulvar cancer. *Br J Cancer* 1998; 78: 1091–1094.
13. Gunthert U, Hofmann M, Rudy W, et al. A new variant of glycoprotein CD44 confers metastatic potential to rat carcinoma cells. *Cell* 1991; 65: 13–24.
14. Kaufmann M, Heider KH, Sinn HP, et al. CD44 variant exon epitopes in primary breast cancer and length of survival. *Lancet* 1995; 345: 615–619.
15. Mayer B, Jauch KW, Gunthert U, et al. De-novo expression of CD44 and survival in gastric cancer. *Lancet* 1993; 342: 1019–1022.
16. Ropponen KM, Eskelinen MJ, Lipponen PK, et al. Expression of CD44 and variant proteins in human colorectal cancer and its relevance for prognosis. *Scand J Gastroenterol* 1998; 33: 301–309.
17. Shi J, Zhou Z, Di W, et al. Correlation of CD44v6 expression with ovarian cancer progression and recurrence. *BMC Cancer* 2013; 13: 182.
18. Omran OM and Ata HS. CD44s and CD44v6 in diagnosis and prognosis of

- human bladder cancer. *Ultrastruct Pathol* 2012; 36: 145–152.
19. Coradini D, Zorzet S, Rossin R, et al. Inhibition of hepatocellular carcinomas in vitro and hepatic metastases in vivo in mice by the histone deacetylase inhibitor HA-But. *Clin Cancer Res* 2004; 10: 4822–4830.
 20. American Joint Committee on Cancer. *AJCC Cancer Staging Manual*, 7th edn. Chapter 32. Berlin, Germany: Springer, 2010.
 21. Kopp R, Classen S, Wolf H, et al. Predictive relevance of soluble CD44v6 serum levels for the responsiveness to second line hormone- or chemotherapy in patients with metastatic breast cancer. *Anticancer Res* 2001; 21: 2995–3000.
 22. Bendardaf R, Algars A, Elzagheid A, et al. Comparison of CD44 expression in primary tumours and metastases of colorectal cancer. *Oncol Rep* 2006; 16: 741–746.
 23. Arch R, Wirth K, Hofmann M, et al. Participation in normal immune responses of a metastasis-inducing splice variant of CD44. *Science* 1992; 257: 682–685.
 24. Lokeshwar VB, Iida N and Bourguignon LY. The cell adhesion molecule, GP116, is a new CD44 variant (ex14/v10) involved in hyaluronic acid binding and endothelial cell proliferation. *J Biol Chem* 1996; 271: 23853–23864.
 25. Ponta H, Sherman L and Herrlich PA. CD44: from adhesion molecules to signalling regulators. *Nat Rev Mol Cell Biol* 2003; 4: 33–45.